

Preparation and characterization of BSA and chitosan nanoparticles for sustainable delivery system for quercetin

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ABSTRACT

The current study was undertaken in order to evaluate the efficacy of BSA and chitosan nanoparticle for sustainable delivery of Quercetin. BSA and chitosan nanoparticles were prepared by coacervation process and Ionic gelation method respectively and were characterized by Scanning Electron Microscope. The average size of BSA and chitosan nanoparticles was found to be 96nm and 42.7nm respectively. Quercetin loaded BSA and chitosan nanoparticles were smooth and spherical with the average size of 317.1 nm and 44.1nm respectively. The encapsulation efficiency of the BSA and chitosan nanoparticles were calculated and it was found that the drug encapsulation efficiency was more in chitosan than in BSA. The in vitro analysis of Quercetin loaded BSA and chitosan nanoparticles were studied using dialysis bag. The nanoformulation of Quercetin loaded BSA nanoparticles shows a sustainable release of drug in which 56% of the drug was released till 24 hours and it did not change further. For Quercetin loaded chitosan nanoparticles it shows 72% of in vitro drug release for 24 hours and found to be better than BSA nanoparticle. Hence both Quercetin loaded BSA and chitosan nanoparticles were effective in encapsulation and sustainable release which also paves way for the bio availability of Quercetin

INTRODUCTION

Drug delivery is a promising approach for the biotechnologist and pharmacologist to develop a delivery system that can aid the drug to its target and maintaining its availability for their therapeutic action. Efficacy of most of the drugs were limited for several reasons such as poor absorption, poor solubility, nonspecific delivery, short circulating half life, high toxicity, high dosage etc (Orive *et al.*, 2003; Praveen *et al.*, 2012). It is always a dream for the scientist to develop a drug with the higher degree of specificity, solubility, improved efficacy with lesser side effects. Researchers have recognized the nanotechnology as a best solution to overcome the above said problem in developing an effective drug. Numerous nanotechnology platforms such as fullerenes, nanotubes, quantum dots, nanopores, dendrimers, liposomes, magnetic nanoprobes and radio controlled nanoparticles are being identified so far (Faraji and Wipf, 2009). Nanoparticles that are compatible to the biological system have numerous applications

in medicine (Chambers and Mitragotri, 2004; Surendran *et al.*, 2009). They are considered as a best drug carrier system because of easy manipulation of particle size and surface, freedom to modify the fundamental properties such solubility, diffusivity and absorption (Jun *et al.*, 2011). Moreover, smaller in size gives the unique properties of nanoparticle with greater surface area and distinct physical and chemical properties (Li *et al.*, 2012) Drugs that are encapsulated in nanoparticles were found to provide enzymatic protection and sustained release that increases the bioavailability of the drug (Chambers and Mitragotri, 2004). It has been reported that nanoparticles can be a carrier for various pharmacological active substance such as phytochemicals, nutraceuticals and drugs (Rhaese *et al.*, 2003; Zhang *et al.*, 2008). Among numerous nanoparticle, Bovine Serum Albumin (BSA) and chitosan based drug carrier are currently in research in order to deliver various drugs such as low molecular drug, anticancer drug, DNA, vaccine etc (Taheri *et al.*, 2012). BSA nanoparticles are non toxic, biodegradable and biocompatible material, which are easily adaptable to the human body (Jahanshahi *et al.*, 2008; Jun *et al.*, 2011). It has potential usefulness in transporting fatty acids, endogenous and exogenous compounds throughout the body (Fang *et al.*, 2011; Ascenzi *et al.*, 2006; Ascoli *et al.*, 2006).

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BSA are rich in charged amino acids (lysine) that allow the positive and negative charged molecule to adsorb electrostatically without the participation of any other compounds (Tan *et al.*, 2013; Rabea *et al.*, 2003; Jayakumar *et al.*, 2010; Patel *et al.*, 2012; Namasivayam *et al.*, 2014). Chitosan is a polycationic polysaccharide distributed in crustacean kingdom. It is used for biomedical application such as scaffold in tissue engineering, wound dressing, drug carrier and antimicrobial agent (Jameela and Jayakrishnan, 1995; Scheffel *et al.*, 1973; Sahoo *et al.*, 2011). Chitosan nanoparticle is unique in targeting the drugs to various organs such as liver, colon, kidney and lungs and thus provides a suitable carrier for various drugs especially for cancer (Chakraborty *et al.*, 2012). Quercetin (3, 5, 7, 3', 4'-pentahydroxyflavone) is bioflavonoid found in some fruits vegetables, food and beverages (Formica and Regelson, 1995). It forms the backbone for other flavonoids such as rutin, hesperidins, Naringenin and tangeritin (Lakhanpal and Rai, 2007). It is an antioxidant and found to regulate lipid peroxidation by chelating metal ions and also used as a radioprotector against UV radiation induced injury and cell death (Kandaswami *et al.*, 2005). It is a potent pharmacological active agent which has a wound healing, anti-inflammatory, anticancerous and antiviral properties (Comalada *et al.*, 2005; Kakran and Sahoo, 2011; Jagtap *et al.*, 2009). Quercetin incorporated collagen matrices were found to be a potent wound healer (Gomathi *et al.*, 2003). In spite of its pharmacological properties, its use is highly limited because of poor solubility and poor absorption in oral administration (Kakran and Sahoo, 2011). Hence the formulation of Quercetin that enhances the bioavailability will improve its pharmacological action. It has been reported that nanovehicle can overcome the above said problem. In this aspect our purpose of the current study is to evaluate the production and characterization of BSA and chitosan nanoparticle and to study the efficacy of Quercetin encapsulation and sustainable release of Quercetin from BSA and chitosan nanoparticle

MATERIALS AND METHODS

The chemicals for the preparation of BSA and chitosan nanoparticles were obtained from Reachem Chemicals. Quercetin (Extra pure) was obtained from Sisco Laboratories, Mumbai. All other chemicals used were analytical grade and high purity chemicals were purchased from SRL chemicals. Deionized water was used throughout the experiment.

Preparation of BSA Nanoparticles

The method adopted for the preparation of BSA nanoparticle was desolvation method (Jahanshahi *et al.*, 2008; Langer *et al.*, 2003). 1% BSA was prepared in deionised water at pH 7 and titrated with acetone by stirring at 700 rpm using magnetic stirrer at room temperature until the solution turned turbid (Sailaja and Amareshwar, 2012). 25% of Glutaraldehyde was added drop wise as a cross linker and the mixture was maintained in stirring condition for 3 hours. The resulting mixture

was centrifuged at 5000rpm for 30 minutes. The supernatant was discarded. The pellet was washed with acetone and then with deionized water for further purification. The pellet was collected, dried and characterized.

Preparation of Chitosan Nanoparticles

Ionic gelation method was followed for the preparation of chitosan nanoparticles (Calvo *et al.*, 1997). 0.3% chitosan was prepared in 1.5% of acetic acid. The chitosan nanoparticles were prepared with drop wise addition of Sodium Tripolyphosphate (1mg/ml) under magnetic stirring at room temperature for 1 hr. The resulting mixture was centrifuged at 15000 rpm for 20 minutes. The pellet was collected and was used for characterization.

Quercetin loaded in BSA Nanoparticles

The BSA nanoparticles were loaded with the drug Quercetin and were checked for its entrapment efficiency. The protocol for the encapsulation of the drug was referred from the studies led by Namasivayam *et al.*, (2014) with minor modification. 1% BSA was prepared and 0.5g of Quercetin was dissolved in it. The solution was titrated with acetone until the solution became turbid. 25% of glutaraldehyde was added to the solution for cross linking and the solution was agitated for 3 hours in a magnetic stirrer. The solution was ultrasonicated to reduce the size of the particle which was then centrifuged at 5000rpm for 30 minutes. The supernatant was subjected for encapsulation efficacy and the pellet was collected for further characterization.

Quercetin loaded in Chitosan Nanoparticles

0.15 g of Quercetin was dissolved in 0.3% of chitosan prepared in 1.5% of acetic acid which was titrated against Sodium Tripolyphosphate (TPP) kept in magnetic stirrer. The solution was then subjected to ultrasonication which reduced the particle size and centrifuged at 5000rpm for 30 minutes. The supernatant was subjected for encapsulation efficacy and the pellet was collected for further characterization.

Encapsulation Efficiency

To check the encapsulation efficiency, the free Quercetin that was present in the supernatant after centrifugation was measured in UV Spectrophotometer at 370nm. The encapsulation efficiency was calculated using the formula:

$$\text{Entrapment Efficiency (\%)} = \frac{\text{Total Quercetin} - \text{Quercetin in supernatant}}{\text{Total Quercetin}} * 100$$

In vitro drug release

In order to study the *in vitro* drug release dialysis bag from Himedia (Membrane -50) was used. 10 mg of the Quercetin loaded BSA and Chitosan nanoparticles were taken and were dispersed in 1ml of phosphate buffer (0.1M/L) solution which was placed in a Himedia dialysis membrane bag. The dialysis membrane bag was immersed in 150ml phosphate buffer solution

which was continuously stirred. 5ml of samples were collected at definite time intervals which were replaced with phosphate buffer solution. The drug release was determined by taking O.D in UV-Vis Spectrophotometer at 370nm (Namasivayam *et al.*, 2014; Shaikh *et al.*, 2009).

Characterization of BSA and Chitosan nanoparticles

The produced BSA and Chitosan nanoparticles were characterized using Scanning Electron Microscope (SEM) analysis, Carl Zeiss, Germany before and after encapsulation the drug reveals the size, shape and structure of the nanoparticle. It also tells whether the drug has been loaded into the prepared BSA and chitosan nanoparticles.

RESULTS AND DISCUSSION

Characterization of BSA and Chitosan Nanoparticle

The size and shape of nanoparticle was determined by using Scanning Electron Microscope. It was clear that most of the morphologies of both chitosan and BSA nanoparticle were semisphere with diameter ranges between 25-120nm. The images of the nanoparticle for BSA and chitosan were illustrated in Fig 1 and Fig 2 respectively. The average size of BSA nanoparticle was 96nm which matched with previous study.

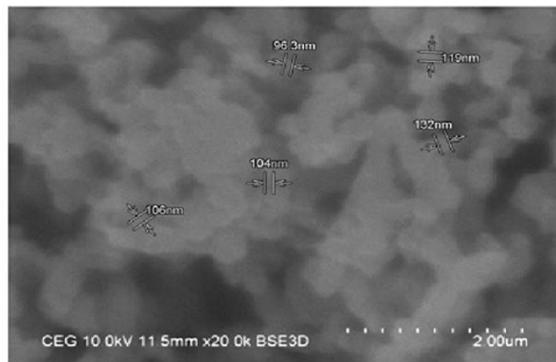


Fig. 1: SEM image of BSA nanoparticle.

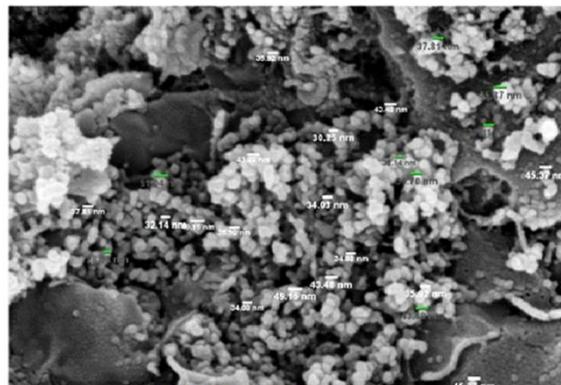


Table 1: Drug encapsulation efficiency of BSA and Chitosan nanoparticles.

| S.No | Nanoparticle | % of encapsulation |
|------|--------------|--------------------|
| 1 | BSA | 59 |
| 2 | Chitosan | 61 |

In vitro drug release of BSA and chitosan nanoparticles

The *in vitro* drug release was studied in BSA and chitosan nanoparticles using the dialysis method. Fig 5 displayed a graph showing the *in vitro* drug release of BSA nanoparticles in which 56% of the drug was released till 24 hours and was not found to change further. Fig 6 displayed a graph showing 72% of *in vitro* drug release of chitosan nanoparticles. The *in vitro* drug release profile of BSA was studied by Yu et al (2014) which confirmed that BSA nanoparticle exhibited a good sustainable release profile due to which it can be used a drug delivery system. Similarly Dounighi et al (2012) studied the *in vitro* release of scorpion venom as an antigen delivery system in which the chitosan nanoparticle showed an initial burst of release of approximately 60% in first ten hours, which was in accordance with our results.

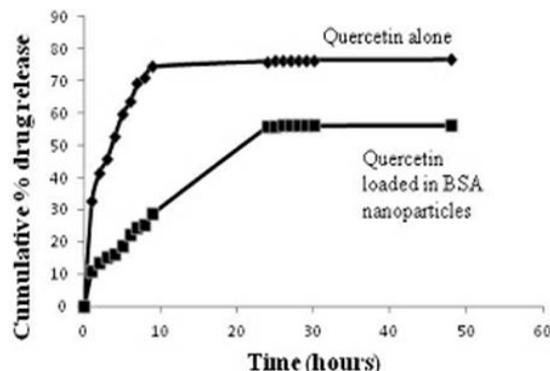


Fig. 5: Graph showing cumulative drug release of quercetin loaded BSA nanoparticles.

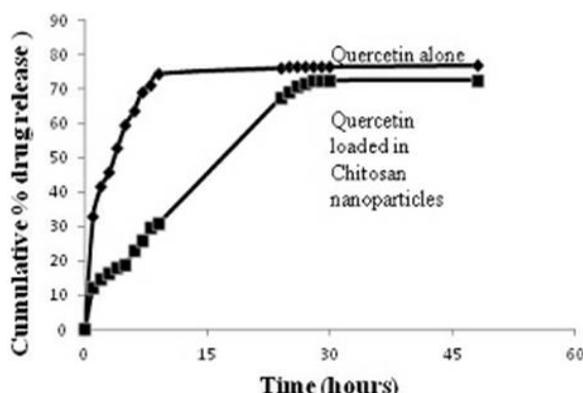


Fig. 5: Graph showing cumulative drug release of quercetin loaded BSA nanoparticles.

CONCLUSION

In this study, BSA and chitosan nanoparticles were prepared by using desolvation and ionic gelation method and this was confirmed by SEM analysis with the minimum particle size. These nanoparticles also showed a maximum Quercetin

encapsulation efficiency. It was clear that successful encapsulation of Quercetin in chitosan nanoparticle will provide the bioavailability of Quercetin in the system. The pharmacological properties of Quercetin and their uses in medicine were limited due to their poor availability our encapsulation result for BSA and chitosan nanoparticle overcomes their Quercetin limitation by increasing their availability and thus BSA and chitosan can be considered as an attractive polymers that can be used as a good carrier for drugs. The *in vitro* studies reveal that the BSA and chitosan nanoparticle loaded with Quercetin showed a sustainable release of about 56% and 72% respectively. Therefore the current study suggest that the pharmacological properties of Quercetin can be increased by using both BSA and chitosan nanoparticle as a good drug delivery system. From the present study it was found that chitosan nanoparticle when compared with BSA showed increased encapsulation efficiency and sustainable release of the Quercetin. Further studies have to be done by encapsulating other drugs into the chitosan nanoparticle to make it as a better drug delivery system.

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