Separation, Quantification and Control of Enatiomers of the Key Starting Material of Dextromethorphan Hydrobromide

Ajit Anerao*, Vishal Solase, Amol More, Nitin Pradhan
R&D Centre (API), Wanbury Ltd., EL-16, TTC Industrial Estate, Mahape, Navi Mumbai 400710, India.

ABSTRACT

Objective: Enantiomeric resolution of racemic 1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline, a key starting material in the synthesis of the antitussive agent dextromethorphan hydrobromide. Material and method: Chiral liquid chromatographic method is developed and enantiomers are resolved on a Chiralpak ID-3 column using gradient mode of elution. Mobile phase A and B containing a mixture of 0.1% diethylamine in acetonitrile and 0.1% diethylamine in methanol respectively are optimized with the timed programme. The flow rate is 1.0 mL/minute and the column temperature was maintained at 40°C. Ultraviolet detection was performed at 280 nm wavelength and the injection volume was 20 μL. Results: The resolution between the enantiomers is found to be more than 2.5. The developed method is extensively validated and proved to be robust. The limit of detection and limit of quantification of unwanted (R)-enantiomer is found to be 0.0075% and 0.025% with respect to test concentration, respectively. The percentage recovery of (R)-enantiomer is ranged from 88.6 to 106.6 in (S)-(-)-1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline sample solution. Conclusion: The proposed method is found to be suitable and accurate for the quantitative determination of (R)-enantiomer in the key starting material of the dextromethorphan hydrobromide bulk drugs.

INTRODUCTION

Dextromethorphan hydrobromide is the pharmacologically active ingredient of NUEDEXTA that acts on the central nervous system (CNS). The chemical name is Morphinan, 3-methoxy-17-methyl-, (9α, 13α, 14α), hydrobromide monohydrate (Rxlist, 2017). Dextromethorphan is the d-isomer of 3-methoxy-N-methyl-morphinan, a synthetic analog of codeine. D-Methorphan is widely used all over the world as an over-the-counter (OTC) medication to treat a cough (PDR, 2018; Magellan Rx Management, 2016; Cersósimo and Micheli, 2007; EMA, 2013). Levomethorphan is the 1-isomer of 3-methoxy-N-methyl-morphinan and, unlike d-methorphan, is a potent narcotic analgesic. For this reason, levomethorphan is not commercially available and in some countries, it is listed as a controlled substance (Bortolotti et al., 2013). Dextromethorphan is an official drug listed in the United State Pharmacopeia (USP) (USP, 2017) as well as European Pharmacopeia (EP) (EDQM, 2017). In USP 40, the unwanted levomethorphan is tested by high-performance liquid chromatography (HPLC) method and controlled with the limit 0.10% while in EP 9.0 it is controlled with an optical rotation method. Recently in the month of March 2018, EP has published a draft monograph by including enantiomeric selective HPLC method in Pharmeuropa (Pharmeuropa, 2017). Owing to serious incidents due to the abnormally high concentration of levomethorphan in dextromethorphan, it is proposed by Pharmeuropa to add a test for enantiomeric purity by LC, which is more specific than the current test of optical rotation. The chiral center responsible for the formation of levomethorphan is generated in the synthetic process while cyclization and formation of the key starting material, 1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline. Racemic 1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline is further resolved in the synthetic process using resolution agents like tartaric acid, mandelic acid or (R)-2-(6-methoxy-2-naphthyl) propionic acid (Divi et al. 2012; Demian et al., 2000). As per International Council of Harmonization...
(ICH) guideline Q3A R2, during drug development, it is necessary to list out and discuss all possible process impurities, degradants, raw materials, intermediates, reagents and catalysts used in the manufacturing drug substance. Impurity formation, their source of origin, control, fate and purge to be studied in order to establish safety, efficacy, and quality of drug substance (ICH, 2006). On basis of the synthetic chemistry of manufacturing dextromethorphan, it is required to control the unwanted enantiomer i.e. (R)-(+) and capillary electrophoresis (Krafft et al. 2018; Lurie and Kimberly, 2005) but only one method is found to control the unwanted enantiomer in key starting material (Guangyue et al. 2016). In that reported method, both enantiomers were separated on chiralcel OJ-H column with mobile phase a mixture of hexane, isopropyl alcohol, and ethanolamine but this method is not selective. The d-mandelic acid used for the resolution of the racemic 1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline is not eluting in this method. Hence the method is not suitable for the intended purpose. The major objective of the present work is to develop a method for the separation of R-isomer and S-isomer of 1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline and the method should be selective and have a low limit of detection and quantification (LOD and LOQ) values. Further, the use of immobilized chiral HPLC column ensures the column life, unlike glycoprotein-based columns. This method was successfully validated according to the ICH guidelines of method validation (ICH, 2005).

MATERIALS AND METHODS

Method of analysis

Instrumentation

Shimadzu high-performance liquid chromatography (HPLC) system LC-2010 CHT with UV detector with LC solutions software or its equivalent.

Chemicals and reagents

HPLC column used by Daicel Corporation, Chiralpak ID-3, length 250 mn, internal diameter 4.6mm, particle size 3μm. (S)-1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline standard was procured from Roopa Industries Ltd, Hyderabad, India of potency 92.6% and (R)-1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline standard was synthesized at Wansbury R&D Center of potency 90.6%. Both standards were characterized by NMR, mass, IR and SOE and structures were confirmed. Acetonitrile, methanol and diethylamine HPLC grade from Merck was used in the experimentation. During method development, Daicel chiral stationary phases Chiralpak IG, Chiralpak AD-H, Chiralpak OJ-H and Phenomenex Lux amylose-2 were used for the trial purpose.

Chromatographic parameters

HPLC method was performed using a Chiralpak ID-3 HPLC column. Separation and peak symmetry were achieved with gradient elution of mobile phase A and B. Mobile phase A was 0.1% diethylamine in acetonitrile and mobile phase B was 0.1% diethylamine in methanol. The gradient elution with timed programme T0/A:B T0/100:00; T0/00:100; T0/00:10; T0/100:00 and T0/100:00 with flow rate 1.0 mL/min was applied. The column temperature was maintained at 40°C. Ultraviolet detection was performed at 280 nm wavelength and the injection volume was 20 μL. Run time for blank, system suitability solution and test solution is 40.0 minutes and for a standard solution, it is 10.0 minutes.

Solution preparation

Diluent preparation

A mixture of an equal volume of methanol and acetonitrile was prepared.

Standard stock solution-A

Accurately weighed and transferred 30.0 mg of (R)-1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline standard into 50 mL volumetric Flask. Dissolved and diluted up to the mark with diluent.

Standard stock solution-B

Transferred the standard stock solution-A 5.0 mL into 50 mL volumetric Flask. Dissolved and diluted up to the mark with diluent.

Standard solution

Further transferred 10.0 mL of standard stock solution-B into 20 mL volumetric flask and dilute up to mark with diluent.

System suitability solution

Accurately weighed and transferred 60.0 mg of (S)-1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline standard into 20 mL volumetric flask. Transferred 10.0 mL of standard stock solution-B, dissolved and diluted up to the mark with diluent.

Test solution

Accurately weighed and transferred 60.0 mg of the test sample into 20 mL volumetric flask. Dissolved and diluted up to the mark with diluent.

Procedure

Injected blank followed by system suitability solution, standard solution six times and test preparation in duplicate. The chromatograph was run for 40.0 minutes and the chromatogram recorded. The retention time of (S)-1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline is 6.1 minutes and (R)-1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline is 8.0 minutes. Both isomer peaks were integrated into system suitability and test solution and unwanted enantiomer were calculated by external standard addition method.
Acceptance criteria for system suitability

The peaks of (S)-1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline and R enantiomer in system suitability solution was found a base to base separated and the resolution between both isomers was more than 2.5.

RESULTS & DISCUSSION

Analytical method development

1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline is an optically active compound with the single chiral center. The molecular formula is $C_{17}H_{17}NO$ and molecular weight 257.37 (CAS, 2018). Optical rotation of S enantiomer is $\left[\alpha\right]_{D}^{20} = -152^\circ$, $c = 1$ in chloroform with CAS No. 30356-07-1 (CAS, 2018). Optical rotation of R enantiomer is $\left[\alpha\right]_{D}^{20} = +152^\circ$, $c = 1$ in chloroform with CAS No. 30356-08-2 (Sigma-Aldrich, 2018; CAS, 2018). Refer structure of both isomers in Figures. 1 and 2. It is UV active compound with strong chromophores of phenyl and quinoline aromatic rings. The solubility of both isomers was checked in various polar to non-polar organic solvents and found that those are soluble in methanol, acetonitrile, isopropyl alcohol, methylene chloride and chloroform. It is very slightly soluble in n-hexane but found practically insoluble in water. On basis of chemical properties and literature survey, it was decided to start method development using normal phase chiral HPLC technique with ultra-violet (UV) detector.

![Structure of the key starting material](image)

Fig. 1: Structure of the key starting material (S)-1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline.

![Structure of unwanted isomer](image)

Fig. 2: Structure of unwanted isomer (R)-1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline.

Method evaluation of the reported method on chiralpak AD-H column was performed with a mobile phase a mixture of hexane, isopropyl alcohol, and ethanolamine. In this method, both enantiomers were separated but the method is non-selective. The d-mandelic acid used for the resolution of the racemic 1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline is not eluting in this method. Mandelic acid gets eluted after increasing composition of isopropyl alcohol but both enantiomers were found merging. Using the same mobile phase composition chiralpak AD-H column with stationary phase amyllose tris(3,5-dimethylphenylcarbamate) was tried. Here enantiomers were not separated base to base and peaks were asymmetrical with tailing factor more than 2.0 refer Figure 3. Maximum resolution of more than 4.0 with clearly separated peaks was observed on two stationary phases namely chiralpak IG-3 i.e. amyllose tris(3-chloro-5-methylphenylcarbamate) and Lux Amylose-2 i.e. amyllose tris(2-methyl-5-chlorophenylcarbamate). Enantiomer peaks were symmetrical but D-mandellic acid was not eluting out from the column. Refer Figures 4 and 5. After few trials of stationary and mobile phases chiralpak ID-3 column was selected where both enantiomers were resolved as well as D-mandellic acid and other known process impurities also eluting out from the column. Chiralpak ID-3 column i.e. amyllose tris(3-chlorophenylcarbamate) immobilized on 3 μm silica gel was found suitable for the analysis. Here (S)-1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline is eluting at a retention time (RT) 6.1 minutes and unwanted R enantiomer at 8.0 minutes. With gradient elution of methanol and acetonitrile, D-mandellic acid was eluting at RT 15.2 minutes. In the mixture of methanol and acetonitrile as mobile phase, both enantiomers were separated base to base but peaks were not symmetrical having tailing more than 4.0. The core structure of the analyte consist of octahydroisoquinoline which is secondary amine has strongest basic pKa 9.36. On basis of the chemical property of the compound decided to add basic additive diethylamine in the mobile phase and found both enantiomer peaks were become sharp and symmetrical without compromising resolution.

Analytical method validation

Selectivity

Selectivity is the ability of the method to determine accurately and specifically the analyte of interest in the presence of other components in a sample matrix that may be expected to be present in the sample matrix under the stated conditions. Selectivity of the method was evidenced by comparing blank, R and S isomers of 1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline and other specified impurities separate injections. There is no interfering peak at the retention time of R and S isomers of 1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline observed. Both isomer peaks were well resolved from all another specified impurity.

Solution stability

Drug stability in Active Pharmaceutical Ingredient (API) is a function of storage conditions and chemical properties of the drug and its impurities. The solution stability till nine hours of 1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline was checked by injecting standard and spike system suitability solutions separately. Both solutions were prepared fresh before injection.
and immediately injected and same were injected after nine hours. The peak area of R and S isomers of 1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline of the freshly prepared standard solution was observed 201998 and 218382 after nine hours it was 202366 and 218803 respectively. Thus, no significant change in R and S isomers was observed till nine hours. Similarly, solution stability of system suitability solution was checked. In a fresh solution area of R, enantiomer was 483966 and it was found unchanged 483214 till fifteen hours.

Fig. 3: Chromatogram of isomers partially resolved on chiralpak AD-H column.

Fig. 4: Chromatogram of isomers well resolved on chiralpak IG-3 column.

Fig. 5: Chromatogram of isomers well resolved on Lux amylose-2 column.
Limit of detection and quantification

The limit of detection (LOD) is the lowest concentration of the analyte in a sample that can be detected but not necessarily quantified. The obtained LOD value of (R)-1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline is discussed.

\[ \text{LOD} = 3.3 \times \sigma / S \]

The limit of quantitation (LOQ) is the lowest concentration or amount of the analyte that can be determined quantitatively within an acceptable level of repeatability precision and trueness.

\[ \text{LOQ} = 10.0 \times \sigma / S \]

Where, \( \sigma \) = the standard deviation of the response and \( S \) = slope of the calibration curve.

LOD and LOQ concentration of unwanted R enantiomer are reported with respect to (S)-1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline test solution concentration. LOD is 0.0075% i.e. 0.075 mg/g and LOQ is 0.025% i.e. 0.25 mg/g. The precision of LOQ is checked by injecting six replicate injections. Relative standard deviation (RSD) of the peak area of (R)-1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline at LOQ level is observed 1.25% which proves consistency and reproducibility of the method at a low level.

Linearity

Under the experimental conditions, the peak area of unwanted R enantiomer vs. concentration plot for the proposed method was found to be linear over the range of LOQ level, 25%, 50% 100%, 120% and 150% of the specified limit with a regression coefficient is 0.999. The regression coefficient (\( r^2 \)) more than 0.99 is generally considered as evidence of an acceptable fit of the data to the regression line. Refer Figure 8 of linearity plot where the concentration of (R)-1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline standard was plotted on abscissa that is independent variable and peak area is on the ordinate.

Accuracy

Accuracy can be defined as the closeness of agreement between a test result and the accepted reference value. The accuracy of the method was determined by recovery study. Analytical method may be considered validated in terms of accuracy if the mean value is within ± 20% of the actual value. During recovery study (S)-1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-
octahydroisoquinoline batch was analyzed and then R enantiomer was spiked in the (S)-1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline at LOQ level, 50%, 100% and 150% with respect to the limit of R-enantiomer that is 1.0%. The recovery is found at 88.6%, 106.6%, 94.7% and 95.3% respectively which proves that the method is capable to quantify the analyte accurately and results are reliable even at trace level.

**Ruggedness study**

Ruggedness refers to the (intra-laboratory tested) behavior of an analytical process when small changes in the environment and/or operating condition are made. The ruggedness of the method was evaluated by estimating % RSD of (R)-1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline standard solution tested by two different analysts on different days. One batch of (S)-1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline was spiked with unwanted R-enantiomer at 100% level and such six different preparations were prepared by each analyst. RSD of the content of R enantiomer of total twelve preparations was found to be 1.9%, which proves that there is no variation in day to day analysis.

![Fig. 8: Linearity plot of unwanted isomer (R)-1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline.](image)

**Table 1.** Method validation summary.

<table>
<thead>
<tr>
<th>S.NO.</th>
<th>PARAMETER</th>
<th>ACCEPTANCE CRITERIA</th>
<th>RESULTS</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Selectivity</td>
<td>R and S enantiomer peaks should be well separated from all known and unknown peaks of (S)-1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline</td>
<td>Method is selective.</td>
</tr>
<tr>
<td>2</td>
<td>Solution stability</td>
<td>Report Result</td>
<td>Solutions are stable up to 15 hrs</td>
</tr>
<tr>
<td>3</td>
<td>Limit of detection</td>
<td>LOD of R enantiomer</td>
<td>0.037% wtr test concentration</td>
</tr>
<tr>
<td>4</td>
<td>Limit of quantification</td>
<td>LOQ of R enantiomer</td>
<td>1.3% wtr test concentration</td>
</tr>
<tr>
<td>5</td>
<td>LOQ precision</td>
<td>% RSD for six replicates of LOQ level standard solutions is NMT: 10.0%</td>
<td>0.999</td>
</tr>
<tr>
<td>6</td>
<td>Linearity</td>
<td>Correlation: NLT 0.99</td>
<td></td>
</tr>
<tr>
<td>7</td>
<td>Accuracy</td>
<td>Recovery should be between 80% to 120%</td>
<td></td>
</tr>
<tr>
<td>8</td>
<td>Method precision</td>
<td>% RSD for results of six test preparations is NMT: 10%</td>
<td>0.75%</td>
</tr>
<tr>
<td>9</td>
<td>Intermediate precision</td>
<td>% RSD for results of twelve test preparations (Method precision and Intermediate precision) is NMT: 10%</td>
<td>1.9%</td>
</tr>
<tr>
<td>10</td>
<td>Robustness: Column temperature (35°C and 45°C)</td>
<td>% RSD of R enantiomer at 35°C</td>
<td>2.5%</td>
</tr>
<tr>
<td></td>
<td></td>
<td>% RSD of R enantiomer at 45°C</td>
<td>1.0%</td>
</tr>
</tbody>
</table>

**Robustness study**

Robustness is a measure of the capacity of the analytical procedure to remain unaffected by small but deliberate variations in method-performance parameters, which provides an indication of its reliability during normal usage. Robustness of the method was determined by analyzing the standard solution and 100% spike solution with a deliberate change in the critical parameter column temperature ± 5°C. System suitability criteria were passing each time and RSD of R enantiomer peak area of six replicate preparations was observed 1.04. Refer to Table 1 of method validation summary.

**CONCLUSION**

The levomethorphan is an unwanted isomer of dextromethorphan hydrochloride drug substance. The chiral center is generated during synthesis of key starting material i.e. (S)-1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline. The R enantiomer of 1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-
octahydroisoquinoline is leading to the formation of the unwanted levomethorphan. Here the quantification of R enantiomer in (S)-1-(4-Methoxybenzyl)-1,2,3,4,5,6,7,8-octahydroisoquinoline method was developed to control the further formation of levomethorphan in the final drug product. The proposed method was validated as per ICH guideline and checked for accuracy and reliability of the result. The chiral HPLC method satisfies all validation parameters like system suitability, precision, specificity, accuracy, the linearity of detector response, ruggedness and robustness. It indicates that the method is stable and suitable for the quantification of R enantiomer in the KSM of dextromethorphan. Hence, the validated method can be used for routine analysis in quality control laboratories in the pharmaceutical industry.

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COMPLIANCE WITH ETHICAL STANDARDS

Conflict of Interest

Author Ajit Anerao, Vishal Solase, Amol More, Nitin Pradhan declare that they have no conflict of interest.

Ethical Approval

This article does not contain any studies with human participants or animals performed by any of the authors.

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