Journal of Applied Pharmaceutical Science Vol. 15(07), pp 027-036, July, 2025 Available online at http://www.japsonline.com DOI: 10.7324/JAPS.2025.234398 ISSN 2231-3354



Review on phenolic constituents and pharmacological activities of genus *Ononis*

Aya Chouman¹^{*}, Abdalla El-lakany^{1,2}, Maha Aboul-ela^{1,2}, Mohamad Ali Hijazi¹

¹Department of Pharmaceutical Sciences, Faculty of Pharmacy, Beirut Arab University, Beirut, Lebanon. ²Department of Pharmacognosy, Faculty of Pharmacy, University of Alexandria, Alexandria, Egypt.

ARTICLE HISTORY

Received on: 28/12/2024 Accepted on: 16/04/2025 Available Online: 05/06/2025

Key words: Ononis, phytochemical constituents, flavonoids, pharmacological activities, structure-activity relationship.

ABSTRACT

Plants of genus *Ononis* belong to family Fabaceae and are distributed in Europe, Atlantic Islands, West Asia, and North Africa and more than 75 species in the genus have been identified worldwide. Previous phytochemical studies showed a diversity of bioactive constituents, specifically flavonoids and isoflavonoids, that have been responsible for the different actions exerted by plants of this genus. This study represents a comprehensive review of all data present in literature related to the phenolic constituents isolated from *Ononis* plants and their pharmacological activities. The study revealed also the essential structure-activity relationship of *Ononis* compounds and briefly highlighted some toxicity studies. Data was collected by searching journals, books, periodicals, and databases. The results demonstrated a wide variety of flavonoids and isoflavonoids, isocoumarins, and resorcinols reported from different *Ononis* species. Flavonoidal compounds were the main constituents responsible for the various pharmacological activities such as antioxidant, anti-inflammatory, antimicrobial, cytotoxic, enzyme inhibitory, and antidiabetic effects. The reason behind these activities was mainly the degree and position of hydroxylation and methoxylation on the flavonoids' rings. The findings of this study suggest *Ononis* plants and/or their components are promising candidates for treatment of wide range of ailments as a preventive measure or in combination with conventional therapies. Further studies would be required to describe the detailed mechanistic pathways for promising compounds and their clinical applications.

INTRODUCTION

Papilionaceae, Leguminosae, or Fabaceae, also named the pea, legume, or bean family, belongs to the Fabales order. With 19,400 species and 740 genera, Fabaceae is the thirdlargest plant family after Orchidaceae (orchid family) and Asteraceae (aster family), and it is one of the world's estimated 12 flowering plants [1]. The Fabaceae family has over 490 species, making it the second-largest family of medicinal plants, the majority of which have been utilized as traditional medicines, making it one of the most interesting families in research [2]. The family features aquatic plants, woody lianas, climbing annuals, shrubs, subshrubs, trees, and herbs. Stems are twining, erect, or climbing. Leaves are rarely simple and mostly compound [3]. Flowers are rarely zygomorphic, actinomorphic, or unisexual. They are bisexual, mainly in corymbs, heads, racemes, panicles, or spikes [4].

The genus *Ononis*, commonly named restharrow, is part of the Fabaceae family. It is widely spread in West Asia, the Atlantic Islands, North Africa, and Europe, and more than 75 species in the genus have been identified worldwide, particularly near the coasts of the Mediterranean Sea [5]. The scientific interest in the genus *Ononis* has increased recently as it contains several active components that have therapeutic benefits and are cost-effective. Mainly, phenolic compounds, particularly flavonoids, have been extracted. Mainly, phenolic compounds, particularly flavonoids, have been extracted. Genus *Ononis* exhibits diverse pharmacological activities, including diuretic, antitussive, anti-inflammatory, analgesic, antioxidant [6,7], antimicrobial [6], and anticancer effects [8]. Additional uses include the treatment of urinary tract infections, rheumatism, wound healing, eczema, and other skin diseases

^{*}Corresponding Author

Aya Chouman, Department of Pharmaceutical Sciences, Faculty of Pharmacy, Beirut Arab University, Beirut, Lebanon. E-mail: a.chouman @ bau.edu.lb

^{© 2025} Aya Chouman *et al.* This is an open access article distributed under the terms of the Creative Commons Attribution 4.0 International License (https:// creativecommons.org/licenses/by/4.0/).

[9]. In this work, we attempt to summarize and review the phenolic constituents of plants belonging to the genus *Ononis*, their pharmacological effects, structure–activity relationship, and reported mechanisms of action.

METHODS

The keywords *Ononis*, phytochemical constituents, pharmacological activities, and structure-activity relationship were searched. The search was obtained from journals and books in databases, such as Scopus, ScienceDirect, Web of Science, PubMed, Google Scholar, and The Cochrane Library, from 1990 to 2024.

RESULTS AND DISCUSSION

Phytochemical profile

Ononis species are the source of several pharmacologically important flavonoids and isoflavonoids. Flavonoids are classified as secondary metabolites. Their primary structural components are phenolic or polyphenolic groups found at different positions of a benzopyrone ring. They are divided into several categories based on the oxidation of the carbon ring, their degree of unsaturation, and their chemical structure. The several subclasses of flavonoids comprise isoflavonoids, flavanones, anthoxanthins (flavanol and flavanone), flavans , flavanones, flavanones, and anthocyanidins [10].

Flavonoids and isoflavonoids have been described from *Ononis* species so far, including the isoflavones and the less prevalent isoflavonoids, other types of constituents, including resorcinol derivatives [11], isocoumarins [12], phenolic lactones [13], and terpenoids [7,14], have been isolated from *Ononis* species. According to most data in the literature, flavonoids are very abundant in herbs, stems, fruits, flowers, vegetables, and seeds. It is highly suggested they are found in chloroplasts. Table 1 summarizes the phenolic profile of different *Ononis* species.

Pharmacological effects of some Ononis plants

Antioxidant and anti-inflammatory effects

Crude extracts (ethyl acetate, n-butanol, and petroleum ether) of *Ononis mitissima* plant showed moderate antioxidant activities. In addition, at 500 µg/ml, these extracts exhibited medium anti-inflammatory activity, with inhibition percentages of 33%, 11%, and 22% in comparison to diclofenac (86%) for ethyl acetate, n-butanol, and petroleum ether extracts, respectively [15]. Another study showed that ethyl acetate and methanolic root extracts of *O. spinosa* reduced inflammation by 40.4% and 35.4%, respectively [5]. Moreover, the leaf extract of Tunisian *Ononis natrix* demonstrated a high reducing power (ED50 = 100 µg/ml), with a low IC₅₀ value (29 µg/ml) corresponding to an increased 2,2-diphenyl-1-picrylhydrazyl scavenging ability. With 60.94 mg of GAE/g DW, it also demonstrated a high level of overall antioxidant activity. [6].

Antimicrobial effects

Many studies reported on the antimicrobial effect of the Ononis species. Ononis natrix extract demonstrated efficacy against strains of gram-positive bacteria, particularly against *Staphylococcus aureus ATC 25923* and *Staphylococcus epidermidis* strains [16]. Similarly, *O. spinosa* stopped *Bacillus subtilis, S. aureus, Pseudomonas aeruginosa, and Escherichia coli* growth. Moreover, potential antimicrobial activity against *Candida albicans, MRSA, P. aeruginosa, Bacillus cereus, E. coli, Salmonella typhimurium,* and *Aspergillus niger* has been reported from *Oxalis hirta* and *Oxalis sicula* ethanolic extracts [17]. The same study showed that chloroform extract of *Oxalis arvensis* leaves had significant inhibitory activity against *E. coli* and *C. albicans*, showing respective MIC values of 12.75 µg/ml and 51 µg/ml. [17].

Cytotoxic effect

By stopping the growth of MDA MB-231 breast cancer cells, Jordanian *O. natrix* revealed a potential antitumor effect with an IC₅₀ of 29–41 µg/ml compared to that of tamoxifen (IC₅₀ 11 µg/ml) [18]. Turkish *O. natrix* extracts, on the other hand, exhibited an apoptotic effect against PC3 cancerous cell line at a very small concentration (0.1 µg/ml) [16].

Enzyme inhibitory effect

The dichloromethane extract of *O. spinosa* demonstrated a concentration-dependent suppression of TNF- α and IL-8 production from lipopolysaccharide-stimulated human neutrophils [19]. In addition, the ethyl acetate extract revealed high inhibitory activity of cholinesterase on both BChE (0.93 mg GALAE/g extract) and AChE (1.46 mg GALAE/g extract), unlike the water extract, which was ineffective on BChE. Conversely, the ethyl acetate extract had no inhibitory activity of tyrosinase, but the water extract had significant activity (52.81 mg KAE/g extract). Regarding glucosidase and amylase inhibition, the most effective extracts were the ethyl acetate (0.74 mmol ACAE/g and 17.52 mmol ACAE/g) and methanol extracts (0.59 mmol ACAE/g and 19.94 mmol ACAE/g) [16].

Antidiabetic effect

Preclinical studies on *O. natrix* decoction have demonstrated its effectiveness in lowering blood glucose levels in rats [20]. Ononis extracts can improve glucose and starch tolerance by activating the glucose transporter type-4 (Glut-4) receptor in experimental rats [21]. Moreover, aqueous extracts of *O. natrix* had the ability to restore the islets of Langerhans suffering from alloxan-induced tissue damage in albino Swiss mice [22].

Structure–activity relationship of phenolic constituents from *Ononis*

The medicinal importance of *Ononis* plants is explained by their phenolic contents, mainly flavonoids. These compounds account for the anti-inflammatory, antioxidant, and anticancer characteristics of *Ononis* plants [23].

Anticancer

Formononetin, one of the most known isoflavones in the *Ononis* species, revealed anticancer activity against various types of cancers such as colon, breast, prostate, nasopharyngeal, Table 1. Phytochemistry of Ononis species and their chemical structures.

Flavonoids											
R4 I											
R_3 R_5											
R_2											
$\left[\begin{array}{c} & \\ \end{array} \right]$											
R											
Compound	R	R,	R,		R,	R,	R,	Plant Species	References		
Gardenin B	OCH,	OCH ₃	OCH ₃	H	4 H	OCH ₃	H				
Xanthomicrol	OCH,	OCH,	OCH,	Н	Н	OH	Н	Ononis natrix	[24]		
Hemopexin	OCH ₃	OH	OCH ₃	Н	Н	OCH ₃	Н	(actial parts)			
5,4'-diOH-6,7,8,3'-tetraOMe	OCH ₃	OCH ₃	OCH ₃	Н	OCH_3	OH	Н				
Sideritoflavone	OCH ₃	OCH ₃	OCH ₃	Н	OH	OH	Н				
Acrosin	OCH ₃	OH	OCH ₃	Н	OH	OCH ₃	Н				
5,2',4'-triOH-6,7,8,5'- tetraOMe (Agecorynin D)	OCH ₃	OCH ₃	OCH ₃	ОН	Н	ОН	OCH ₃				
Pectolinarigenin	OCH,	OH	Н	Н	Н	OCH,	Н	Ononis natrix			
Hispidulin	OCH,	OH	Н	Н	Н	OH	Н	(aerial parts)	[25]		
Desmethylsudachitin	OCH ₃	OH	OCH ₃	Н	Н	OH	Н				
Nevadensin	OCH ₃	OH	OCH ₃	Н	Н	OCH ₃	Н				
Nepetin	OCH ₃	OH	Н	Н	OH	OH	Н				
Jacoesidin	OCH ₃	OH	Н	Н	OCH ₃	OH	Н				
Cirsiliol	OCH ₃	OCH ₃	Н	Н	OH	OH	Н				
HO											
		Ĭ				I I					
			\checkmark	····	~^^	\triangleleft	_ОН				
		~	Ĭ		γ	Í					
			Ö				N				
				~	OCH3	Ť	ЮН				
						Сон					
Compound Plant Species Reference											
(3S)-7-hydroxy-4'-methoxy-isoflavanone 3'-β-d- Ononis angustissima											
glucopyranoside (aerial parts)											
Flavonols											
R											
ОН											
HO											
			\bigwedge	$ \longrightarrow $							
				$ \downarrow $							
			Ý	\uparrow	`ОН						
I II он о											

Continued

Compound			R	Plant S	Species		Re	ferences			
Quaraatin	Ononis natrix subsp. hispanica						[16]				
Querceilli	(aerial parts)					[10]					
Kaempferol			Н	Ononis	natrix (leaves)		[26]				
Flavones											
Compound		R		R1	Plant	Species		References			
Apigenin		ОН		Н			1				
Luteolin		OH		OH	Ononis (aerial	<i>natrix</i> subs parts)	sp. hispanica	[16]			
Luteolin 7-methyl ether		ОН		OCH ₃	(F					
Chryseriol		OH		OCH ₃	Ononis	natrix (aer	ial parts)	[25]			
Velutin	(OCH ₃		OCH ₃		[×]	1 /				
Compound	Plant Species Reference							ce			
(tectorigenin 7-O-glucoside)	Ononis spinosa (roots) [27]										
				.0.							
$\begin{array}{c} \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\$											
Compound	R	R ₁	R ₂	R ₃	R ₄	R ₅	Plant Species	References			
Formononetin	ОН	OCH ₃	Н	Н	Н	Н	Ononis spinosa (roots)	[27]			
Ononin (formononetin 7-O-glucoside)	Glucosi-de	OCH ₃	Н	Н	Н	Н	Ononis angustissim (aerial parts)	[7]			
Calycosin	ОН	OCH ₃	ОН	Н	Н	Н	Ononis spinosa (roots)	[28]			
Genistein	ОН	ОН	Н	Н	ОН	Н	Ononis spinosa (roots)	[27]			



Compound	R	R ₁	Plant	species		Refe	rences
Medicarpin	ОН	Н	Ononi	s spinosa (roots)		[2	7,28]
3,4,9-trimethoxypterocarpan	OCH ₃	OCH	. Ononi	s vaginalis (root	s)	[29]
		Pheno	lic Lactones				
	HO			OH			
Compound	Plants	species				Reference	
	Ononis	s spinosa					
Ononilactone	Ononis	s arvensis				[13]	
	(hairy	root cultures)					
		ин	HO				
Compound	Plant	species				Reference	
Bulatlactone 2"-O-β-d-glucoside	Ononis (hairy	s spinosa & On root cultures)	onis arvensis			[13]	
		Dihydro	isocoumarins				
R	R1 O			R ₃ R ₄	CH	l3	
Compound	R	R1	R2	R3	R4	Plant Species	References
3,4-Dihydro-8-hydroxy-6-methoxy-3-(6- hydroxyundecyl)-1H-2-benzopyran-1-one	OCH ₃	ОН	Н	ОН	Н	Ononis natrix (aerial parts)	[24]
6,8-Dihydroxy-3-(8'-hydroxyundecyl)-3,4- dihydroisocoumarin	ОН	ОН	Н	Н	ОН		
3,8-Dihydroxy-6-methoxy-3-undecyl-3,4- dihydroisocoumarin	OCH ₃	ОН	ОН	Н	Н	Ononis natrix	[30]
6,8-dihydroxy-3-(6-oxoundecyl)-3,4- dihydroisocoumarin	ОН	ОН	Н	Н	Н	(aerial parts)	[20]
8-hydroxy-3,6-dimethoxy-3-undecyl-3,4- dihydroisocoumarin	OCH ₃	ОН	OCH ₃	Н	Н		

8-Hydroxy-6-methoxy-3-undecyl-3,4- dihydroisocoumarin	OCH ₃	ОН	Н	Н	Н		
6,8-Dihydroxy-3-undecyl-3,4- dihydroisocoumarin	ОН	ОН	Н	Н	Н		
6,8-Dihydroxy-3-(6-hydroxyundecyl)-3,4- dihydroisocoumarin	ОН	ОН	Н	ОН	Н		
8-Hydroxy-6-methoxy- 3-(6-hydroxyundecyl)- 3,4-dihydroisocoumarin	OCH ₃	ОН	Н	ОН	Н		
6,8-Dimethoxy-3-undecyl-3,4- dihydroisocoumarin	OCH ₃	OCH ₃	Н	Н	Н	(aerial parts)	[12]
6,8-Dihydroxy- 3-(6-oxoundecyl)-3,4-dihydroisocoumarin	ОН	ОН	Н	=0	Н		
8-Hydroxy-6-methoxy-3-(6-oxoundecyl)- 3,4-dihydroisocoumarin	OCH ₃	OH	Н	=0	Н		
6,8-Dimethoxy-3-(6-oxoundecyl)-3,4- dihydroisocoumarin	OCH ₃	OCH ₃	Н	=0	Н		
6,8-Diacetoxy- 3-(6-acetoxyundecyl)- 3,4. dihydroisocoumarin	O-COCH ₃	O-COCH ₃	Н	O-COCH ₃	Н		
	но			<u> </u>	СН3		
Compound	Plai	nt Species				Reference	
6,8-Dihydroxy-3-(7'-oxoundecyl)-3,4- dihydroisocoumarin	[30]						
	(aer		derivative				
	R	itesoi enio					
	но	R ₁		R ₂	СН3		
Compound		R		R ₁	R ₂	Plant species	Reference
5-(2'-acetoxytridecyl) resorcinol methyl eth	ner	OCH ₃	O-0	CH ₃ CO	Н	2	
5-(2'-acetoxy-tridecyl) resorcinol		OH	O-0	CH ₃ CO	Н	<i>Ononis</i> <i>natrix</i> subsp	
5-(2'-hydroxytridecyl) resorcinol methyl et	her	OCH ₃		OH	OH	hispanica	[31]
5-(2',8'-dihydroxytridecyl) resorcinol		OH		OH	OH	(aerial parts)	
5-(2',8'-dihydroxytridecyl) resorcinol meth	oH	OCH ₃	0	ОН	ОН		
	но		, Ľ	$\sim \sim$	СН3		
Compound	Reference						
-(2'-hydroxy-8'-oxytridecyl) resorcinol (aerial parts)						[31]	
	OCH3	- F.m. (2)					

Compound		Reference						
	Ononis natrix			[20]				
5-(7-Oxotridecyi)-3-methoxyphenol	(aerial parts)		[30]					
R	0 R2	~~~	R ₃	СНз				
Compound	R	R ₁	R ₂	R ₃	Plant species	Reference		
1-O-Methyl-5-(2'-acetoxytridecyl) resorcinol	CH ₃	Н	CH ₃ CO	Н				
1-O-Methyl-5-(2'-acetoxy-8'-oxotridecyl) resorcinol	CH ₃	Н	CH ₃ CO	=0	<i>Ononis natrix</i> (aerial parts)			
1-O-methyl-5-(2'-acetoxy-8'-hydroxytridecyl) resorcinol	CH ₃	Н	CH ₃ CO	ОН				
1-O-Methyl-5-(2'-hydroxytridecyl) resorcinol	CH ₃	Н	Н	Н				
1-O-Methyl-5-(2'-hydroxy-8'-oxotridecyl) resorcinol	CH ₃	Н	Н	=0		[11]		
1-O-Methyl-5-(2',8'-dihydroxytridecyl) resorcinol	CH ₃	Н	Н	OH				
5-(2'-Acetoxy-8'-oxotridecyl) resorcinol	Н	Н	CH ₃ CO	=0				
5-(2'-Acetoxy-8'-hydroxytridecyl) resorcinol	Н	Н	CH ₃ CO	OH				
5-(2'-Acetoxy-8'-oxotridecyl) resorcinol diacetate	CH ₃ CO	CH ₃ CO	CH ₃ CO	=О				
5-(2',8'-Diacetoxytridecyl) resorcinol diacetate	CH ₃ CO	CH ₃ CO	CH ₃ CO	O-CH ₃ CO				

bladder, cervical, lung, laryngeal, glioma, adrenal medulla, multiple myeloma, and osteosarcoma cancer. It arrests the G0/G1 phase of the cell cycle in ES2 and the G1 phase in human ovarian and lung cancerous cells and triggers apoptosis in breast cancerous cells [32]. Data about formononetin structure–activity relationships are extremely limited. One study suggested that the presence of a 2–3 double bond, a 4-carbonyl group, and ortho- as opposed to meta-hydroxylation in the B ring greatly increased the cytotoxic activity. Comparing 3-hydroxylated molecules to their non-hydroxylated counterparts, the latter showed noticeably greater cytotoxicity [33].

As for the antioxidant activity, the total number of hydroxyl groups has a significant influence. For example, apigenin, kaempferol, luteolin, and quercetin reduced nitric oxide and phagocytosis in a dose-dependent manner, and the phenolic hydroxyl groups number was proportional to their antioxidant effect [34]. Additionally, the presence of a 3',4'-catechol structure in the B ring greatly increased the potential to inhibit lipid peroxidation. Thus, flavonoids are most efficient in scavenging peroxynitrite, superoxide, and peroxyl radicals. Due to oxidation on the flavonoid B ring with a catechol group, a relatively stable ortho-semiguinone radical that acts as a potent scavenger is produced. In contrast, flavones without a catechol system form unstable radicals when oxidized, showing less scavenging potential [35]. It is worth to mention that flavones and catechins appear to have the most potent activity against reactive oxygen species [35].

Concerning the antimicrobial activity, several structurally distinct flavonoids, such as isoflavones, flavones, isoflavanones, and flavanones, show that *Streptococcus sobrinus* and *Streptococcus mutans* growth was inhibited by 5-hydroxyisoflavanones and 5-hydroxyflavanones with one,

two, or three additional hydroxyl groups at the 7, 2', and 4' positions, with position 2' hydroxylation being crucial for the anti-staphylococcal activity. Additionally, hydroxyl groups at position 5 of flavones and flavanones are critical for their anti-MRSA action. It was also evident that chalcones work better than flavanones or flavones against MRSA. On the contrary, it has been observed that methoxy groups significantly reduce flavonoids' antibacterial activity [36].

For type 2 diabetes mellitus, quercetin has the highest IC_{50} value to inhibit α -glucosidase and dipeptidyl peptidase IV. The existence of the C-2-C-3 double bond and the C-4 ketonic group is crucial to its antidiabetic properties. However, acetylation, methylation, and hydroxyl groups reduce the antidiabetic *in-vitro* effects of flavonoids [37].

Toxicity and safety assessments

Till this date, no studies assessing the toxicity and safety of the Ononis species have been reported, so the literature is lacking in this regard. However, there are several investigations that have evaluated the safety of Ononis-based compounds. For example, quercetin, one of the most common and known flavonoids in the Ononis species, has shown conflicting results. One study proved the safety of quercetin when taken orally for 98 days in male and female mice (doses ~12.5, 25, or 50 mg/kg of body weight) [38]. On the other hand, higher doses of quercetin >2,000 mg/kg were associated with hepatotoxic stress in mice liver [39], and a dose of 3807 mg/kg was lethal 22 hours after administration to mice [40]. Moreover, sodium formononetin-30-sulphonate, a water-soluble derivative of formononetin, was considered safe and showed no side effects at doses <100 mg/kg in dogs [41], and formononetin was safe at a dose of 1.5 mg/kg in mice [42]. The lethal dose of 50% was

also considered to be 103.6 mg/kg, and no side effects were exhibited at doses <50 mg/kg [43].

The findings of the collected studies showed no discrepancies, contraindications, or conflicting viewpoints concerning pharmacological activities and associated structureactivity relationships. On the contrary, the main results of the different studies were consistent, enhancing the credibility of the available literature. However, it is important to mention that while most biological activities of Ononis flavonoids and their structure-activity relationship are well documented, there is a lack of studies investigating their cytotoxic activity. Accordingly, the cytotoxic activity of Ononis flavonoids needs to be thoroughly investigated to determine a detailed structure-activity relationship that could pave the way in the process of drug discovery. In addition, the available studies are mostly in vitro evaluations except for the antidiabetic activity, which has been studied in vivo. In addition, it is noteworthy to mention that the toxicity of some Ononis flavonoids has been assessed, but the Ononis plants or their extracts have not been studied in this aspect, thereby necessitating the need for further investigations. All in all, this is the first review article that summarizes the constituents of different Ononis species and their pharmacological activities, as well as highlighting their structure-activity relationship, thereby offering a collective review of these species. The use of several recent studies emphasizes the growing interest in the Ononis species and the importance of their therapeutic potential in various diseases and stimulates further investigations of these plants.

CONCLUSION

This review of genus *Ononis* highlights the presence of diverse bioactive compounds, mainly flavonoids and isoflavonoids, responsible for the pharmacological potential of these species. Based on the study findings, *Ononis* species could be considered important therapeutic remedies in curing oxidative stress, inflammation, infections, diabetes, and some cancers. It opens the gates for researchers to prepare more potent derivatives from the phenolic and flavonoid content of the *Ononis* plant. These derivatives could serve as novel molecule in the process of drug discovery. Furthermore, the study highlighted the therapeutic potential of the *Ononis* plant as preventive, treatment, and even adjuvant measures for various diseases. Preclinical and clinical studies will be recommended to support its clinical applications.

ACKNOWLEDGMENTS

The authors thank Beirut Arab University for providing the support throughout the manuscript preparation. The authors thank Acdlabs.com (ChemSketch free version) for making it possible to create all of the figures.

AUTHOR CONTRIBUTIONS

All authors made substantial contributions to conception and design, acquisition of data, or analysis and interpretation of data; took part in drafting the article or revising it critically for important intellectual content; agreed to submit to the current journal; gave final approval of the version to be published; and agree to be accountable for all aspects of the work. All the authors are eligible to be an author as per the International Committee of Medical Journal Editors (ICMJE) requirements/guidelines.

FINANCIAL SUPPORT

This research was not supported by funding sources in the public, private, or not-for-profit sectors.

CONFLICTS OF INTEREST

The authors report no financial or any other conflicts of interest in this work.

ETHICAL APPROVALS

This study does not involve experiments on animals or human subjects.

DATA AVAILABILITY

The data supporting the results of this research are accessible in standard research databases such as PubMed, Scopus, ScienceDirect, Google Scholar, The Cochrane Library, Web of Science, and/or public domains that are accessible via keywords or DOI numbers.

PUBLISHER'S NOTE

All claims expressed in this article are solely those of the authors and do not necessarily represent those of the publisher, the editors and the reviewers. This journal remains neutral with regard to jurisdictional claims in published institutional affiliation.

USE OF ARTIFICIAL INTELLIGENCE (AI)-ASSISTED TECHNOLOGY

The authors state that they have not utilized artificial intelligence (AI) tools for writing and editing of the manuscript, and no pictures were modified using AI.

REFERENCES

- Tekdal D. Plant genes for abiotic stress in legumes. Abiotic Stress and Legumes. Singh VP, Singh S, Tripathi DK, Prasad SM, Bhardwaj R, Chauhan DK, editors. Academic Press; 2021. Pp 291–301.
- 2. Gao T, Yao H, Song J, Liu C, Zhu Y, Ma X, *et al.* Identification of medicinal plants in the family *Fabaceae* using a potential DNA barcode ITS2. J Ethnopharmacol. 2010;130(1):116–21.
- Xu Z, Deng M, Xu Z, Deng M. Fabaceae or Leguminosae. Identification and Control of Common Weeds. Vol. 2. 2017. Pp 547– 615. Available from: https://doi.org/10.1007/978-94-024-1157-7 43
- 4. Maroyi A. Medicinal uses of the *Fabaceae* family in Zimbabwe: a review. Plants. 2023;12(6):1255.
- Öz BE, İşcan GS, Akkol EK, Süntar İ, Keleş H, Acıkara ÖB. Wound healing and anti-inflammatory activity of some *Ononis* taxons. Biomedicine Pharmacother. 2017;91:1096–105.
- 6. Mhamdi B, Abbassi F, Abdelly C. Chemical composition, antioxidant and antimicrobial activities of the edible medicinal *Ononis natrix* growing wild in Tunisia. Nat Prod Res. 2015;29(12):1157–60.
- Mezrag A, Malafronte N, Bouheroum M, Travaglino C, Russo D, Milella L, *et al.* Phytochemical and antioxidant activity studies on *Ononis angustissima L.* aerial parts: isolation of two new flavonoids. Nat Prod Res. 2017;31(5):507–14.
- Ghribi L, Waffo-Téguo P, Cluzet S, Marchal A, Marques J, Mérillon JM, *et al.* Isolation and structure elucidation of bioactive compounds

from the roots of the Tunisian *Ononis angustissima L*. Bioorgan Med Chem Lett. 2015;25(18):3825–30.

- 9. Al-Snafi AE. The traditional uses, constituents and pharmacological effects of *Ononis spinosa*. IOSR J Pharm. 2020;10(2):53–9.
- Ullah A, Munir S, Badshah SL, Khan N, Ghani L, Poulson BG, et al. Important flavonoids and their role as a therapeutic agent. Molecules. 2020;25(22):5243.
- Cañedo LM, del Corral JM, San Feliciano A. 5-Alkylresorcinols from Ononis natrix. Phytochemistry. 1997;44(8):1559–63.
- San Feliciano A, del Corral JM, Cañedo LM, Medarde M. 3, 4-Dihydroisocoumarins from *Ononis natrix*. Phytochemistry. 1990;29(3):945–8.
- Gampe N, Szakács Z, Darcsi A, Boldizsár I, Szőke É, Kuzovkina I, et al. Qualitative and quantitative phytochemical analysis of *Ononis* hairy root cultures. Front Plant Sci. 2021;11:622585.
- Abu Zarga MH, Al-Jaber HI, Al-Qudah MA, Al-Aboudi AM. A new cyclic polyketide and other constituents from *Ononis spinosa* growing wildly in Jordan and their antioxidant activity. J Asian Nat Prod Res. 2022;24(3):290–5.
- Besbas S, Mouffouk S, Haba H, Marcourt L, Wolfender JL, Benkhaled M. Chemical composition, antioxidant, antihemolytic and anti-inflammatory activities of *Ononis mitissima L*. Phytochem Lett. 2020;37:63–9.
- Yerlikaya S, Zengin G, Mollica A, Baloglu MC, Celik Altunoglu Y, Aktumsek A. A multidirectional perspective for novel functional products: *in vitro* pharmacological activities and in silico studies on *Ononis natrix* subsp. *hispanica*. Front Pharmacol. 2017;8:600.
- Dénes T, Bartha SG, Kerényi M, Varga E, Balázs VL, Csepregi R, *et al.* Histological and antimicrobial study of *Ononis arvensis L*. Acta Biol Hung. 2017;68:321–33.
- Al-Zereini WA. Ononis natrix and Salvia verbenaca: two Jordanian medicinal plants with cytotoxic and antibacterial activities. J Herbs Spices Med Plants. 2017;23(1):18–25. Available from: https://doi.or g/10.1080/10496475.2016.1241200
- Spiegler V, Gierlikowska B, Saenger T, Addotey JN, Sendker J, Jose J, *et al.* Root extracts from *Ononis spinosa* inhibit IL-8 release via interactions with toll-like receptor 4 and lipopolysaccharide. Front Pharmacol. 2020;11:889.
- Al-Mubideen BF, Al-Serhan AA, Amarin JZ, Al-Dweikat A, Al-Muhaisen RA, Shreikh YA, *et al. Ononis natrix L.* lowers the blood glucose concentration in wistar rats with streptozotocin-induced diabetes mellitus. Endocr Metab Immune Disord Drug Targets. 2021;21(5):854–8.
- Al-Mterin MA, Aboalhaija N, Zihlif MA, Afifi FU. Effects of *Ononis* natrix on glucose and lipid metabolism: an *in vivo* study. J Res Pharm. 2024;28(1):278–88.
- El Khiat A, Bouftini K, Lafhal K, Tastift MA, El-Mansoury B, Ali DA, *et al.* Evaluation of the Anti-hyperglycemic activity of *Ononis natrix* aqueous extract against alloxan-induced experimental model of insulinopenic diabetes in albino Swiss mice. Academic J. 2024;39(4):57–64.
- Kanso MA, Hijazi MA, El-Lakany A, Aboul-Ela M. Review on phytochemical constituents and pharmacological activities of genus *Galium*. J Appl Pharm Sci. 2024;14(9):046–56.
- Al-Khalil S, Masalmeh A, Abdalla S, Tosa H, Iinuma M. N-arachidylanthranilic acid, a new derivative from *Ononis natrix*. J Nat Prod. 1995;58(5):760–3. Available from: https://doi.org/10.1021/ np50119a018
- Wollenweber E, Dörr M, Rivera D, Roitman JN. Externally accumulated flavonoids in three Mediterranean *Ononis* species. Zeitschrift fuer Naturforschung C. 2003;58(11-12):771–5.
- 26. Gampe N, Darcsi A, Lohner S, Béni S, Kursinszki L. Characterization and identification of isoflavonoid glycosides in the root of Spiny

restharrow (*Ononis spinosa L.*) by HPLC-QTOF-MS, HPLC-MS/ MS and NMR. J Pharm Biomed Anal. 2016;123:74–81.

- 27. Gampe N, Dávid DN, Takács-Novák K, Backlund A, Béni S. *In vitro* and *in silico* evaluation of *Ononis* isoflavonoids as molecules targeting the central nervous system. PLoS One. 2022;17(3):e0265639.
- Gampe N, Darcsi A, Nagyné Nedves A, Boldizsár I, Kursinszki L, Béni S. Phytochemical analysis of *Ononis arvensis L*. by liquid chromatography coupled with mass spectrometry. J Mass Spectrometry. 2019;54(2):121–33.
- 29. Abdel-Kader MS. Phenolic constituents of *Ononis vaginalis* roots. Planta Medica. 2001;67(04):388–90.
- Yousaf M, Al-Rehaily AJ, Ahmad MS, Mustafa J, Al-Yahya MA, Al-Said MS, *et al.* A 5-alkylresorcinol and three3, 4-dihydroisocoumarins derived from *Ononis natrix*. Phytochem Lett. 2015;13:1–5.
- Barrero AF, Sánchez JF, Rodríguez I. N-Δ13-Docosenoylanthranilic acid and alkylresorcinols from *Ononis natrix* subsp. *hispanica*. Phytochemistry. 1990;29(6):1967–9.
- Jiang D, Rasul A, Batool R, Sarfraz I, Hussain G, Mateen Tahir M, et al. Potential anticancer properties and mechanisms of action of formononetin. BioMed Res Int. 2019;2019(1):5854315. Available from: https://doi.org/10.1155/2019/5854315
- Plochmann K, Korte G, Koutsilieri E, Richling E, Riederer P, Rethwilm A, *et al.* Structure-activity relationships of flavonoidinduced cytotoxicity on human leukemia cells. Arch Biochem Biophy. 2007;460(1):1–9.
- 34. Tian C, Liu X, Chang Y, Wang R, Lv T, Cui C, *et al.* Investigation of the anti-inflammatory and antioxidant activities of luteolin, kaempferol, apigenin and quercetin. South African J Bot. 2021;137:257–64.
- 35. Kumar S, Pandey AK. Chemistry and biological activities of flavonoids: an overview. Sci World J. 2013;2013(1):162750.
- Cushnie TT, Lamb AJ. Antimicrobial activity of flavonoids. Int J Antimicrob Agents. 2005;26(5):343–56.
- Sarian MN, Ahmed QU, Mat So'ad SZ, Alhassan AM, Murugesu S, Perumal V, *et al.* Antioxidant and antidiabetic effects of flavonoids: a structure-activity relationship based study. BioMed Res Int. 2017;2017(1):8386065.
- Cunningham P, Patton E, VanderVeen BN, Unger C, Aladhami A, Enos RT, *et al.* Sub-chronic oral toxicity screening of quercetin in mice. BMC Complementary Med Ther. 2022;22(1):279.
- Singh P, Sharma S, Rath SK. A versatile flavonoid Quercetin: study of its toxicity and differential gene expression in the liver of mice. Phytomed Plus. 2022;2(1):100148.
- 40. Dibal NI, Garba SH, Jacks TW. Acute toxicity of quercetin from onion skin in mice. Pharm Biomed Res. 2020;6(4):269–76.
- Li C, Li G, Gao Y, Sun C, Wang X. A 90-day subchronic toxicity study with sodium formononetin-3'-sulphonate (Sul-F) delivered to dogs via intravenous administration. Regul Toxicol Pharmacol. 2016;77:87–92.
- 42. Singh KB, Dixit M, Dev K, Maurya R, Singh D. Formononetin, a methoxy isoflavone, enhances bone regeneration in a mouse model of cortical bone defect. Br J Nutr. 2017;117(11):1511–22.
- Pingale TD, Gupta GL. Acute and sub-acute toxicity study reveals no dentrimental effect of formononetin in mice upon repeated ip dosing. Toxicol Mech Methods. 2023;33(8):688–97.

How to cite this article:

Chouman A, El-lakany A, Aboul-ela M, Ali Hijazi M. Review on phenolic constituents and pharmacological activities of genus *Ononis*. J Appl Pharm Sci. 2025;15(07):027–036. DOI: 10.7324/JAPS.2025.234398