Antioxidant enzyme activity and lipid peroxidation in rat liver exposed to celecoxib and lansoprazole under epinephrine-induced stress

Bohdan Melekh\textsuperscript{1,2}, Iryna Ilkiv\textsuperscript{3}, Andrii Lozynskyi\textsuperscript{4*}, Alexander Sklyarov\textsuperscript{3}

\textsuperscript{1}Department of Pathophysiology, Danylo Halytsky Lviv National Medical University, 69 Pekarska, Lviv, 79010, Ukraine.
\textsuperscript{2}St. Mary's Hospital, 6-8 Marienstraße, Vechta, 49377, Germany.
\textsuperscript{3}Department of Biochemistry, Danylo Halytsky Lviv National Medical University, 69 Pekarska, Lviv, 79010, Ukraine.
\textsuperscript{4}Department of Pharmaceutical, Organic and Bioorganic Chemistry, Danylo Halytsky Lviv National Medical University, 69 Pekarska, Lviv, Ukraine.

**ARTICLE INFO**

**Article history:**
Received on: 22/02/2017
Accepted on: 25/05/2017
Available online: 30/10/2017

**Key words:**
Celecoxib, lansoprazole, oxidative stress, liver.

**ABSTRACT**

The effect of exposure of celecoxib and lansoprazole on antioxidant enzymes and lipid peroxidation has been studied in liver of rats. We considered the content of malondialdehyde (MDA), and activity of superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx), and glutathione reductase (GR). MDA content was significantly decreased for 35% in rats treated with celecoxib and for 72% in combination with lansoprazole. The increase of SOD activity after administration of celecoxib in the liver was 29% and observed decrease of CAT (12%), GPx (5%) and GR (33%) activities. Similarly, an increase in the antioxidant enzymes was found after administration of celecoxib and lansoprazole such as increase of SOD activity (44%) and decrease of CAT (16%), GPx (9%) and GR activities (46%). The current study shows that exposure of celecoxib and lansoprazole increases the endogenous antioxidant defense system and decreases the lipid peroxidation in rat liver tissues under epinephrine-induced stress and confirms the important role of these drugs in the reduction of stress-induced cellular toxicity.

**INTRODUCTION**

In a healthy human body an effective balance exists between the antioxidant and prooxidant system and the ability of cells to protect themselves from oxidant damage. Disruption of this balance has been attributed to several factors including age, genetics, diet, individual behavior, stress and external stressors (Bagchi \textit{et al.}, 1998; Söderholm and Perdue, 2001). It has been also considered that the lipid peroxidation damage is involving in aging and pathological disorders. Some phases of ulcerative colitis, atherosclerosis, neuronal ceroid lipofuscinosis, Alzheimer's disease, liver injury and oxygen toxicity have been discussed in relation to lipid peroxidation (İnal \textit{et al.}, 2001). On the other hand, lipid peroxidation in cells and tissues plays an important role in the biosynthesis of prostaglandin and in the mechanisms of immune response especially during peroxynitrite, glutathionyl and protein-tyrosyl radicals derived from macrophages (de Menezes and Augusto, 2001; Costantini and Møller, 2009).

In addition, it has been proposed that lipid peroxidation and antioxidant enzyme activity play a key role in inflammation development (Wiseman and Halliwell, 1996). Thus, the inflammatory process activates a series of receptors and transcription factors such as c-Jun amino terminal kinase, toll-like receptors, nuclear factor-B and the receptor for advanced glycation end products, which lead to β-cell dysfunction and apoptosis (Goldberg, 2009). In addition, the inflammation involved in the releasing of chemokines such as macrophage migration inhibition factor (MIF), monocyte chemoattractant protein (MCP)-1 and others from stressed tissues.
Moreover, this mediators released by stressed cells have effects on proinflammatory gene activation and chemokine–induced proliferation producing cytokines such interferon-γ, TNF-α, IL-1, IL-6, IL-18, and others. The underlying factors that stimulate expression of these genes include oxidized lipids, reactive oxygen species (ROS), reactive nitrogen species (RNS) and advanced glycation end products (AGEs) (Goldberg, 2009).

Wide spread problems, such as wounds, menstrual pain and inflammation, are nowadays treated by non-steroidal anti-inflammatory drugs (NSAIDs) which inhibit the activity of cyclooxygenases (Luong, 1996). However, the non-steroidal anti-inflammatory drug (NSAID) therapy in treatment of various diseases has great benefits, but these benefits are offset by the occurrence of potentially complications related to NSAID-induced gastrointestinal injury (Graham et al., 2005). Thus, the traditional NSAIDs such acetylsalicylic acid, indomethacin and ketoprofen inhibiting both cyclooxygenase isoenzymes (COX-1 and COX-2) in the gastrointestinal tract and reduce the intrinsic ability of the gastric mucosa to resist injury induced by endogenous and exogenous agents (Fiorucci et al., 2005). The identification and characterization of the COX-2 enzyme inhibitors which are considered from experimental models supporting the concept that selective COX-2 inhibitors spare the gastrointestinal tract. Thus, these new generation of drugs have been shown to be effective in reducing inflammation while causing significantly less gastrointestinal damage than standard NSAIDs (Reuter et al., 1996; Ilkiv et al., 2007). Released in 1998, celecoxib was the first specific inhibitor of COX-2 available for clinical use. In the treatment of patients with arthritis, pain, menstrual cramps, and colonic polyps, therapeutic doses of celecoxib have proven to be equi-efficacious when compared with other traditional NSAIDs.

For patients with rheumatoid arthritis and heart failure, celecoxib is one of the most commonly prescribed drugs in the U.S. (Davies et al., 2000). It should be noted that the action of COX-2–selective NSAIDs are similar to that of a non-selective NSAID plus a proton-pump inhibitor for patients at high risk for ulcer complications. Lansoprazole is one example of a proton pump inhibitor that is effective for treating various acid-related diseases and shows greater efficacy than histamine-2 receptor antagonists in terms of complete symptom resolution and ulcer healings.

Also not only using different non-steroidal anti-inflammatory drugs, but emotional stress and external stressors lead to gastrointestinal damage. Therefore, stress models such as epinephrine-induced is widely used to induce gastric mucosal lesions in animals. Thus, this model reproduce systemic and local consequences of stress exposure to the upper gastrointestinal (GI) tract, resulting in the formation of bleeding gastric lesions and a decrease in mucosal microcirculation. These effects could be caused by gastric constriction of the blood vessels, leading to hypoxia, ischemia, oxidative stress and thereby leading to formation of gastric injury (Fomenko et al., 2014). In addition, the epinephrine administration caused in liver tissues the changes between pro-oxidant production and antioxidant defence system in response to hepatic oxidative stress (Polavarapu et al., 1998). In view of these considerations, the aim of the present study was to evaluate if acute epinephrine-induced stress model in rats modifies the antioxidant defense system and induces LPO in liver in the presence of celecoxib as selective COX-2 inhibitor and lansoprazole as proton-pump inhibitor, in order to determine the involvement of ROS formation on epinephrine-induced stress. To this end, GSH-Px, GR, SOD and CAT activities, as well as MDA levels, as measurement of LPO, were measured in liver homogenates from rats treated with COX-2 and proton-pump inhibitors under epinephrine-induced stress.

MATERIAL AND METHODS

Animals

All experiments were carried out using male albino rats weighing 180–220 g in accordance with the norms of the European Convention for the Protection of Vertebrate Animals Used for Experimental and Other Scientific Purposes (1986), as well as the Ethical Committee of Lviv National Medical University. The rats were fed standard chow and water ad libitum, and were housed in room with controlled temperature (22 ± 1 °C), humidity (65–70%) and light cycle (12 h light/dark). The rats were deprived of food for 18 h before the experiment, but had free access to water. Six rats were used in each group.

Study protocol

The study comprised of the following series of experiments: 1) intact animals were used as controls; 2) epinephrine-induced stress in rats were induced by intraperitoneal injection of epinephrine in dose of 2 mg/kg daily during 5 days (Martinez-Augustin et al., 2000); 3) selective COX-2 inhibitor, celecoxib (Pfizer, USA) was introduced in a dose of 10 mg/kg in rats on the background of epinephrine-induced stress during two weeks (n = 6); 4) experimental group, animals received lansoprazole (Pharma Life, Ukraine) in dose 30 mg/kg intragastrically (via an orally introduced polyethylene tube) once daily per 30 min after administration celecoxib (10 mg/kg) on the background of epinephrine-induced stress during two weeks (n = 6). Under general anesthesia, rats were sacrificed by decapitation and immediately subjected to necropsy. Livers were quickly removed, immediately washing out the blood with ice-cold 0.9% saline solution, weighted and stored at -70 °C. Homogenates of the tissues were prepared as 1.0 g/10 mL in 250 mM sucrose, 1 mM EDTA, 1 mM DL-dithiothreitol and 15 mM Tris HCl (pH 7.4), using HS-30E homogenizer (WiseStir, Republic of Korea). Each homogenate was centrifuged for 30 min at 800 g. The resulting supernatant fraction was used to determine enzyme activities and MDA levels. The protein concentrations of the supernatant were determined by the method described by Bradford (1976).

Enzyme activities

SOD activity was measured using quercetin as the substrate after suitable dilution method as described by Kostyuk et al. (1990). The assay mixture in a total volume of 1 mL consisted
0.1 mol/L sodium phosphate buffer (pH 7.8) and 0.08 mmol/L EDTA at a 1:1 proportion. The 0.1 mL of tissue sample (1:1000) after dilution was added to 2.3 mL of distilled water, after which 1 mL of assay mixture with EDTA and sodium phosphate buffer. The increase in absorbance due to oxidation of quercetin at 0 and 20 min was measured spectrophotometrically at 406 nm. In the blank, tissue sample was substituted by equal quantities of distilled water. One unit of SOD activity is defined as the quantity of enzyme that inhibited quercetin oxidation by 50% under given experimental conditions.

CAT activity was assayed according to the method of Korolyuk et al. (1988). The reaction was started by the addition of 0.1 mL of tissue sample to 1 mL of 4% ammonium molybdate and 2 mL of 0.03% H2O2 solution. One unit of catalase activity is defined as the amount of enzyme required to clear 1 μmol of H2O2 per minute per gram of tissue. The breakdown of hydrogen peroxide in the reaction mixture was measured spectrophotometrically at 410 nm.

The activity of GPxs in the tissue homogenates was measured spectrophotometrically as described by Moin (1986). The assay mixture contained 0.8 mL of 0.1 mol/L Tris-HCl with 12 mmol/L sodium acetate and 6 mmol/L EDTA, pH 8.9, 0.2 mL of 1:50 or tissue homogenates, 0.1 mL of 0.01 mol/L 5,5-dithiobis-2-nitrobenzoic acid, 1 mL of 20 mmol/L t-butylhydroperoxide, and 0.1 mL of 4.8 mmol/L GSH. The decrease in the absorbance at 412 nm was followed spectrophotometrically. GR activity in the tissue homogenates was performed by the method of Glatzle et al. (1974). GR activity was determined spectrophotometrically by measuring NADPH oxidation at 340 nm. The reaction mixture contained tissue samples (0.05–0.2 mg), 0.1 mM NADPH, 0.5 mM EDTA, 0.1 M potassium phosphate (pH 7.5), 200 mM KCl. After 5 min of preincubation (37 °C), the reaction was initiated by addition of 1 mM GSSG.

Measurement of lipid peroxidation levels

Estimates of lipid peroxidation levels were evaluated by the thiobarbituric acid reactive substances (TBARS) procedure, described by Timirbalatov and Seleznev (1981) on homogenized tissues. This method involves reaction of TBA with the degradation product of lipid peroxidation, MDA, under conditions of high temperature and acidity to generate a colored adduct that is measured spectrophotometrically. Briefly, 2 mL of distilled water was added to 0.1 mL of tissue sample, which followed by 1 mL of TBA reagent and 1 mL of trichloracetic acid and, and the mixture heated in a boiling water bath for 10 min before the addition of butanol. After cooling, the mixture was centrifuged for 10 min. Absorbance in the organic phase was determined at 532 nm and samples were compared to a blank.

Statistical methods

Each experiment was performed in triplicate and average values were recorded. Results are expressed as the means ± SD. The data were evaluated statistically using Student’s t-test. In general, a value of p<0.05 was regarded to be statistically significant and marked with an asterisk.

RESULTS AND DISCUSSION

The present study demonstrates that the antioxidant defense systems are modified by celecoxib and lansoprazole administration, altering the activities of antioxidant enzymes, and as a consequence, suggesting the role of ROS in the pathogenesis of stress-induced damage. This is the first study concerning about the changes in the activities of antioxidant enzymes produced by administration of selective COX-2 and proton-pump inhibitors in the liver of rats. Thus, the liver has been described as the important organ involved in the intracellular redox homeostasis, due to the formation of various intracellular antioxidant enzymes responsible for ROS clearance and the function of the key systemic antioxidant agent, GSH/GSSG (Klaassen, 2001). In addition, the liver plays an important role in metabolism of xenobiotics, therefore, its oxidative status might be affected by the administration of various drugs. Our results indicate that most reactive biochemical products that enter the reactions with enzymes are MDA. They consist of reaction products with MDA and occur as finished products of lipoperoxidation. As result, the content of malondialdehyde (MDA) increased 3.5-fold as compared to control rats. The selective COX-2 inhibitor, celecoxib, elicited a decrease of MDA content for 35% and for 72% in combination with lansoprazole as compared with indices of stress group (Figure 1).

![Fig. 1: Effect of celecoxib and lansoprazole (10 and 30 mg/kg) on malondialdehyde (MDA) levels under epinephrine-induced stress in rat liver tissues. The results are expressed as mean ± SD for 6 rats per group; *p<0.05 in comparison of control group; #p<0.05 versus the indices of stress.](Image)

Simultaneously, we found interrelationships between administration of celecoxib and lansoprazole and the activity of the important antioxidant enzymes SOD, CAT, GPx, and GR and TBARS products in the liver of rats. The activities of SOD found in liver from experimental rats are shown in Figure 2.
Fig. 2: Effect of celecoxib and lansoprazole (10 and 30 mg/kg) on the activity of superoxide dismutase (SOD) under epinephrine-induced stress in rat liver tissues. The results are expressed as mean ± SD for 6 rats per group; *p<0.05 in comparison of control group; #p<0.05 versus the indices of stress.

The celecoxib administration increased SOD activity for 29% (p<0.01) and celecoxib + lansoprazole for 44% (p<0.01) as compared with stress group. The administration of celecoxib decreased in liver catalase activity for 12%. Indices of the catalase activity in celecoxib + lansoprazole pretreated rats subjected to epinephrine-induced stress were similar to those of the group treated with celecoxib (Figure 3).

Fig. 3: Effect of celecoxib and lansoprazole (10 and 30 mg/kg) on the activity of catalase (CAT) under epinephrine-induced stress in rat liver tissues. The results are expressed as mean ± SD for 6 rats per group; *p<0.05 in comparison of control group; #p<0.05 versus the indices of stress.

Administration of celecoxib under acute epinephrine-induced stress conditions, did not significantly decrease GPx activity. In the liver the percentages of GPx reduction were 5 and 9% in rats treated with celecoxib and celecoxib + lansoprazole, respectively, when compared with stress values (Figure 4).

Changes in GR activities, the enzyme involved in generation of GSH from GSSG, are shown in Figure 5. Liver glutathione reductase activity in the rats after administration of celecoxib was about 33% lower than stress group. In rats after administration celecoxib + lansoprazole liver glutathione reductase also decreased, but to a final value about 12% higher than that in intact rats.

Fig. 4: Effect of celecoxib and lansoprazole (10 and 30 mg/kg) on the activity of glutathione peroxidase (GPx) under epinephrine-induced stress in rat liver tissues. The results are expressed as mean ± SD for 6 rats per group; *p<0.05 in comparison of control group; #p<0.05 versus the indices of stress.

Our results show an increase of activity of SOD and decrease the activities of CAT GPx, GR in liver of rats administrated with selective COX-2 and proton-pump inhibitors when compared to stress group. These results suggest decreased
production of oxygen free radicals by celecoxib and lansoprazole. The endogenous scavenger, SOD, which removes the superoxide anion radicals by converting them into hydrogen peroxide (H₂O₂) and O₂, was significantly increased in the two experimental groups. The observed increase in liver SOD enzyme activity after administration of the tested drugs may be a consequence of oxidative activation of enzyme protein or increased of their synthesis. Therefore, the increase in the activity of SOD in liver tissues of celecoxib and lansoprazole treated rats might indicate a reduction accumulation of superoxide anion radical with oxidative stress, contributing decrease liver toxicity (Zelko et al., 2002). In addition, SOD enzymes in human cells work in conjunction with H₂O₂-removing enzymes such as CAT and GPx. Thus, the relative contributions of CAT and GPx, in decomposition of endogenously produced hydrogen peroxide, follow tissue specificity, especially in liver and renal tissues (Kaushik and Kaur, 2003). The decreased activities of GPx and CAT in liver tissues of the treated rats indicate the highly reduced capacity to scavenge hydrogen peroxide produced in these tissues, with an reduction of oxidative stress and generation of ROS in response to administration of selective COX-2 and proton-pump inhibitors (Escobar et al., 1996). The observed decrease in glutathione reductase activities after administration of celecoxib and lansoprazole indicates a decrease in the conversion of oxidized glutathione (GSSG) back to its reduced form (GSH), which indicate further reduction of ROS during oxidative challenge (Circu and Aw, 2010). In addition, the changes in glutathione reductase activities follow qualitatively a similar pattern to those of follow qualitatively a similar pattern to those of lipogenesis and of several enzymes involved in lipid biosynthesis, resulting the decrease of MDA levels and cellular free radical stress (Knapen et al., 1999).

**CONCLUSION**

Our results support the hypothesis that enzymatic free radical defense systems can be impaired in epinephrine-induced stress rats and that the resultant enhanced oxidative stress can contribute to pathological liver injury. However, the exposure of celecoxib and lansoprazole to epinephrine-induced stress results in an increase in the endogenous antioxidant defense system together with a decrement in lipid peroxidation in rat liver tissues, suggesting an important role of these drugs in the pathogenesis of stress-induced cellular toxicity.

**Financial support and sponsorship:** Nil.

**Conflict of Interests:** There are no conflicts of interest.

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How to cite this article: