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# Biophysical investigation of curcumin based nanocomposite for wound dressing application

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# ABSTRACT

Curcumin (cur) loaded polyvinyl alcohol (PVA), chitosan (Cs) and PVA/Cs blend (50:50) were prepared with different concentrations of curcumin (0, 5, 10, and 15 wt%) using the solution casting technique and studied for wound dressing application. Physical characterization of the composites achieved through dielectric spectroscopy (DS), fourier transform infrared spectroscopy (FTIR), scanning electron microscope (SEM) and transmission electron microscope (TEM). The wound dressing characteristics were evaluated by the water vapor transmission rate (WVTR), antimicrobial studies and cell viability assay. The scanning electron microscope revealed good dispersion of the nanocurcumin in the matrix. The dielectric properties of all composites were studied through the measurements of the permittivity  $\varepsilon'$  and dielectric loss  $\varepsilon''$ . These results show that both values for PVA/cur nanocomposites greatly increase with increasing curcumin content which was found to be higher than those of the two individuals. On the opposite side, both  $\varepsilon'$  and  $\varepsilon''$  decrease with increasing curcumin content in case of Cs/cur and PVA/Cs/cur nanocomposites. The obtained electrical conductivity for all composites lies in the order of  $(10^{-9}-10^{-14}/S \text{ cm}^{-1})$ . WVTR of the samples decreased by increasing the curcumin %. Results of the cell viability assay confirmed that the addition of the curcumin has no toxic effect on the cells. PVA/cur nanocomposites displayed excellent antimicrobial activity.

# **INTRODUCTION**

Functional biomaterials research has built on new drug delivery systems and enhanced scaffolds for regenerative medicine that is recently one of the most developing fields in the life sciences. Polyvinyl alcohol (PVA) and chitosan (Cs) are biopolymers that are widely used in this field due to their biocompatibility, nontoxicity, noncarcinogenicity, and their swelling properties (Baker *et al.*, 2012; Dash *et al.*, 2011).

Polymer blends are physical mixtures of two or more polymers. Usually, the properties of the blend are superior to the component homopolymers. Some studies on the polyvinyl

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alcohol (PVA)/chitosan blends have been reported. Nakano *et al.* (2007) studied the mechanical and thermal properties of chitosan and polyvinyl alcohol (PVA). The characteristics of the blend films studied using DSC, X-ray, viscoelastic and positron annihilation techniques. The results recommended that the miscibility of chitosan and PVA could be confirmed by entanglement of the amorphous chain segments of chitosan and PVA.

The adhesion and growth of fibroblasts on the PVA and PVA/Cs membranes were investigated by Chuang *et al.* (1999). They concluded that PVA membrane can be modified by blending with chitosan. In addition, they observed that, for cell culture, the PVA/Cs blend was more favorable than the pure PVA.

Curcumin (diferuloylmethane, 1, 7-bis (4-hydroxy-3methoxyphenyl)-1, 6-heptadiene-3, 5-dione) is a polyphenolic compound which is a natural product obtained from an Asian spice called turmeric. Curcumin is a member of the ginger family. It is believed to have many biological activities such as antioxidant,

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anti-inflammatory, and even anticancer effect. Several studies suggest that it might reduce inflammation and pain symptoms resulted from osteoarthritis and rheumatoid arthritis (Weber *et al.*, 2005; Shankar *et al.*, 2012; Fan *et al.* 2013).

Curcumin has been revealed high cytotoxicity towards different cancer cells, but its instability and water-insolubility make its bioavailability extremely low and thus it usually displays low anticancer activity in *in-vivo* tests (Tang *et al.*, 2010). Several studies have demonstrated the therapeutic effects of curcumin (Aggarwal and Kuzhuvelil, 2009; Duvoix *et al.*, 2005; Phan *et al.*, 2001).

Curcumin-loaded Poly (lactic acid) (PLA) membranes were studied to be used as a drug-eluting stent. The anticoagulation assay conducted through the platelet adhesion test, enlightening that the anticoagulation behavior of the membranes was higher than the pure PLA membrane and the anticoagulation improved by increasing curcumin percent (Chen *et al.*, 2010).

The release of curcumin from PVA/curcumin composites and its effectiveness towards hepatocarcinoma were studied by El-Nashar *et al.* (2016). They concluded that the rate of release for curcumin depends on pH of the media and that (PVA/cur) composites have a hopeful result on human liver cancer cell HEPG<sub>2</sub>.

Novel hydrogel-silver nanoparticle-curcumin composites have been studied by Varaprasad *et al.* (2011) as wound dressing and antibacterial application. The results demonstrate that combining the natural compound curcumin enhances the antimicrobial effect and the composites have potential applications in the wound dressing or burn wounds.

PVA and Cs blend in different compositions with and without incorporation of curcumin have been prepared by Vidyalakshmi *et al.* (2004) and evaluated as drug delivery system. Mechanical properties of the membranes such as percentage elongation at break and tensile strength were evaluated.

The overall goal of this work is to study the result of loading curcumin with different concentrations (0, 5, 10, and 15 wt%) on the dielectric and biological properties of PVA, chitosan and PVA/Cs blend (50:50) to be used for medical applications, especially for wound healing.

# MATERIALS AND METHODS

#### Materials

Chitosan: poly (D-glucosamine), beta-(1, 4)-2-Amino-2-deoxy-D-glucose  $(C_6H_{11}NO_4)_n$  (degree of deacetylation (DD) = 75–92%). Houston, Texas, USA.

Polyvinylalcohol:  $(C_2H_4O)_n$  supplied from Qualikems fine chemicals Pvt.Ltd., New Delhi. Commercial Curcumin purchased from markets.

Gram-Negative Bacteria: (Escherichia coli (ATCC25922), Salmonella enterica (ATCC 25566), Klebsiella pneumonia (ATCC 10031) and Psedomonas aeruginosa (ATCC 27853). Gram Positive Bacteria: (Bacillus subtilis (NRRL-B-4219), Staphylococcus aureus (ATCC 6538) and Streptococcus pyogenes (ATCC19615). Yeasts: (Candida albicans (ATCC10231).

HFB4 (Human normal melanocyte) cell line was purchased from Vacsera (Giza, Egypt).

# Methods

#### Synthesis of nano curcumin

Curcumin solution was prepared by dissolving curcumin powder in ethanol. Under ultrasonication condition, this solution was added to boiling water in drop-wise mode. The solution was sonicated for about 30 min. After which, the mixture was stirred at 800 rpm for about 20 min till we obtained the orange colored precipitate. Afterward, the supernatant was removed and the obtained pellet was used for the study.

#### Preparation of curcumin nanocomposites

The PVA was dissolved in aqueous solution by using tetrahydrofuran (THF), heated in a water bath and stirred for 30 min. chitosan dissolved in 2% aqueous acetic acid solution at room temperature with stirring to prepare chitosan solution.

Curcumin-loaded PVA, CS, and PVA/Cs blend (50:50) nanocomposites were prepared. Curcumin of various concentrations, i.e., 0, 5, 10, and 15 wt% were dissolved in ethanol with PVA,  $C_s$ , and PVA/Cs (50/50). The resulting solution was stirred carefully few hours until a homogenous yellow color solution is obtained. Finally, the solution was poured into glass Petri dish and left to evaporate the solvent at room temperature overnight.

# Transmission electron microscope (TEM)

The particle size of the nano-curcumin powder was determined using Transmission electron microscope (TEM) model: JEM-HR 2100 and accelerating voltage 200 kV (Japan).

# Scanning electron microscope (SEM)

The homogeneity of the samples and distribution of the curcumin powder in the polymer matrix was investigated by Scanning electron microscope, SEM/EDX, Philips XL30 (Japan).

# Fourier transform infrared (FTIR)

The infrared spectrum was recorded by a JASCO FT/ IR 300 E Fourier transform infrared (FTIR) Spectrometer (Tokyo, Japan).

#### Dielectric measurements

An LCR meter type AG-411 B (Ando electric, Japan) was used to measure the dielectric properties in the frequency range 100 Hz up to 100 kHz at room temperature (25°C). The bridge directly measured the capacitance C, loss tangent tan  $\delta$  and ac resistance ( $R_{ac}$ ) and from those parameters, the permittivity  $\epsilon$ , dielectric loss  $\epsilon$ ", and  $R_{dc}$  were calculated. A guard ring capacitor type NFM/5T Wiss Tech. Werkstatten (WTW) GMBH Germany was used as a measuring cell. Standard materials used to calibrate the cell and the experimental error in  $\epsilon$  and  $\epsilon$ " were found to be  $\pm 3\%$  and  $\pm 5\%$ , respectively.

#### Water vapor transmission rate (WVTR)

The water vapor transmission rate (WVTR) was measured according to the monograph of the European pharmacopeia (Razzak and Darwis, 2001). Glass bottles were filled with 20 ml water. Circular shape samples with thickness 0.75 mm adhered to the bottles as a cap as seen in Figure 1. The bottles were weighed  $(W_i)$  and kept at 37°C in an incubator for 24 hours. After 24 hours the bottles are weighed  $(W_i)$  and the WVTR was calculated from:

$$WVTR(g/m2/h) = \frac{Wi \cdot Wf}{24A} \times 10^6 \qquad 1$$

where A is the surface area of the circular sample in (mm<sup>2</sup>).



Fig. 1: Experiment for WVTR.

#### Cell viability assay

HFB4 (Human normal melanocyte) was preserved in Dulbecco's Modified Eagle's Medium (DMEM). The medium was supplemented with 10% fetal bovine serum and incubated at 37°C in 5% CO<sub>2</sub> and 95% humidity. The cell line was sub-cultured using trypsin 0.15%. After 24 h of seeding 10000 cells per well (in 96 well plates), the medium was changed to serum-free medium containing a final concentration of the tested samples of 100  $\mu$ g/ml in triplicates. The cell was treated for 48 h. 100  $\mu$ g/ml doxorubicin was used as positive control and 0.5% DMSO was used as negative control. Cell viability was determined using the MTT (3-(4, 5-dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium bromide) assay (Mosmann, 1983). The equation used for calculation of percentage cytotoxicity:

percentage cytotoxicity = 
$$(av(x)/(av(NC))) \times 100$$
, 2.

where Av: average, X: absorbance of the sample well measured at 595 nm with reference 690 nm, NC: absorbance of negative control measured at 595 nm with reference 690.

#### Antimicrobial assay

Composites loaded with 0 and 10 wt% cur were subjected to microbiology study using a disk diffusion methodology (BSAC Disk Diffusion Method for Antimicrobial Susceptibility Testing, Version 4, 2005). Isosensitest agar (Oxoid, Basingstoke, UK). The sample code as follow:

1	2	3	4	5	6
PVA	PVA/cur	Cs	Cs/cur	PVA/Cs	PVA/Cs/cur

The tested samples placed onto the inoculated plates and incubated overnight at 37°C. The diameters of the inhibition zones formed around the disks were measured. Levofloxacin antibiotic discs with potency (5 mcg) were used as a standard antibiotic (Rahman *et al.*, 2007; Andrews, 2002).

#### **RESULTS AND DISCUSSION**

#### Transmission electron microscope (TEM)

Figure 2 shows the TEM image of nano-curcumin particles. The micrograph shows spherical ultrafine particles in the nano grade with diameter ranges from 2-4 nm.



Fig. 2: TEM of Nano-Curcumin.

#### Scanning electron microscope (SEM)

The morphology of the PVA, Cs and PVA/Cs blend and their cur nanocomposites with different concentrations were studied using SEM technique and only PVA/Cs blend and 10% cur loaded samples presented in Figure 3 for the sake of brevity.

The surface morphology of pure PVA, Cs and PVA/Cs samples are found to be smooth and homogenous. The addition of nanocurcumin particles causes changes in the surface morphology of such polymers. The curcumin nanoparticles are homogeneously distributed in the polymer matrix with some aggregations at the higher concentration.

# Fourier transform infrared spectroscopy (FTIR)

To investigate the interaction between nano curcumin with PVA, Cs and PVA/Cs blend, FTIR spectra were measured and recorded in the spectral range from 400-4000 cm<sup>-1</sup> and represented in Figures 4-8 and the band assignments of pure PVA and chitosan in Table 1. The characteristic bands of Cs and PVA in this study are in agreement with that reported before in the literature (Kamel *et al.*, 2017; Wang *et al.*, 2004; El-Hefian *et al.*, 2010).

The FTIR spectrum of the PVA/Cs blend (Fig. 4) is different from that of the pure homopolymers. By comparing the spectra of pure Cs with the blend, the absorption peak at 3520 cm<sup>-1</sup> characteristics for –OH and –NH stretching vibrations shifted to a 3620 cm<sup>-1</sup> in the blend. The band at 1595 cm<sup>-1</sup> in the pure chitosan spectrum characteristic for Amide II, N-H bending vibration and the symmetric-NH<sub>3</sub><sup>+</sup> deformation is shifted to 1582 cm<sup>-1</sup> with an increase in the intensity. These results reflect the chemical interaction between the two polymers. A similar observation was mentioned in previous work on PVA/Cs studies (El-Hefian *et al.*, 2010).

The chemical formula for curcumin is  $C_{21}H_{20}O_6$ . The chemical structure and FTIR spectrum are given in Scheme 1 and Figure 5, respectively.

Ν	Main IR band assignments of PVA	Main IR band assignments of Cs		
Wave number cm <sup>-1</sup>	ave number cm <sup>-1</sup> Band assignments		Band assignments	
3600	OH stretching vibrations	3503	O-H Stretching vibration and amine N-H bonds	
2934	CH stretching vibration of alkyl groups	2898	C-H vibration	
1747	C=O stretching vibration of the carbonyl group	1660	Amide I and antisymmetric- $NH_3^+$ deformation	
1640	acetyl C=O group	1585	Amide II, N-H bending vibration and the symmetric- $NH_3^+$ deformation	
1470	The corresponding bending of $\mathrm{CH}_{\mathrm{2}}$ vibrations	1434	C-H and O-H deformation vibrations	
1370	The corresponding wagging of $\mathrm{CH}_{_2}$ vibrations	1277	C-OH stretching	
1146	The C-C and C-O stretching vibration band	1169 1115	CO stretching and OH deformation vibrations of the primary and secondary alcohol groups in the chitosan	
1046	C-O stretching			



Fig. 3: SEM of (a) PVA/cur 10% (b) Cs/cur 10% (c) PVA/Cs and (d) PVA/Cs/cur 10%.



Fig. 4: FTIR spectra for PVA, Cs, and PVA/Cs blend.

As illustrated in scheme 1, the curcumin structure has 3 chemical entities: 2 aromatic rings containing o-methoxy phenolic groups, linked with 7 carbon linker consisting of an  $\alpha$ , $\beta$ unsaturated  $\beta$ -diketone moiety (Priyadarsini and Kavirayani 2014; Zhao *et al.*, 2015).

Figure 5 shows the FTIR spectrum of curcumin. From the figure, a broadband appears at 3417 cm<sup>-1</sup> attributed to the phenolic O–H stretching, 2 sharp peaks at 2920 and 2852.2 cm<sup>-1</sup> attributed to CH Stretching, 1706 cm<sup>-1</sup> for C=O, 1641 cm<sup>-1</sup> for C=C stretching, 1464 for v (OH), 1105 for C-O-C stretching modes, 723 for cis CH vibration of aromatic ring.

Figure 6 represents the FTIR spectra of PVA/cur nanocomposites with different concentrations of curcumin (0, 5, 10, and 15 wt%). From the figure, it can be noticed that a new band appeared at 1660 cm<sup>-1</sup> as a result of curcumin (stretching vibration peak of the carbon-carbon double bond in curcumin and

Table 1: Main IR band assignments of PVA and Cs.

the intensity of this band increases by increasing the curcumin concentration. It is noticed also increasing in the intensities of the bands at 2963. 2173, 1773, 1471, and 1145 cm<sup>-1</sup> after addition of curcumin. From the results of the FTIR, it appears that PVA and curcumin present in the nanocomposite without chemical interaction between them.



Scheme 1: Structure of curcumin.

For Cs/cur nanocomposites (Figure 7), the 1662 cm<sup>-1</sup> band vibration characteristic for the Amide I and antisymmetric- $NH_3^+$  of chitosan was shifted to 1672 cm<sup>-1</sup>. The band at 2898 cm<sup>-1</sup> is increased up to 10% cur. Increased intensity of 1595, 1440 and 1172 cm<sup>-1</sup> bands also noticed with increasing cur%. The resulted spectra may indicate the presence of interaction between the positively charged amino group of chitosan and negatively charged hydroxide group of curcumin (Das *et al.*, 2010).

Figure 8 shows the FTIR spectra of PVA/Cs/cur nanocomposites with different concentrations of curcumin. The curcumin loaded PVA/Cs blend (Figure 8) has shown all the characteristic bands of the pure blend with a slight shift of the bands at 1160, 1582, and 1671 cm<sup>-1</sup>. The disappearance of the band at 1582 cm<sup>-1</sup> from the spectrum of 5 and 10% cur loaded chit/PVA blend. The stretching vibration at 3627 cm<sup>-1</sup> corresponding to OH/ NH<sub>2</sub> groups has shifted to 3649 cm<sup>-1</sup>, indicating that the curcumin has been bonded to the functional groups of the chitosan and PVA.



Fig. 5: FTIR spectrum of curcumin.

#### **Dielectric properties**

The permittivity  $\varepsilon'$  and dielectric loss  $\varepsilon''$  for PVA, Cs, and PVA/Cs blend loaded with different concentrations of curcumin versus the applied frequency f, at room temperature 25°C are illustrated graphically in Figure 9. It is notable that the permittivity  $\varepsilon'$  for the whole samples shows a relatively high value at low-frequency range, decreased with increasing frequency and almost remained independent of the applied external field at the high-frequency range. It may be accomplished that; at a lower frequency, the dipoles within the system reorient themselves to retort to the applied electrical field. Whereas at a higher frequency,

the decrease of permittivity values was attributed to the lag of molecules towards the applied electric field and this might be explained by the issue of dipoles orientation during this frequency (Srivastava *et al.*, 2014). Moreover, it had been additionally observed that the values of  $\varepsilon'$  of PVA/cur nanocomposites greatly increase with increasing curcumin content which was found to be higher than those of the two individuals. This finding may be due to some sort of humidity of cur itself. On the opposite side, the permittivity decreases with increasing curcumin content in case of CS/cur and PVA/CS/cur nanocomposites as a result of the fact that the value of  $\varepsilon'$  for cur is about 8.5 (Khan, 2014) while it is 6.9 for PVA and 33.3 for Cs at f = 100 Hz.



Fig. 6: FTIR spectra of PVA/cur nanocomposites with different concentrations of curcumin (a) 0%, (b) 5 wt%, (c) 10 wt%, and (d) 15 wt%.

The high permittivity of CS/cur and PVA/Cs/cur may be due to the polar acetate and  $NH_3^+$  ions forming during the preparation of films by dissolving chitosan in 2% acetic acid solution.

The increase in permittivity of the matter could be related to the various forms of polarization. The well-known types of polarizations are electronic polarization, atomic polarization as well as orientation polarization attributed to the orientation of dipoles parallel to the applied electric field (Johns and Charoen, 2011). In addition, in the case of heterogeneous materials, a chance for interfacial polarization to be appears as a result of the distinction in conductivities of the phases. According to the type of polarization, the time needed to reach equilibrium level varies. In our case, which is considered to be a polar molecule, at low frequencies all the four types polarization contribute the permittivity leading to high permittivity, at lower frequencies. By increasing the frequency, both interfacial polarization and the orientation polarization contributions vanished which is the cause of the reduction of the permittivity at higher frequencies. The orientation polarization requires a longer time to reach static field value incomparable with both electronic and atomic polarization. Therefore, the orientation polarization decreases with increase in frequency compared to electronic and atomic polarizations at lower frequency region. At lower frequencies, the surface polarization generally happens at abundant. Anyhow the permittivity  $\varepsilon'$  was found to follow the order Cs > PVA/Cs/cur > PVA nanocomposites.



**Fig. 7:** FTIR spectra of Cs/cur nanocomposites with different concentrations of curcumin (a) 0%, (b) 5 wt%, (c) 10 wt%, and (d) 15 wt%.



Fig. 8: FTIR spectra of PVA/Cs/cur nanocomposites with different concentrations of cur (a) 0%, (b) 5%, (c) 10%, and (d) 15%.



Fig. 9: The permittivity  $\varepsilon'$  and dielectric loss  $\varepsilon''$  for cur loaded PVA, Cs and PVA/Cs with different conc. versus the applied frequency f, at room temperature 25°C.

As a result of the inability of polarization mechanism of the molecules to follow the change of the oscillating of the electric field, the dielectric loss  $\varepsilon''$  was formed. This type of the relaxation process in a composite is defined as the time taken for the dipoles to come back to its original random orientation. It doesn't take place instantaneously but the polarization tailing exponentially. Minimum loss obtained if the relaxation time is smaller or similar to the rate of the oscillating electric field. On the other hand, the polarization cannot follow the oscillating frequency leading to the energy absorption and or dissipated as heat in case of faster electric field oscillation than the relaxation time (Lovell *et al.*, 1993). The values of  $\varepsilon''$  obtained for the nanocomposites under investigation is found to follow the order Cs > PVA/Cs > PVA. Also, it is seen that the values of  $\varepsilon''$  are higher at low-frequency range beside the fact that  $\varepsilon''$  versus applied frequency curves are much complicated and broader than one Debye process. This finding reflects more than one relaxation process is expected to happen in such complicated nanocomposites (Kamel *et al.*, 2015; Motawie *et al.*, 2016).

Analyses of these curves were done by using Fröhlich and Havriliak-Nagami functions as mentioned elsewhere (Abd-El-Messieh and Abd-El-Nour 2003). Dielectric spectroscopy is a strong tool for studying the dynamics of polymers in an exceedingly wide range frequency. The various regimes of the spectra may be detected and therefore the dynamics of the primary and secondary relaxations were obtained.



Fig. 10: Example of the analyses of  $\varepsilon''$  data for 10% cur loaded PVA/Cs Fitting the experimental data after subtraction of the losses due to dc conductivity with Fröhlich and Havriliak Negami functions.

Example of the analysis is illustrated graphically in Figure 10. From this figure, an absorption region obtained at lower frequency range was fitted by Fröhlich function with distribution parameter p = 3 and relaxation time  $\tau_1$  about 0.003 s. These values are found to be independent on either the type of polymer or curcumin content in the composite. This region could be attributed to Maxwell-Wagner effect which is expected to be at the lower frequency range as a result of the multi-constituents of the investigated systems. The second absorption region,  $\tau_2$  which was fitted by Fröhlich distribution could be due to some local molecular motions rather than the main chain motion as it is expected to be frozen since the measurements were carried out at 25°C i.e. lower than the glass transition Tg of the polymers under investigations (Abou-Aiad *et al.*, 2006). The relaxation time  $\tau_2$  associated with this region is highly affected by the type of polymer and the percentage of curcumin and follows the order Cs > PVA/Cs > PVA.

The obtained data of  $\tau_2$  for all nanocomposites under investigations are plotted graphically in Figure 11. From this figure, it is clear that  $\tau_2$  increases in the same order CS > /PVA/ Cs > PVA but  $\tau_2$  values for PVA/Cs composites are much closer to those for Cs rather than PVA indicating a nonlinear relationship. As an example, Figure 11b represents the variation of PVA content versus  $\tau_2$  for PVA/Cs/cur 10%. From this figure, it is clear that the deviation from the line connecting the two individuals was a positive deviation. This finding reflects an interaction between the two polymers which justify the results obtained before from FTIR spectroscopy. This interaction increases the molar volume of the rotating units and then the relaxation time.



Fig. 11: Relaxation time  $\tau_2$  versus (a) cur content and (b) PVA content for 10% cur loaded PVA/Cs.

For evaluation of the insulating materials, the conductivity studies are so important parameter. This is due to the fact that the main objective for an insulator material is to test its capability to oppose the leakage of electric current (Johns and Charoen, 2011). The electrical conductivity  $\sigma$  was calculated from measured resistance and illustrated graphically in Figure 12. From this figure, it is found that Cs/cur nanocomposites possess the highest  $\sigma$  values while PVA/cur are the lowest ones. As seen in figure 12 the  $\sigma$  values are found to decrease by increasing curcumin content in both C<sub>s</sub> and PVA/CS nanocomposites. While on the other hand, they increase by increasing cur content in PVA/

cur nanocomposites. Anyhow, the electrical conductivity of both Cs (Martins *et al.*, 2014) and PVA (El-Houssiny *et al.*, 2012) are found to be comparable with those found in the literature.

However, the obtained electrical conductivity for all nanocomposites under investigation lies in the order of the antistatic range  $10^{-9}$ – $10^{-14}$ /S cm<sup>-1</sup> (Huang, 2002) which recommend being used for antistatic applications such as skin contact.

#### Water vapor transmission rate (WVTR)

WVTR is the ability of a dressing material to control water loss from the wound. Moderate WVTR is recommended to

offer a moist environment to accelerate healing. High WVTR may cause dryness of the wound, while low WVTR may cause storing of wound exudates which may result in retardation of the wound healing (Xu *et al.*, 2016). The values of the WVTR for normal skin have been reported to be in the range 8.5 g/m<sup>2</sup>/h to 11.6 g/

 $m^2/h$  (Gonzalez *et al.*, 2012). The WVTR values for cur loaded PVA, Cs and PVA/Cs blend are shown in Table 2. From this table, it is obvious that addition of cur decreases the WVTR of the composites. WVTR values follow the order PVA > Cs > PVA/Cs.



Fig. 12: Electrical dc conductivity versus cur content.



Fig. 13: Zone of inhibition of 0 and 10% loaded cur PVA, Cs and PVA/Cs, PVA/Cs blend against various pathogenic microorganisms.

#### Cell viability assay

The cell viability effect of pure and 10% cur loaded PVA, Cs and PVA/Cs blend against the human normal melanocyte cell line (HFB4) was investigated and presented in Table 3. Cultures of the cell line were treated with extracts first at one concentration of 100  $\mu$ g/ml and the results indicated the all samples showed cell viability greater than 80% which confirm that addition of cur has no toxic effect on the cells.

#### Antimicrobial assay

Figure 13 represents the Zone of inhibition of pure and 10% cur loaded nanocomposites against 8 pathogenic microorganisms and the Inhibition zone diameter (millimetre) of the pure and 10% cur loaded PVA, Cs and PVA/Cs nanocomposites is represented in Table 4. The results illustrated that the samples PVA, Cs, Cs/cur 10% have very weak antimicrobial effect on all tested standard microorganisms, while the sample PVA/Cs/ cur 10% has moderate antimicrobial activity showed on some gram negative bacteria (Klebsiella pneumonia) & gram positive bacteria (Streptococcus pyogenes) with inhibition zone diameter (11 mm), also the PVA/Cs blend showed low to moderate on all tested bacterial either gram positive or gram negative bacteria with inhibition zone diameter ranged (7-14 mm). On the other hand there is no antimicrobial effect on yeasts (Candida albicans) by all tested samples except the sample PVA/cur 10% which showed moderate antimicrobial effect with inhibition zone diameter (19 mm), moreover, the sample PVA/cur 10% was the best one which showed moderate to strong antimicrobial effect on all tested standard microorganisms especially on Klebsiella pneumonia which is pathogen that mainly involved in many infections and can be linked with a high rate of mortality. Previous work on curcumin confirmed its efficiency against this pathogen (Gunes et al., 2016).

When chitosan and PVA/Cs blend are loaded with curcumin, they have chemically interacted as supposed by FTIR and dielectric results. This interaction may lead to limit the inhibitory effect of curcumin as observed in the antimicrobial test. On the other hand no or few interactions between PVA and curcumin so the active compounds of curcumin are free to inhibit microorganisms in the antimicrobial test and this may be the interpretation of the increased antimicrobial efficiency of the PVA/cur than cs/cur nanocomposites.

 Table 2: Water vapor transmission values for cur loaded PVA, Cs and PVA/Cs blend.

6l.	WVTR values g/m <sup>2</sup> /h					
Sample	0% cur	5% cur	10% cur	15% cur		
PVA	37.81	35.82	30.78	24.42		
Cs	19.90	17.78	16.80	16.72		
PVA/Cs blend	11.94	9.29	9.28	8.625		

**Table 3:** Cell viability of pure and 10 wt% cur loaded PVA, Cs and PVA/Cs blend (100 µg/ml) on human normal melanocyte cell line (HFB4).

Comula	Cell viability%			
Sample	HFB4 (human Normal melanocyte)			
PVA	91.3			
PVA/cur 10%	88.6			
Cs	91			
Cs/cur 10%	91.1			
PVA/Cs	84.1			
PVA/Cs/cur 10%	83.5			

Table 4: Inhibition zone diameter (millimeter) of the pure and 10% cur loaded PVA, Cs and PVA/Cs nanocomposites.

i	1	2	3	4	5	6	I I787 ++
Microorganism	PVA	PVA/cur	Cs	Cs/cur	PVA/Cs	PVA/Cs/cur	LEV.
E. coli	Nil*	12.0	Nil	Nil	8.0	Nil	24.0
Sal. enterica	Nil	24.0	Nil	Nil	10.0	Nil	38.0
K. neumoniae	Nil	26.0	Nil	Nil	9.0	11.0	27.0
P. aeruginosa	Nil	18.0	Nil	Nil	14.0	Nil	30.0
Str. pyogenes	Nil	20.0	Nil	Nil	11.0	11.0	24.0
Staph. aureus	Nil	21.0	Nil	Nil	9.0	Nil	26.0
B. subtilis	Nil	15.0	Nil	Nil	7.0	Nil	36.0
Can. albicans	Nil	15.0	Nil	Nil	Nil	Nil	32.0

\*Nil: No antimicrobial activity recorded.

\*\*LEV: Levofloxacin, used as a standard antibiotic.

# CONCLUSION

PVA, chitosan and their 50:50 blend nanocomposite with and without curcumin have been prepared by solution casting method using different concentrations of curcumin (0, 5, 10 and 15) wt%. Results of the FTIR suggest an interaction between the nanocurcumin and chitosan and also between the curcumin and the PVA/Cs blend. Results of the SEM indicate good dispersion of the curcumin within the polymers matrix. Cur loaded PVA have the highest WVTR than chitosan and PVA/Cs blend. The Low WVTR induced by cur loading suggests that these types of dressings are more suitable for nonexudative or dry wounds. The cell viability data indicated that all samples showed cell viability greater than 80% which confirmed that the addition of curcumin has no cytotoxic effect on human normal melanocytes. PVA/cur nanocomposites are the best which showed moderate to strong antimicrobial effect on all tested standard microorganisms. Both  $\varepsilon'$ and  $\varepsilon''$  values follow the order CS > /PVA/Cs > PVA. The obtained

electrical conductivity for all composites under investigation lies in the order of the antistatic range  $(10^{-9}-10^{-14}/\text{S cm}^{-1})$  which recommend such composites to be used for antistatic applications. The investigation indicates that the PVA/cur nanocomposites have high potential as wound dressing materials and *in-vivo* study for these materials are recommended.

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