

Chemical constituents and cytotoxic effect from the barks of *Goniothalamus chinensis* Merr. & Chun. growing in Vietnam

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ABSTRACT

Goniothalamus chinensis Merr. & Chun is a medicinal plant which has cytotoxicity, antitumor, antifungal, antimalaria and antituberculosis and antioxidant activity. The styryl-lactone, alkaloid and acetogenin are the main group compounds in species of *Goniothalamus*. In this study, we aimed to investigate phytochemical and cytotoxic effect of compounds from bark of *Goniothalamus chinensis* Merr. & Chun. The bark of *Goniothalamus chinensis* Merr. & Chun grown in Vietnam was extracted by methanol. Three compounds were isolated using on Sephadex LH-20 and preparative glass-backed TLC plates. The compound's structure were characterized on the basis of spectroscopic data and by comparing their physicochemical and spectral data with those published in literatures. The cytotoxicity of three compounds was evaluated against four cancer cell lines: epithelial cancer (KB), liver (Hep G2); breast cancer (MCF-7) and lung (LU-1). We have been isolated three compounds. Their structures were identified as Goniothalamin (**1**), Aristolactam BII (**2**), 3-Methyl-1H-benz[*f*]indole-4,9-dione (**3**). Isolated compounds were tested cytotoxic activity against different cancer cell lines. The compound **1** demonstrated the highest potency against the epithelial cancer cell line with an IC₅₀ of 4.18 ± 0.60 µg/mL. Other compounds have weak cytotoxic activity.

INTRODUCTION

With the tropical climate, Vietnam has diversity of plant and animals species. In total, Vietnam has more than 12000 plant species; of which nearly 4000 species can be used in traditional medicine. The genus of *Goniothalamus* contains about 160 species in the world, mostly in Southeast Asia, among which about 19 are distributed in Vietnam (Wiert, 2007). Some species have been used as folk medicines for healing, for digestion and as a tonic plant. Main compounds in *Goniothalamus* are styryl-lactone, alkaloid and acetogenin which have cytotoxic, antitumor, insecticides, antifungal, antimalaria, antituberculosis and antioxidant activities (Blázquez *et al.*, 1999). Orlikova *et al.*, have showed goniothalamine, a styryl-lactone isolated from

Goniothalamus macrophyllus, have cytotoxic and anti-inflammatory effects. This compound also inhibits the tumor necrosis factor- α -induced NF- κ B activation and have anti-leukemic potential (Orlikova *et al.*, 2013). Other study demonstrated that goniothalamine C, a styryl lactone, was extracted from *Goniothalamus cheliensis*, strongly inhibited platelet-derived growth factor-BB-induced vascular smooth muscle cell migration and proliferation (Sun *et al.*, 2014). Goniothalamine, an active compound extracted from *Goniothalamus griffithii*, also inhibits human leukemic cells undergo apoptosis via intrinsic and extrinsic pathways (Petsophonakul *et al.*, 2013) and cervical cancer cells (HeLa) (Alabsi *et al.*, 2012). Furthermore, goniothalamine also showed potent gastroprotective activity on gastric ulcers models in rats. The mechanism is goniothalamine may induce the production of sulfhydryl compounds and prostaglandins (Vendramini-Costa *et al.*, 2014). For now, species of *Goniothalamus* continue being interested by many researches. However, there has been no report about the studies on the chemical constituents and bioactivity of this plant growing in Vietnam.

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Therefore a study about the chemical composition of *Goniothalamus chinensis* Merr. & Chun is much needed. In this study, from barks of *Goniothalamus chinensis* Merr. & Chun grown in Vietnam, have extracted, isolated and identified three compounds: Goniothalamine (1), Aristolactam BII (2), 3-Methyl-1H-benz[f]indole-4,9-dione (3). Furthermore, we have evaluated the cytotoxic of these compounds on four cell line epithelial cancer (KB), liver (Hep G2); breast cancer (MCF-7) and lung (LU-1).

MATERIALS AND METHODS

Plant material

Barks of *Goniothalamus chinensis* Merr. & Chun. (Kingdom: Plantae; Phylum: Tracheophyta; Class: Magnoliopsida; Order: Magnoliales; Family: Annonaceae) were collected in Ha Giang province, Vietnam during 2013 and authenticated by the School of Medicine and Pharmacy, Vietnam National University, Hanoi, Vietnam (SMP-VNU). A voucher specimen (No. SMP-2013-0018) has been deposited at the Herbarium of SMP-VNU.

General experimental procedures

Melting points were measured on Mikroskopheiztisch PHMK-50 (VEB WaegetechnikRapido, Germany). The FT-IR spectra were recorded on an IMPACT-410FT-IR spectrometer (CARL ZEISS JENA). The NMR [^1H (500 MHz), ^{13}C (125 MHz), and DEPT-90 and 135 MHz] spectra were recorded on an AVANCE spectrometer AV 500 (Bruker, Germany) in the Institute of Chemistry, Vietnam Academy of Science and Technology (VAST). Chemical shifts were reported in ppm downfield from TMS with J in Hz. Electrospray Ionization Mass Spectra (ESI-MS) were recorded on a Varian Agilent 1100 LC-MSD mass spectrometer. Analytical TLC was performed on Kieselgel 60 F₂₅₄ (Merck) plates (silica gel, 0.25 mm layer thickness) and RP-18 F₂₅₄ (Merck) plates (0.25 mm layer thickness). Spots were visualized using ultraviolet radiation (at 254 and 365 nm) and by spraying with 10% H₂SO₄, followed by heating with a heat gun. Column chromatography was performed on silica gel (70–230 and 230–400 mesh, Merck). Organic solvents were of analytical grade.

Extraction and isolation

Goniothalamus chinensis Merr. & Chun barks of 1.7 kg was dried and extracted with methanol (5L × 24 hours × 3 times) at room temperature. The methanol extracts were combined and then evaporated to dryness *in vacuo* at 40°C to yield crude extract (40.3 g). This crude extract was chromatographed on Sephadex LH-20 column and eluted with gradient of CH₂Cl₂/MeOH to yield ten fractions F1-F10. Fraction F3 was further separated over a silica gel column and eluted with *n*-hexane/acetone (9:1) to yield eight subfractions F3.1-F3.8. Fraction 3.7 was further separated on Sephadex LH-20 column and eluted with MeOH/CH₂Cl₂ (95/5) to yield two fractions F3.7.1 and F3.7.2. Fraction F3.7.2 was applied on preparative glass-backed TLC plates, coated with silica gel 60 and eluted with gradient of *n*-hexane/acetone to yield compound 1

(14 mg). Fraction 3.8 was further separated on Sephadex LH-20 column and eluted with MeOH/CH₂Cl₂ (95/5) to yield compound 3 (17.5 mg). Fraction F4 was chromatographed over a silica gel column and eluted with *n*-hexane/acetone (9:1) to yield fifteen subfractions F4.1-F4.15. Fraction F4.13 was crystallized on solvents CH₂Cl₂-MeOH (1:1) to yield compound 2 (6.4mg).

Goniothalamine (1): was obtained as a light yellow, solid powder, melting point 82-84°C; $R_f = 0,44$ (*n*-hexane-CH₂Cl₂ 20/80, v/v); $[\alpha]_D^{27} = +81,0^\circ$ (C 0,2; EtOH). FT-IR (KBr) ν_{\max} (cm⁻¹): 2925, 1707, 1664, 1627, 1454, 1382, 1244, 1066, 966, 822, 759, 699, 584, 512, 440. UV λ_{\max} MeOH nm (log ϵ): 210 (4.43); 251 (4.32); 282 (3.25); 290 (3.03). (+)-ESI-MS: m/z 223 [M+Na]⁺. $^1\text{H-NMR}$ (500 MHz, CDCl₃): δ (ppm) 7.40 (2H, m, H-10, H-14); 7.34 (2H, m, H-11, H-13); 7.27 (1H, m, H-12); 6.92 (1H, ddd, $J=4.0; 5.0; 9.5$ Hz, H-4); 6.73 (1H, d, $J=16.0$ Hz, H-8); 6.27 (1H, dd, $J=6.5; 16.0$ Hz, H-7); 6.09 (1H, dt, $J=2.0; 10.0$ Hz, H-3); 5.10 (1H, ddd, $J=1.0; 6.5; 15.5$ Hz, H-6); 2.54 (2H, m, H-5). $^{13}\text{C-NMR}$ (125 MHz, CDCl₃): δ (ppm) 163.8 (C-2); 144.5 (C-4); 135.8 (C-9); 133.1 (C-8); 128.7 (C-11, C-13); 128.3 (C-12); 126.7 (C-10, C-14'); 125.7(C-7); 121.7(C-3); 77.9 (C-6); 29.9 (C-5).

Aristolactam BII (2): was obtained as a yellow, crystalline powder, melting point 248-250°C; $R_f = 0,38$ (*n*-hexane-acetone 67/33, v/v). FT-IR (KBr) ν_{\max} (cm⁻¹): 3449, 3194, 2926, 2855, 1717, 1644, 1607, 1462, 1379, 1031, 838, 720, 624, 530, 466. UV λ_{\max} MeOH nm (log ϵ): 205 (3.59); 232 (3.56); 263 (3.49); 276 (3.54); 286 (3.53); 317 (2.97); 385 (2.94). (+)-ESI-MS: m/z 302 [M+Na]⁺; 581 [2M+Na]⁺. $^1\text{H-NMR}$ (500 MHz, CDCl₃ + CD₃OD): δ (ppm) 9.12 (1H, m, H-5); 7.72 (1H, s, H-2); 7.72 (1H, m, H-8); 7.47 (2H, m, H-6, H-7); 7.00 (1H, s, H-9); 4.02 (3H, s, 4-OCH₃); 3.98 (3H, s, 3-OCH₃). $^{13}\text{C-NMR}$ (125 MHz, CDCl₃ + CD₃OD): δ (ppm) 170.2 (C=O); 154.4 (C-3); 151.5 (C-4), 134.8 (C-8a), 134.2 (C-10), 128.9 (C-8), 127.4 (C-5), 127.4 (C-7), 126.9 (C-5a), 125.8 (C-6), 124.2 (C-10a), 121.2 (C-1), 120.8 (C-4a), 109.5 (C-2), 106.2 (C-9), 60.2 (4-OCH₃), 56.8 (3-OCH₃).

3-Methyl-1H-benz[f]indole-4,9-dione (3): was obtained as a yellow, solid powder, melting point 248-249°C; $R_f = 0,44$ (*n*-hexane- CH₂Cl₂ 25/75, v/v). (+)-ESI-MS: m/z 210 [MH]⁺, (+)-ESI-MS: m/z 212 [M+H]⁺; (+)-HR-ESI-MS m/z 212,0710 [M+H]⁺, FT-IR (KBr) ν_{\max} (cm⁻¹): 3429, 3322, 2938, 1647, 1588, 1507, 1403, 1238, 1031, 932, 714. UV (MeOH) λ_{\max} nm (log ϵ): 258 (4,07); 330 (3,32). $^1\text{H-NMR}$ (500 MHz, CDCl₃+ CD₃OD): δ (ppm) 8.08 (1H, m, H-5); 8.01 (1H, m, H-8); 7.59 (2H, m, H-6, H-7); 6.83 (1H, m, H-2); 2.34 (3H, s, CH₃-10). $^1\text{H-NMR}$ (500 MHz, DMSO): δ (ppm) 12.65 (1H, br s, H-1); 8.18 (2H, m, H-5, H-8); 7.76 (2H, m, H-6, H-7); 7.13 (1H, m, H-2); 2.31 (3H, s, CH₃-10). $^{13}\text{C-NMR}$ (125 MHz, CDCl₃

+ CD₃OD): δ (ppm) 182.3 (C-9); 175.7 (C-4); 134.5 (C-8a); 133.4 (C-4a); 133.2 (C-6, C-7); 132.7 (C-9a); 126.5 (C-5); 126.0 (C-8); 125.5 (C-2); 125.1 (C-3a); 122.8 (C-3); 11.0 (CH₃-10).

Cytotoxicity Assay

Cytotoxicity of these compounds was evaluated against four types of cancer cell lines: epithelial cancer (KB), liver (Hep G2); breast cancer (MCF-7) and lung (LU-1) by 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay. Cell lines were cultured in Dulbecco's Modified Eagle medium (DMEM) with 10% foetal bovine serum, at 37°C, 5% CO₂ and 95% humidity. Cells were plated in 96-well flat bottom tissue culture plates and allowed to density of approximately 3x10⁴/mL then begin experiment. The cells were incubated with 100 μ L of compound at a range of concentration (from 1 μ g/mL to 1000 μ g/mL) for 72 hours. Then, 0.2 mg/mL MTT reagent was added to each well, and the plate was incubated for 4 hours at 37°C. Next, DMSO (100 μ L) was added and the plates shaken for 5 min. The absorbance for each well was measured at 540 nm in a microtiter plate reader and the percentage cell viability (CV) was calculated manually using the formula:

$$CV = \frac{\text{Average abs of duplicate drug wells}}{\text{Average abs of control wells}} \times 100$$

The effects of compounds were expressed by IC₅₀ values (the drug concentration reducing the absorbance of treated cells by 50% with respect to untreated cells) (Monks *et al.*, 1991).

RESULTS AND DISCUSSION

Isolation and Chemistry

Goniothalamine (1)

Goniothalamine (1) was obtained as a light yellow, solid powder, melting point 82-84°C and the optical rotation $[\alpha]_D^{27} = +81,0^\circ$ (c 0,2; EtOH). The ring γ -lactone α,β -unsaturated showed typical absorption bands arising from carbonyl group (1707 cm⁻¹) in IR spectrum, maximum absorption wavelength at λ_{max} 210 nm (log ϵ 4,43; MeOH) in UV spectrum and δ_C 163,8 in ¹³C-NMR spectrum. The molecular formula was established as C₁₃H₁₂O₂ based on a molecular ion peak at m/z 223 [M + Na]⁺. The ¹H-NMR spectrum showed the presence of phenyl group with five signals proton at δ_H 7,27- 7,40 (5H, m, C₆H₅), signals of 4 proton olefinic at δ_H 6,92 (1H, ddd, J=4,0; 5,0; 9,5 Hz, H-4); 6,73 (1H, d, J=16,0 Hz, H-8); 6,27 (1H, dd, J=6,5; 16,0 Hz, H-7); 6,09 (1H, dt, J=2,0; 10,0 Hz, H-3). The ¹H-NMR spectrum of compound 1 also exhibited the presence of a methylene signal at δ_H 2,54 (m, 2H, H-5) and a methoxy signal at δ_H 5,10 (1H, ddd, J=1,0; 6,5; 15,5 Hz; H-6). The ¹³C-NMR and DEPT spectra of compound 1 indicated the presence of 13 carbon atoms in the molecule. There are a carbonyl carbon (δ_C 163,8), one CH₂ group at δ_C 29,9, one sp³ methine oxygenated carbons at δ_C 77,9; nine methine sp² group at δ_C 121,7-144,5 and one quaternary carbon sp³ at δ_C 135,8. The signal in COSY spectra allowed correlating from C-3 to C-8. The coupling constants J(16,0 Hz) between H-7 and H-8 indicated that

two protons are in trans position. The coupling constants J (9.5 Hz) between H-3 and H-4 indicated that two protons are in cis position. Based on the above evidence and the literature data (Wattanapiromsakul *et al.*, 2005) compound 1 was identified as goniothalamine. This compound is styryl lactone has been isolated from many Goniothalamus species. It has antitumor activity in vitro and in vivo (de Fátima *et al.*, 2005; Mereyala and Joe, 2001; Tian *et al.*, 2006). Furthermore, it has antifungal (Fatima *et al.*, 2008) and antioxidant activity (Kim *et al.*, 2012).

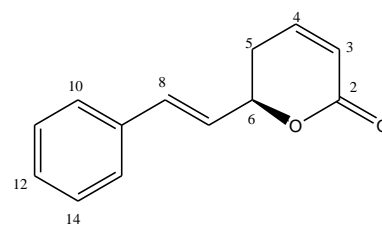


Fig. 1: Structures of compound 1.

Aristolactam BII (2)

Compound 2 was obtained as a yellow, crystalline powder, melting point 248-250°C, gave positive result with Dragendorff agent. The IR spectrum showed typical absorption bands arising from NH (ν_{max} 3449 cm⁻¹) and C=O (ν_{max} 1717 cm⁻¹). The UV spectrum in solvent MeOH showed typical maximum absorption wavelength of phenanthrene structure at λ_{max} nm (log ϵ): 232 (3.56), 263 (3.49), 276 (3.54), 286 (3.53), 317 (2.97) and 385 (2.94). The molecular formula was established as C₁₇H₁₃NO₃ based on a molecular ion peak at m/z 302 [M + Na]⁺. The ¹³C-NMR spectra of compound 2 indicated the presence of 17 carbon atoms in the molecule. There are a carbonyl carbon (δ_C 170.2), two methoxy groups (δ_C 60.2 and 56.8), six methine sp² group at δ_C 106.2; 109.5; 125.8; 127.4; 127.4; 128.9 and eight quaternary carbon sp³.

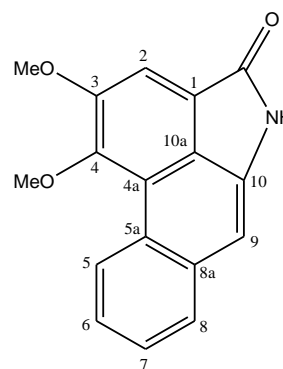


Fig. 2: Structures of compound Aristolactam BII (2);

The ¹H-NMR spectrum of compound 2 exhibited the presence of 6 signals proton of aromatic ring at δ_H 9.12 (1H, m, H-5); 7.72 (1H, s, H-2); 7.72 (1H, m, H-8); 7.47 (2H, m, H-6 + H-7); 7.00 (1H, s, H-9) and two methoxy groups OCH₃ at δ_H 4.02 (3H, s, OCH₃); 3.98 (3H, s, OCH₃). HMBC spectrum showed the correlation between H-9 with C-8, C-5a, C-8a, C-10a and C-10;

between H-5 with C-7, C-8a and C-5a, between H-8 with C-6, C-7, C-5a and C-9. Then it can deduced the correlation between C-8 with C-8a, C-9, C-10 and C-10a; C-5 with C-5a, C-8a and C-7. There is also the correlation between H-2 with C-3, C-4, C=O and C-1, hence C-2 correlates with C=O through C-1. The correlation between protons of methoxy group (δ_H 4.02) with C-3 (δ_C 154.4) and protons of methoxy group (δ_H 3.98) with C-4 (δ_C 151.5) indicated that two methoxy OCH₃ group linked with carbon C-3 and C-4. Based on the above evidence and the literature data (Marques *et al.*, 2011) compound **2** was identified as Aristolactam BII. This compound has neuroprotective (Kim *et al.*, 2004) and cytotoxicity with P-388 leukemia and A-549 lung adenocarcinoma epithelial cell line (Tsai *et al.*, 2005).

3-Methyl-1H-benz[f]indole-4,9-dione (3)

Compound **3** was obtained as a yellow, solid powder. The molecular formula was established as C₁₃H₉O₂N based on a molecular ion peak at *m/z* 212,0710 [M+H]⁺. The IR spectrum showed typical absorption bands arising from NH (ν_{max} 3429 cm⁻¹) and carbonyl C=O (ν_{max} 1647 cm⁻¹).

The ¹H-NMR spectrum of compound **3** exhibited the presence of 5 signals proton of aromatic ring at δ_H 8.08 (1H, m, H-5); 8.01 (1H, m, H-8), 7.59 (2H, m, H-6 and H-7) and 6.83 (1H, m, H-2); and 1 methyl group at δ_H 2.31 (3H, s). The ¹H-NMR spectrum of compound **3** also exhibited the presence of a proton NH signal at δ_H 12.65 (1H, br s, H-1). The ¹³C-NMR spectra of compound **3** indicated the presence of 13 carbon atoms in the molecule. There are a methyl group; 5 methine sp² groups; and 2 carbonyls group at δ_C 182.3 (C-9) và 175.7 (C-4) and 5 quaternary carbons in aromatic ring at δ_C 122.8 (C-3), 125.1 (C-3a), 133.4 (C-4a), 134.5 (C-8a) and 132.7 (C-9a).

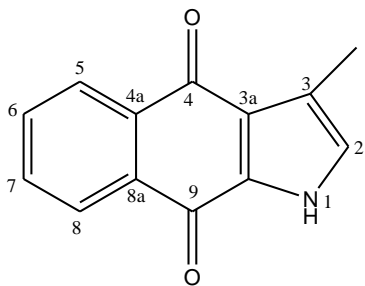


Fig. 3: Structures of compound 3-Methyl-1H-benz[f]indole-4,9-dione (3).

The signal in COSY spectra allowed determining the correlation from H-3 to H-8. HMBC spectrum showed the correlation between H-5 and H-8 with C-4a and C-8a, confirmed the presence of benzene A ring. Also there are correlations between 2 carbonyls C-4 and C-9 with H-5 and H-8 in HMBC spectrum respectively. It allowed confirming the C-4 and C-9 linked to ring A at position C-4a and C-8a. The pyrole C ring also is determined by HMBC spectrum: H-2 correlates with C-3, C-3a and C-9a; CH₃-10 correlates with C-2 and C-3a. The methyl group position at C-3 in pyrole C ring is also confirmed by NOE correlation between H-2 and CH₃-10. Based on the above evidence

and the literature data (Efdi *et al.*, 2010) compound **3** was identified as 3-methyl-1H-benz[f]indole-4,9-dione. This compound was first isolated from *Goniothalamustapis* Miq (Efdi *et al.*, 2010).

Bioactivity

The result of cytotoxicity tests of isolated compounds against epithelial cancer (KB), liver (Hep G2), breast cancer (MCF-7) and lung (LU-1) cell lines are shown in Table 1.

Table 1: Cytotoxicity of isolated compounds

Compounds	IC ₅₀ (μg/mL)			
	KB	HepG2	LU-1	MCF7
Compound 1	4.18 ± 0.60	8.62 ± 1.24	8.04 ± 1.18	13.31 ± 1.45
Compound 2	52.32 ± 5.21	85.29 ± 3.47	98.34 ± 6.13	134.87 ± 7.97
Compound 3	67.32 ± 3.73	94.53 ± 5.32	105.43 ± 2.41	167.89 ± 3.54

Only compound **1** exhibited strong cytotoxic activity against epithelial cancer (KB), liver (Hep G2), breast cancer (MCF-7) and lung (LU-1) cell lines. Previous study have demonstrated *Goniothalam* possess anticancer potential activity through mechanism of induces apoptosis and inhibits TNF- α -induced NF- κ B activation (Orlikova *et al.*, 2013; Seyed *et al.*, 2014). For the compounds **2**, **3**, we have found the IC₅₀ against fours tested cancer cell line were greater over than ten times as compared with compound **1**.

CONCLUSION

From the bark of *Goniothalamus chinensis* Merr. & Chun. we have isolated three compounds, *Goniothalam* (**1**), Aristolactam BII (**2**), 3-Methyl-1H-benz[f]indole-4,9-dione (**3**). The compound **1** showed the highest potency against the epithelial cancer (KB) cell line with an IC₅₀ of 4.18 ± 0.60 μg/mL. Others compounds have weak cytotoxic activity against all cell lines tested.

CONFLICT OF INTERESTS

The authors declare that there is no conflict of interest regarding the publication of this paper.

REFERENCES

- Alabsi, A.M., Ali, R., Ali, A.M., Al-Dubai, S.A., Harun, H., Abu Kasim, N.H., Alsalahi, A. Apoptosis induction, cell cycle arrest and in vitro anticancer activity of goniothalam in a cancer cell lines. Asian Pacific journal of cancer prevention : APJCP, 2012; 13: 5131-5136.
- Blázquez, M.A., Bermejo, A., Zafra-Polo, M.C., Cortes, D. Styryl-lactones from *Goniothalamus* species-A review. Phytochemical Analysis, 1999; 10: 161-170.
- de Fátima, Â., Kohn, L.K., Antônio, M.A., de Carvalho, J.E., Pilli, R.A. (R)-*Goniothalam*: total syntheses and cytotoxic activity against cancer cell lines. Bioorganic & medicinal chemistry, 2005; 13: 2927-2933.
- Efdi, M., Fujita, S., Inuzuka, T., Koketsu, M. Chemical studies on *Goniothalamus tapis* Miq. Natural product research, 2010; 24: 657-662.
- Fatima, A.d., Martins, C.V.B., de Resende, M.A., Ferreira Magalhaes, T.F., Lima, S., Henrique, B., Watanabe, G.A., Gois Ruiz,

A.L., de Carvalho, J.E., Pilli, R.A. Antifungal activity of goniiothalamine enantiomers. *Letters in Drug Design & Discovery*, 2008; 5: 74-78.

Kim, R., Bihud, V., bin Mohamad, K., Leong, K., bin Mohamad, J., bin Ahmad, F., Hazni, H., Kasim, N., Halim, S., Awang, K.. Cytotoxic and Antioxidant Compounds from the Stem Bark of *Goniiothalamus tapisoides* Mat Salleh. *Molecules*, 2012; 18: 128.

Kim, S.R., Sung, S.H., Kang, S.Y., Koo, K.A., Kim, S.H., Ma, C.J., Lee, H.-S., Park, M.J., Kim, Y.C. Aristolactam BII of *Saururus chinensis* attenuates glutamate-induced neurotoxicity in rat cortical cultures probably by inhibiting nitric oxide production. *Planta medica*, 2004; 70, 391-396.

Marques, A.M., Velozo, L., de Moreira, D., Guimaraes, E.F., Kaplan, M. Aristolactams from roots of *Ottonia anisum* (Piperaceae). *Natural product communications*, 2011; 6: 939-942.

Mereyala, H., Joe, M. Cytotoxic activity of styryl lactones and their derivatives. *Current Medicinal Chemistry-Anti-Cancer Agents*, 2001; 1: 293-300.

Monks, A., Scudiero, D., Skehan, P., Shoemaker, R., Paull, K., Vistica, D., Hose, C., Langley, J., Cronise, P., Vaigro-Wolff, A., Gray-Goodrich, M., Campbell, H., Mayo, J., Boyd, M. Feasibility of a High-Flux Anticancer Drug Screen Using a Diverse Panel of Cultured Human Tumor Cell Lines. *Journal of the National Cancer Institute*, 1991; 83: 757-766.

Orlikova, B., Schumacher, M., Juncker, T., Yan, C.C., Inayat-Hussain, S.H., Hajjouli, S., Cerella, C., Dicato, M., Diederich, M. Styryl-lactone goniiothalamine inhibits TNF- α -induced NF- κ B activation. *Food and chemical toxicology : an international journal published for the British Industrial Biological Research Association*, 2013; 59: 572-578.

Petsophonsakul P., Pompimon W., Banjerdpongchai R. Apoptosis induction in human leukemic promyelocytic HL-60 and monocytic U937 cell lines by goniiothalamine. *Asian Pacific journal of cancer prevention : APJCP*, 2013; 14: 2885-2889.

Seyed, M.A., Jantan, I., Bukhari, S.N.A. Emerging Anticancer Potentials of Goniiothalamine and Its Molecular Mechanisms. *BioMed Research International*, 2014: 536508.

Sun, L., Zhao, R., Lan, X., Chen, R., Wang, S., Du, G. Goniolactone C, a styryl lactone derivative, inhibits PDGF-BB-induced vascular smooth muscle cell migration and proliferation via PDGFR/ERK signaling. *Molecules*, 2014; 19: 19501-19515.

Tian, Z., Chen, S., Zhang, Y., Huang, M., Shi, L., Huang, F., Fong, C., Yang, M., Xiao, P. The cytotoxicity of naturally occurring styryl lactones. *Phytomedicine*, 2006; 13: 181-186.

Tsai, I.-L., Lee, F.-P., Wu, C.-C., Duh, C.-Y., Ishikawa, T., Chen, J.-J., Chen, Y.-C., Seki, H., Chen, I.-S. New cytotoxic cyclobutanoid amides, a new furanoid lignan and anti-platelet aggregation constituents from *Piper arborescens*. *Planta medica*, 2005; 71: 535-542.

Vendramini-Costa, D.B., Monteiro, K.M., Iwamoto, L.H., Jorge, M.P., Tinti, S.V., Pilli, R.A., de Carvalho, J.E. Gastroprotective effects of goniiothalamine against ethanol and indomethacin-induced gastric lesions in rats: Role of prostaglandins, nitric oxide and sulfhydryl compounds. *Chemico-biological interactions*, 2014; 224C, 206-212.

Wattanapiromsakul, C., Wangsintaweekul, B., Sangprapan, P., Itharat, A., Keawpradub, N. Goniiothalamine, a cytotoxic compound, isolated from *Goniiothalamus macrophyllus* (Blume) Hook. f. & Thomson var. *macrophyllus*. *Songklanakarin J. Sci. Technol*, 2005; 27: 479-487.

Wiert, C. *Goniiothalamus* species: a source of drugs for the treatment of cancers and bacterial infections? Evidence-based complementary and alternative medicine : eCAM, 2007; 4: 299-311.

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