

Estimation of Bosentan Monohydrate in Male Rabbit Plasma by using RP-HPLC Method

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ABSTRACT

A new method for the quantitative determination of bosentan monohydrate in male rabbit plasma was developed by Reverse Phase HPLC as per standard regulatory guidelines. Precipitation method was used for extracting the drug from the drug spiked plasma using mobile phase. The analyte extracted was injected in to ODS column C18 (250 x 4.6 mm, 5 μ m), temperature was maintained at 25^oC and monitoring of effluent was performed at 273 nm. The mobile phase consist acetonitrile and phosphate buffer (pH 4.6) in 80:20 ratio. The method developed shown high specificity for bosentan monohydrate. The calibration curve for bosentan monohydrate was linear from 0.1-5 μ g/0.5mL in plasma ($R^2=0.9999$). The lower limit of detection is 0.1 μ g/0.5mL. Developed method was verified for precision in intra-day and inter-day and was found within the limits. The accuracy of the developed Reverse Phase HPLC method was found to be 99.8 \pm 1.25-98.7 \pm 1.29% and is found to be reproducible. The method developed was proved to be adequate, sensitive and reproducible for the determination of bosentan monohydrate in rabbit plasma.

INTRODUCTION

Bosentan monohydrate is used as pulmonary arterial anti-hypertensive drug. It acts by blocking endothelien receptor ET₁ and ET₂ where their main action is constriction of pulmonary artery (Dermot *et al.*, 2011, Christian *et al.*, 2008). Bosentan monohydrate is chemically represented as 4-tert-butyl-N-[6-(2-hydroxyethoxy)-5-(2-methoxyphenoxy)-2-pyrimidin-2-ylpyrimidin-4-yl] benzenesulfonamide and its chemical formula is C₂₇H₂₉N₅O₆S (Figure 1) (National Centre for Biotechnology Information, 2017). Previous works reported analytical methods for the estimation of bosentan monohydrate in plasma. For the estimation of related substances in bosentan monohydrate RP-HPLC method was developed (Jadhav *et al.*, 2011). The reported methods are sensitive but are having disadvantages like a narrow

range of linear response, more retention time and lesser precision. Two narrow-bore LC-MS methods were reported (Lausecker *et al.*, 1995, Lausecker *et al.*, 2000).

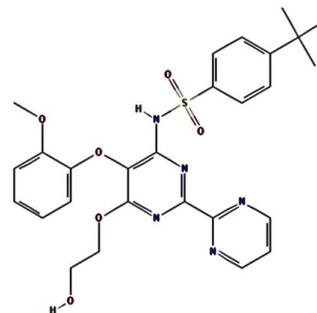


Fig. 1: Chemical structure of bosentan monohydrate.

The reported LC-MS methods involve costly detector and tiresome sample preparation steps. Our aim in the current research study is to develop a new RP-HPLC method for estimation of drug in New Zealand rabbit plasma which is very accurate & sensitive.

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MATERIALS AND METHODS

Chemicals and solvents

Bosentan monohydrate and etodoloac were gift samples received from MSN laboratories, Hyderabad, India. HPLC grade Acetonitrile, HPLC grade Methanol and all other chemicals used were obtained from Merck chemical division, Mumbai. HPLC grade water was obtained from Milli-Q water purification system.

Instrumentation

Chromatography was performed using waters 2695 HPLC. The HPLC system contains high speed auto sampler, column oven, degasser and 2996 PDA detector. Isocratic pump mode was used in this system. Class Empower-2 software was used for monitoring the operations. The column used was ODS (C₁₈ 250 x 4.6 mm, 5 μ L) and the flow rate was 1 mL/min. The temperature of the column was set at 25^oC and runtime was 10 min. The volume of injection loop was 20 μ L and the detection was done at the wave length of 273 nm.

HPLC conditions

Both acetonitrile and phosphate buffer (containing 0.1% orthophosphoric acid and pH was adjusted to 4.6 using triethanolamine) were filtered through 0.45 μ m membrane filter. The solvent system was pumped at the ratio of 80:20v/v to the column at a flow rate of 1 mL/min

Preparation of phosphate buffer (pH 4.6)

7.0g of potassium dihydrogen phosphate was taken in 1000 mL volumetric flask and 900 mL of HPLC grade water was added and stirred well. 100 mL of water was added and the pH was adjusted to 4.6 with 0.1% orthophosphoric acid and triethanolamine solution. Fresh buffer was used on each day.

Preparation of mobile phase

200mL of phosphate buffer (pH 4.6) and 800mL of acetonitrile were taken in 1000mL volumetric flask and degassed for 5 min using sonicator and the resultant mobile phase was filtered through 0.45 μ m membrane filter under vacuum. The same mobile phase was used as diluent.

Preparation of standard solution

10mg of bosentan monohydrate was weighed accurately and transferred in to 100mL volumetric flask. Ensured to dissolve the un-dissolved portions of drug by sonicating and final volume was made with mobile phase. Aliquot of 1 mL was taken and transferred into a 10 mL clean and dry volumetric flask and it was diluted up to the mark with the same diluent (10 μ g/mL). From this 1mL was taken and further diluted to 10mL to get a concentration of 1 μ g/mL.

Collection of blood samples from New Zealand rabbits

Blood samples were collected in heparinized eppendroffs tube before administration of dosage form from left marginal ear

vein of rabbit (CPCSEA No.:1677/PO/Re/S/2012/CPCSEA). From the blood cells plasma was separated by centrifuging blood at 8000rpm for 10 min. The separated plasma was stored at -20^oC until further study.

Extraction of drug from plasma of New Zealand rabbits

250 μ L of plasma containing varying amount of bosentan monohydrate was taken in to a teflon lined cap centrifuging tube followed by addition of 50 μ L of 2 μ g/0.5mL of etodoloac (internal standard) and 2 mL of acetonitrile. The mixture was vortexed for 2 min and centrifuged at 3300 rpm speed until organic layer was separated. Organic layer was collected and filtered through 0.2 μ m membrane filter. 20 μ L of filtrate was injected into the analytical column and the eluents were observed at 273nm. The concentrations in the range of 0.1 to 5 μ g/0.5mL of plasma were prepared. The experiment was repeated for five times for all the concentrations.

Validation

The proposed RP-HPLC method was validated for linearity, precision and accuracy. The precision (% coefficient of variation) was done by checking the inter-day and intra-day variation in the known drug concentration. The accuracy was done by estimating the percent recovery of known amount of drug added to the known amount of pre-analyzed bosentan monohydrate plasma sample (Validation of analytical procedure 1996, ICH guidelines 1996).

Selectivity

An aqueous mixture of bosentan monohydrate (1 μ g/mL) was prepared and injected in to the column to determine the retention time of drug. A drug free plasma sample was chromatographed to observe any interfering peaks at the retention time of drug. Spiked plasma containing 1 μ g/0.5 mL of bosentan monohydrate concentration was chromatographed to check the effect of various endogenous components in retention time (Ravichandran *et al.*, 2010)

Linearity

The linearity of the proposed RP-HPLC method was determined in terms of the correlation coefficient. Calibration curve was taken in to consideration. The peak area ratios of bosentan monohydrate and etodoloac (internal standard) were recorded and the regression of the plasma concentration over its peak area ratio was calculated using the least square method of analysis. Mean of five samples was taken for linearity determination.

Precision

The precision of the method was done by spiking the plasma containing 0.2, 0.5, 1 μ g/0.5mL of bosentan monohydrate containing etodoloac 2 μ g/0.5mL (internal standard), intra-day and inter-day variations were observed using the RP-HPLC method developed. The results obtained were noted. Each sample was

extracted from plasma as described previously. The precision of the developed method was calculated in terms of percent coefficient of variation. Mean of five samples was taken to determine the precision.

Accuracy

The accuracy of the developed RP-HPLC method was done by using recovery studies. Plasma samples containing different concentrations of drug (0.2, 0.5, 1 µg of bosentan monohydrate) was subjected to the developed method and each sample was injected three times. The difference in the measured concentration and that of the added quantity was expressed in terms of percent recovery (Peter *et al.*, 2007, ICH 2005).

RESULTS AND DISCUSSION

Standard calibration curve

The standard calibration curve of peak area ratios of bosentan monohydrate to etodolac in rabbit plasma was found to be linear (Figure 2). Various peak area ratios of bosentan monohydrate to etodolac against the concentration of drug was also reported (Table 1).

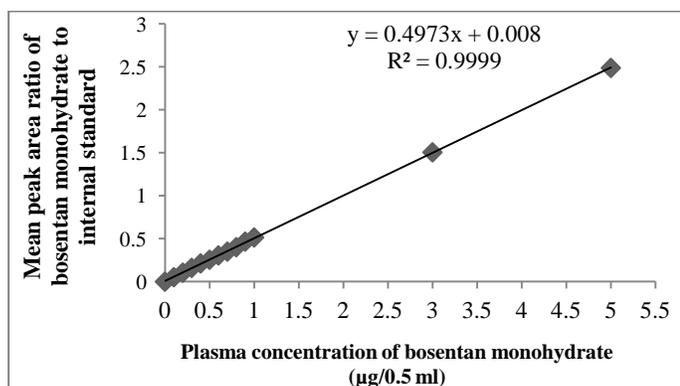


Fig. 2: Standard graph for the estimation of bosentan monohydrate in rabbit plasma

Table 1: Calibration of the RP-HPLC method for the estimation of bosentan monohydrate in rabbit plasma.

Plasma concentration of bosentan monohydrate (µg/0.5mL)	Peak area ratios*	% C.V.
0	0	0
0.1	0.052007	0.15
0.2	0.104633	0.05
0.3	0.160475	0.17
0.4	0.212688	0.03
0.5	0.254922	0.28
0.6	0.306102	0.47
0.7	0.349369	0.51
0.8	0.401659	0.42
0.9	0.466127	0.18
1	0.514828	0.51
3	1.505902	0.70
5	2.488393	0.47

*Mean of five determinations

Selectivity

The method was set with a run time of the 10 minutes. The retention time for bosentan monohydrate and etodolac were 3.6 minutes and 4.1 minutes respectively and was depicted in model chromatogram (Figure 3). The same bosentan monohydrate solution containing internal standard was injected for five times the retention time was found to be same. Blank chromatogram of plain plasma was also provided (Figure 4).

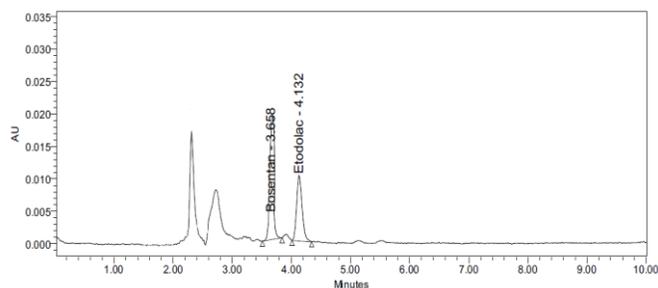


Fig. 3: A model chromatogram for estimation of bosentan monohydrate in rabbit plasma.

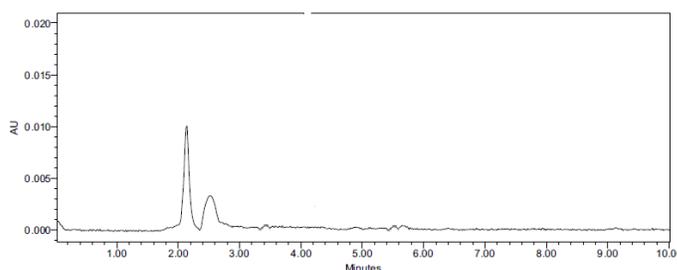


Fig. 4: Blank chromatogram of plasma.

Linearity

The peak area ratios of bosentan monohydrate to internal standard were reproducible which is also indicated by low coefficient of variation (0.03) (Table 1). The statistical analysis results by least square method confirmed a high correlation coefficient of 0.9999 in the range of 0.1 to 5µg/0.5mL (Figure 2) for the plasma concentration of bosentan monohydrate and its respective peak area ratios. The regression equation was found to be $Y=0.4973X+0.008$ for bosentan monohydrate concentration over its peak area ratio with internal standard where Y is the peak area ratio and X is the concentration of bosentan monohydrate. By using the regression equation estimation of the bosentan monohydrate in plasma or in validation study can be done. The lower limit of detection is 0.1 µg/0.5mL.

Precision

The present RP-HPLC method was also validated for intra-day and inter-day variation. The plasma samples containing drug (0.2, 0.5, 1 µg/0.5mL) were extracted as per the procedure described previously, and the resultant filtrate samples were repeatedly injected on the same day and on three different days. The coefficient of variation (CV) was calculated in the peak area of bosentan monohydrate for five replicate injections which was

found to be less than 1.4%. The inter-day variation (3 days and five injections) was found to be less than 1.4% and the results were reported (Table 2). Thus the present RP-HPLC method was proved to be highly reproducible and it was confirmed by the results.

Table 2: Inter-day and intra-day precision for the estimation of bosentan monohydrate in rabbit plasma.

Plasma concentration of bosentan monohydrate ($\mu\text{g}/0.5\text{mL}$)	Concentration of bosentan monohydrate ($\mu\text{g}/0.5\text{mL}$) found (Intra-day)			Concentration of bosentan monohydrate ($\mu\text{g}/0.5\text{mL}$) found (Inter-day)		
	Mean*	% CV	% Precision	Mean*	% CV	% Precision
0.2	0.198	1.2	99.2 \pm 1.2	0.195	1.4	97.5 \pm 1.4
0.5	0.499	1.1	99.8 \pm 1.1	0.496	1.2	99.2 \pm 1.2
1	0.999	1.4	99.9 \pm 1.4	0.994	1.2	99.4 \pm 1.2

*Mean of five determinations.

Accuracy

When a known amount of bosentan monohydrate (0.2, 0.5, 1 μg) was added to pre analyzed plasma samples (1 $\mu\text{g}/0.5\text{mL}$), there was a high recovery (99.8 \pm 1.25%) of the drug indicating that this RP-HPLC method is highly accurate and the results were reported (Table 3).

Table 3: Recovery of bosentan monohydrate after adding known amounts to pre-analyzed plasma samples (1 $\mu\text{g}/0.5\text{mL}$).

Amount of bosentan monohydrate added (μg)	*Mean \pm SD amount (μg) recovered	% Recovery
0.2	0.197 \pm 0.07	98.5 \pm 1.31
0.5	0.499 \pm 0.16	99.8 \pm 1.25
1	0.987 \pm 0.19	98.7 \pm 1.29

*Mean of three determinations.

CONCLUSION

The present RP-HPLC method described above is found to be precise, simple and highly accurate for the estimation of bosentan monohydrate in rabbit plasma samples. The Reverse Phase-HPLC method developed was validated as per standard regulatory guidelines and all the parameters satisfied prerequisites of robust and accurate method. Hence, adopting this method for quality control analysis of the bosentan monohydrate in rabbit plasma is confirmed.

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Conflict of Interests: There are no conflicts of interest.

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