

# The in Silico Study of Nutmeg Seeds (*Myristica fragrans* Houtt) as Peroxisome Proliferator Activated Receptor Gamma Activator Using 3D-QSAR Pharmacophore Modelling

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## ABSTRACT

In this study, we created a pharmacophore models from a dataset of agonists for PPAR gamma receptor using the Catalyst/Hypogen module. A training set consists of 22 compounds activity range between 0.1 to 3,500 nM, were carefully selected. In previous study, molecular docking of macelignan against PPAR $\gamma$  binding pocket showed a free energy binding of -11.07 kJ/mol, interaction with the hydrophobic pocket (diphenyl pocket)(Celik *et al.*, 2007), and a hydrogen bond network (His323, Tyr473, His449 and Ser289). The pharmacophore model (Hypo1), consisting of 5 features, i.e. one hydrogen bond acceptor (HBA), negative ionizable (NI), ring aromatic (RA) and two hydrophobics (HY) features, and one excluded volume. Hypo1 has the lowest total cost value (92.055), the highest cost difference (40.9316), the lowest RMSD (0.591049), and the best correlation coefficient (0.972949). Fourteen natural substances reported from nutmeg seeds (*Myristica fragrans* HOUTT.) were then mapped against Hypo1, and macelignan shows a fair fit value of 7.00102 with an estimated value of 1271.990 nM. This concludes, macelignan in nutmeg might have antidiabetic properties via PPAR $\gamma$  receptor activation.

## INTRODUCTION

Peroxisome Proliferator-Activated Receptors (PPARs) are members of the nuclear receptor super-family that is involved in protein gene expression for energy, glucose and lipid metabolism, proliferation and differentiation of adipocyte, as well as insulin sensitivity (Arck *et al.*, 2010; Hiukka *et al.*, 2010; Farce *et al.*, 2009). Three isoforms of these core receptors have been identified so far: PPAR $\alpha$ , PPAR $\beta/\delta$  dan PPAR $\gamma$ . Each of them has different selectivity towards type of tissue, type of ligand, and finally, unique biological response. PPAR $\gamma$  is mostly expressed in adipose tissue, colon and macrophages, and has been proven for its role in regulating carbohydrate metabolism as well as fatty acid storage. Finding a new and potent ligand for PPAR $\gamma$  is the main focus in research for treating patients with

Type II diabetes mellitus, as it can restore insulin sensitivity. PPAR $\gamma$  agonist has made a big attraction in the clinical management of cardiovascular risk factors associated with metabolic syndrome and Type 2 diabetes mellitus (Pearson, 2009). A few classes of PPAR $\gamma$  agonist have been reported to possess anti-diabetic properties, such as: thiazolidinediones, dihydrobenzofurans, dihydro-benzopyrans, benzofuran benzoxazoles and  $\alpha$ -amino- $\beta$ -phenylpropanoic acid derivatives (Henke, 2004b). Thiazolidinediones (TZDs) is an important class of PPAR $\gamma$  synthetic agonist (Feldman *et al.*, 2008).

TZDs are anti-diabetic agents that work on adipose tissue and increase insulin sensitivity. At this point, it is still used in the effort to treat Type II diabetes mellitus (Pourcet *et al.*, 2006). Despite the clinical benefit of TZDs, they are related with side effects such as an increased body weight, an increased adipogenesis, kidneys fluid retention, and possible increase in cardiovascular disorder occurrences (Jones, 2010).

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Thus, new PPAR $\gamma$  activator with an increased therapeutic efficacy and lesser side effects is necessary. Partial PPAR agonist is recently popular as a promising and new generation of PPAR activator. Partial agonist induces alternative receptor confirmation and as such, recruits different co-activator, which results in a different transcription effect when compared with TZDs. Therefore, it is indicated that such partial agonists will probably offer the required effectiveness and reduce side effects (Chang *et al.*, 2007; Yumuk, 2006). Natural products are an important and promising source in the discovery of new potent drugs (Newman and Cragg, 2007). Here, we created a pharmacophore models from a dataset of agonists for PPAR $\gamma$  receptor using the Catalyst/Hypogen module for predicting the nutmeg seeds compounds. In the previous study, Najjar *et al.* (2011) generated the pharmacophoric space of PPAR $\gamma$  using seven diverse sets of activators (Al-Najjar *et al.*, 2011). However, the compounds screened was not natural compounds. Thus, the purpose of this study was to identify the functional groups or features that activate the human PPAR $\gamma$  as agonist. The approach with identification and modelling of pharmacophore at 2-aryloxy-3-phenyl-propanoic acid has been done with *in silico* method.

## MATERIAL AND METHOD

### Selecting Training Set Compound

Choosing the training set is important for generating a hypothesis in the Hypogen and has to follow certain ground rules, such as a minimum of 16 varieties of structural compound has to be chosen to avoid by chance of correlation. Activity data was limited at least 4 magnitude order. The selected compounds should have simple and clear information. Any redundancy (extreme) in the structural features or in the activity data must be avoided. Inactive compound due to steric factor is unsuitable to be used in the training set because Catalyst is limited to accommodate that such cases.

### Data Set mining

The number of 117 ligands were collected from some literatures as dataset that subtraction as training set and external set. that has activity against PPAR $\gamma$  (Martres *et al.*, 2008; Henke, 2004a; Koyama *et al.*, 2004; Shibata *et al.*, 2012; Oon Han *et al.*, 2007). The database were chosen based on similar bioassay method as shown in Fig. 5. Activity values were presented as Effective Concentration, or EC<sub>50</sub>.

Dataset was then divided into training set and test set. The Most Active (MA) compound gives an important information for pharmacophore modeling and a few Moderately Active (M) compound as well as Less Active (L) compound are also important in spreading the range for activity value as large as possible. A number of 20 ligands that fulfilled the criteria were used as training set and the remaining 25 compounds were selected as a test set based on the formula of 1.1. and 1.2. Training set was selected using formula (1), while the external set was chosen beside of training set. For the training set data, attribute of

uncertainty was set up to show uncertain value with a score of 3 on the spreadsheet. This means, the activity of the actual ligand in the training set would be in the range of 1/3 to 3 times from the ligand's activity.

Activity range of PPAR $\gamma$  compound is classified as followed: *Most Active* (<0,9nM), *Moderate* (0,9nM-316nM) and *Less Active* (>316nM). Compound's activity is determined by calculation with the formula:

$$MA * UncMA - A / UncA > 0.0 \quad (1.1)$$

$$\log(A) - \log(MA) > 3.5 \quad (1.2)$$

Description: MA = compound with highest activity (lowest EC<sub>50</sub>); UncMA=uncertainty of activity is measured and A= activity from active compound (Abu Hammad and Taha, 2009).

## Pharmacophore Modeling

### Generation of Conformation Library of Bioactive Compounds

For the training and test sets molecules, conformational models representing their available conformational space were calculated. All molecules were built using the 2D and 3D sketcher of Hyperchem 7.0, and optimized using MM2 in Hyperchem 7.0. A conformational set was generated for each molecule using the poling algorithm and the best energy option, based on CHARMM force field from Discovery Studio 2.5 (Brooks *et al.*, 1983; Musfiroh *et al.*, 2013). The molecules associated with their conformational models were mapped onto the pharmacophore model using the "best fit" option to obtain the bioactive conformation of each molecule.

### Pharmacophore Models

A number of 255 Conformers was chosen to be minimized as best conformation, and 20 kcal/mol was set as energy threshold as global energy minimum for conformational searching (Wang *et al.*, 2008), this protocol is available in DS 2.5 packages. The best pharmacophore models was validated according to Deng *et al.* (Deng *et al.*, 2008) in terms of cost functions and other statistical parameters which were calculated by HypoRefine module during hypothesis generation. A good pharmacophore model should have a high correlation coefficient, lowest total cost and RMSD values, and the total cost should be close to the fixed cost and away from the null cost. The best pharmacophore model was further validated by external test set and Fischer's randomization test (Abu Hammad and Taha, 2009).

## Molecular Docking

This study used LBD (ligand binding domain) of peroxisome proliferator-activated receptor gamma or PPAR $\gamma$  (PDB id : 3HOD) (Fracchiolla *et al.*, 2009) structure with 2.1 Å resolution. The crystal structure were selected according to best resolution or lower resolution value, and also have R-free and R-value lower than 0.25 (Mughtaridi *et al.*, 2014). The 3D structures of lignan derivatives compounds were constructed using Hyperchem 7, then were optimized using Austin Model 1 (AM1). Docking results have been saved in a dlg file to be analyzed to get

information about ligand orientation, binding energy value and  $K_i$  value (Ikram *et al.*, 2015). Molecular docking method was validated by re-docking of the ligand of crystal as control docking against PPAR $\gamma$  (PDB id: 3HOD). Autodock 4.0 was employed to know the interaction between ligand and receptor (Muchtaridi *et al.*, 2014).

## RESULTS

### 3D-QSAR Pharmacophore Model

The Hypogen that embedded in DS 2.5. was generated 100 models. The results of the pharmacophore hypothesis from the 3D QSAR Pharmacophore were presented in Table 1. The results showed that hypothesis 1, run 9 (Hypo1) was the best hypothesis results in this study. This is based on the value of cost difference, which is the highest value of 40.9316, lowest root mean square error with value of 0.591049, and highest correlation coefficient with value of 0.972949.

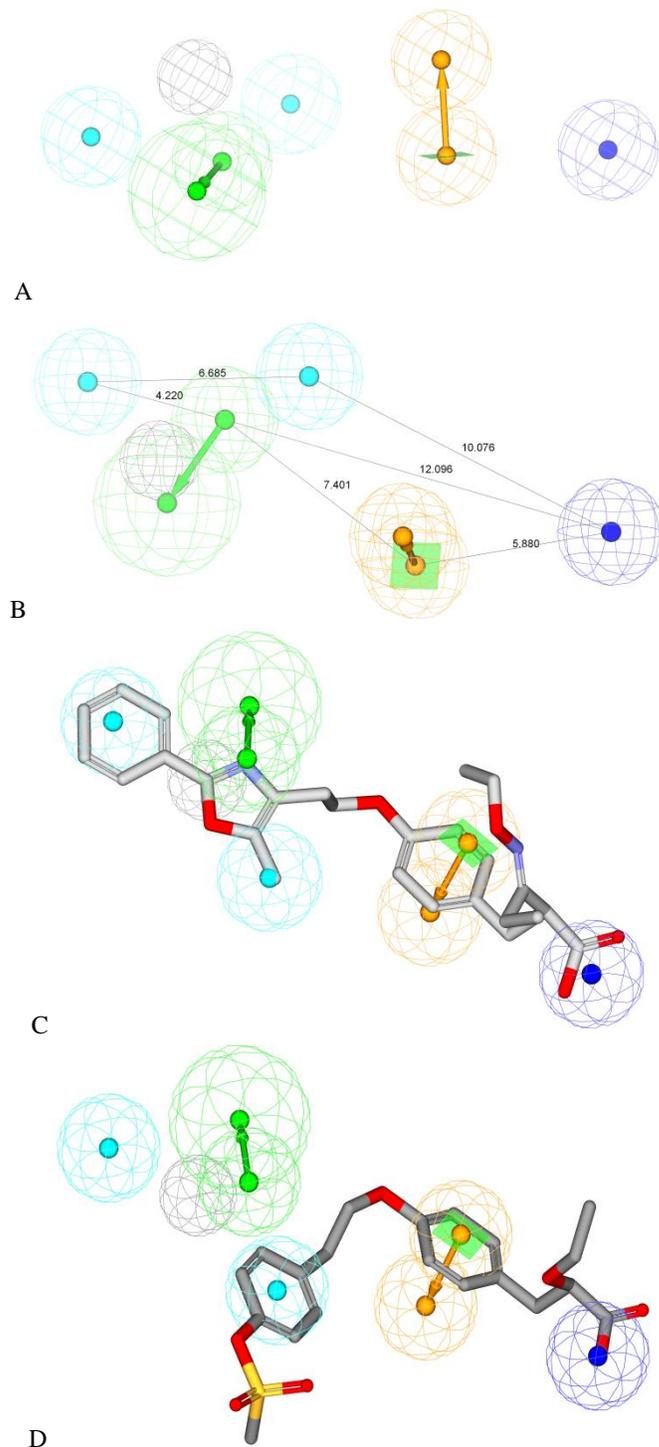
**Table 1:** Hypothesis result of 3D QSAR pharmacophore for training set.

Run no.	Total cost	Cost diff. <sup>1</sup>	RMSD (Å)	Correlation (r)
4	89.4434	40.7476	0.628438	0.96755
5	89.6173	40.5737	0.683979	0.962129
6	93.0029	37.1881	0.788700	0.949402
7	91.2994	38.8916	0.799089	0.947947
<b>9</b>	<b>89.2594</b>	<b>40.9316</b>	<b>0.591049</b>	<b>0.972949</b>

<sup>1</sup>(Null cost-total cost), null cost = 130.191. All cost values are in bits.

This proved that there was more than 97% correlation between estimated activity and actual activity from the training set compounds. Fixed cost, total cost, and null cost are 84.8473, 89.2594 and 130.191, respectively. Ideal difference between null cost and fixed cost are the value in between 70-100 bit. The difference between null cost and fixed cost that lies between 40-60 bits indicates that correlation probability from Hypo1 is 75%-90% (Accelrys, 2002). The bigger difference and the higher probability could identify a good pharmacophore model. Configuration cost value has to be less than 17 and configuration value obtained is 16.4502. (Sutter *et al.*, 2000). The 3D space and distance constraints of these pharmacophore features are shown in Fig. 2B. Pharmacophore features of Hypo1 was presented in Figure 1 (A), consisting of 5 pharmacophore features, employing HBA, NI, RA, two HY and one excluded volume in Figure 1(B), shows the distance between features in Armstrong (Å) units. The distance between features of HY1 and HY2 was 6.6885 Å. HY1 to HBA was 4.220 Å, HBA to NI was 12.096 Å, HBA to RA, 7,401 Å and RA to NI was 5,880 Å. This distance happens to be a combination of electrical and steric features of compounds that is necessary to ensure an optimal interaction of molecules with PPAR $\gamma$  to trigger its biological response (agonist) (Manetti *et al.*, 2003). Figure 1c showed a lot of hydrophobic interactions in the binding pocket PPAR $\gamma$  (hydrophobic character). HBA interacted with the amino acid residue HIS449 (2.4 Å), HIS323 (3.0 Å) and TYR473 (2,8Å). This interaction explained that the ligands should have hydrophobic interactions and hydrogen bonding, to ensure

their biological activity (PPAR $\gamma$  agonist). Mapping results of the training set compound on Hypo1 with the activity (experimental and estimated) and their corresponding error.



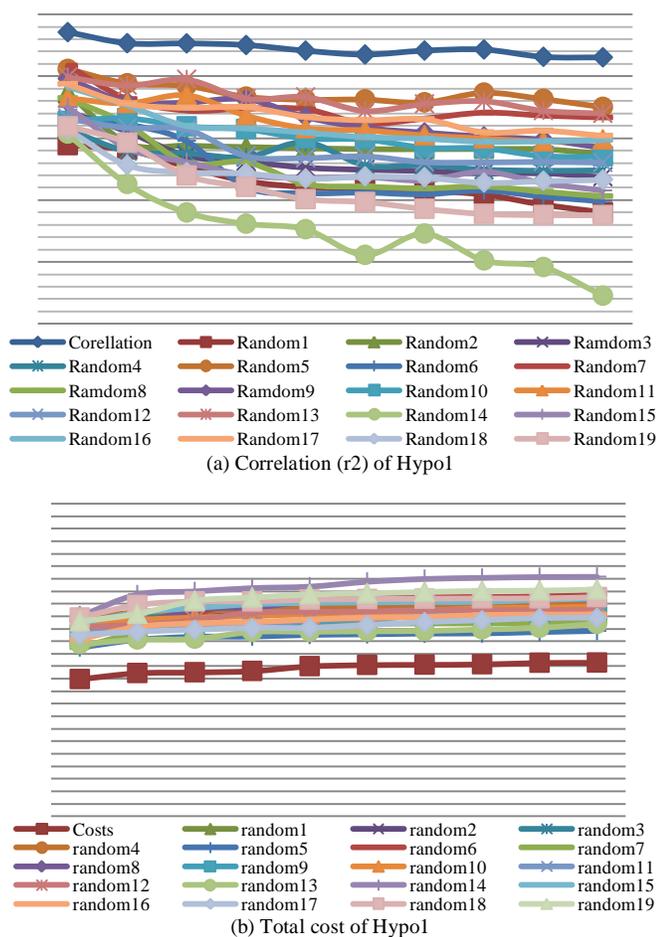
**Fig. 1:** Feature of Best Pharmacophore with Validation by Hyporefine Run in DS 2.5. (a) The best HypoRefine pharmacophore model, Hypo1. (b) 3D spatial relationship and geometric parameters of Hypo1. (c) Mapping of Hypo1 to MA 6 from training set (d) Mapping of Hypo1 to LA 105 from training set. Pharmacophore features are color coded; green: hydrogen-bond acceptor (HBA), cyan – hydrophobic feature (Hy), dark blue – negative ionizable (N), orange – aromatic ring (AR), and grey – excluded volume.

All most active (MA) compounds have been correctly predicted by Hypo1. All MA compounds mapped all pharmacophore features covering HBA, NI, AR and HY. One of MA compounds, i.e. compound 6 with EC50 value of 0.10 nM shows a fit value of 10.85 (Experiment: 0.1nM; Prediction: 0.18nM). Fit value indicates how well the pharmacophore features overlay chemical features in the molecule (Guner, 2005). A higher fit value is better [32]. Figure (3A), showed the compound 6 with HBA feature fitted with nitrogen sp/sp2 hybridization that has free electrons and smaller or no charge. The NI feature fitted against the carboxylate functional group, AR mapped to the benzene and both HY with methyl and phenyl respectively.

## Validation of Pharmacophore Model

### Fischer Randomization test

Fischer Randomization is embedded in the DS 2.5 package to evaluate the best pharmacophore model. CatScramble module within Catalyst was performed to evaluate the models.



**Fig. 2:** The difference in total cost (b) and correlation (a) of hypotheses between the initial spreadsheet and 19 random spreadsheets after CatScramble run.

This statistical program in CatScramble mixes up activity values of all training set compounds to check whether there are strong correlation between the structure and activity.

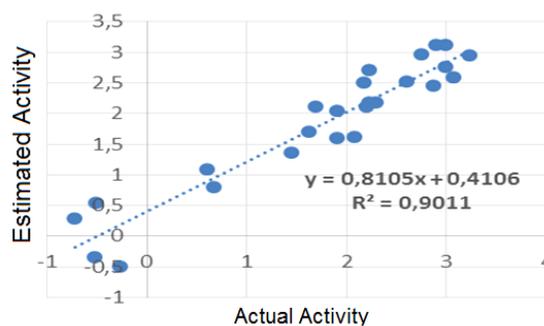
Cross validation test explained that Hypo1 has a significant value of 95% and the results. It could be seen in Figure 2a that the correlation value ( $r$ ) on all pharmacophore models (Random 1 – Random 19) were smaller than the Hypo1 correlation, which showed highest correlation between experimental activity and predictive value. Fischer randomization test was done to assess the predictive quality of Hypo1. COST value indicated that the difference between estimation values and experiment value. COST data obtained from running of 3D QSAR from every hypothesis was created a mold to test for Fischer randomization. Compounds were taken at random from every hypothesis, and then compared with the other hypothesis. Based on the results of Fischer randomization (Figure 2b), it was shown that data cost from training set compound was taken randomly by Hypo1 that showed the lowest COST. Both COST value and correlation showed that Hypo1 was the best hypothesis.

### Test set methods

Subsequently, all the test set molecules were prepared by the same way as that for the training set molecules. Hypo1 was applied to map the 25 test set compounds which gave a correlation coefficient of between experimental and estimated activities as shown in estimated Fig 4.

The results showed that all ligands with MA activity were predicted accurately, except for ligand 6 and 10, where by the ligand is underestimated became M.

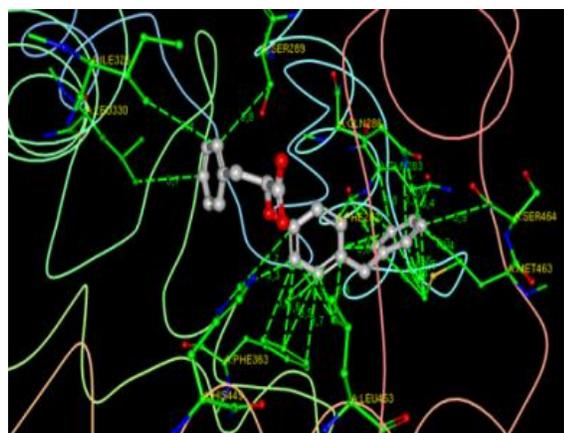
From all the M ligands, only ligand 45 was overestimated to have MA activity. LA ligands were all predicted accurately as well, except for ligand 63, in which it is overestimated into MA. Hypo1 shows a correlation as large as 0.9011, which means that there are good relationship between experimental value and estimated value. A linear regression showing the similarity of experimental activity and estimated activity is shown in Figure 3.



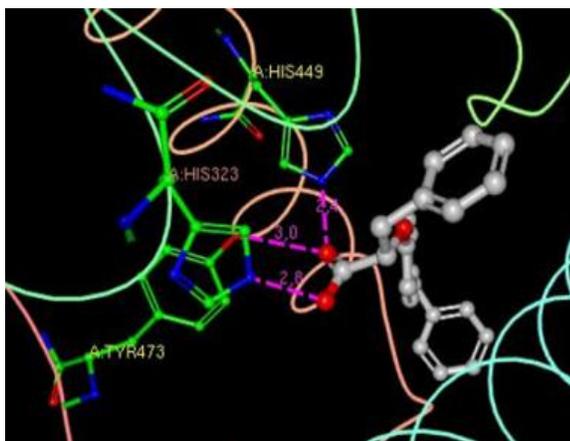
**Fig. 3:** Plot of the correlation ( $r$ ) between the experimental activity and the predicted activity by Hypo1 for the test set molecules and the training set molecules.

Figure 4 showed that the character of the PPAR $\gamma$  binding pocket using software Ligand Explorer incorporated in the Protein Data Bank (PDB) website. Binding mode of the lead compound was shown by the interaction between the amino acid residues with the binding site. Figure 1c shows a lot of hydrophobic interactions in the binding pocket PPAR $\gamma$  (hydrophobic character). Figure 1c and Figure 3 showed that HBA interacted with the amino acid residue

HIS 449 (2.4 Å), HIS 323 (3.0 Å) and TYR 473 (2.8Å). This interaction suggested that the ligands should have hydrophobic interactions and hydrogen bonding, to ensure their biological activity (PPAR $\gamma$  agonist).



A



B

**Fig. 4:** Character of the PPAR $\gamma$  binding pocket using software Ligand Explorer incorporated in the Protein Data Bank (PDB). (a) Hydrophobic interaction in PPAR $\gamma$  binding pocket, (b) Hydrogen bond interaction in PPAR $\gamma$  binding pocket

#### Activity Prediction of Nutmeg Seed Compounds

Dihydro-di-isoeugenol (DHDE) in nutmeg (*Myristica fragrans* Hout) is recently developed as antidiabetic medicine with predicted mechanism of action as a dual agonist, i.e. PPAR  $\alpha$  and  $\gamma$  (Muchtaridi *et al.*, 2014). In this study, 14 compounds derived from nutmeg plant, thiazolidinediones and lead compound 2-arloxy-3-phenyl propanoic acid (3HOD) which is expected active, was mapped by Hypo1 model.

Reason for mapping is to approach that even with different chemotypes, it can have good mapping results with Hypo1. From a total of 22 compounds, Malabaricone B from nutmeg extract shows highest fit value (8.10119) with estimated activity of 100.997 nM. However, this compound still doesn't have any research on its antidiabetic activity towards PPAR $\gamma$  receptor *in vitro* or *in vivo*. Antidiabetic currently being used in Indonesia now is Pioglitazone, with a fit value of 6.87839. Hypo1 predicted

EC50 1686.99nM where experimental activity value of 1280nM (Casimiro-Garcia *et al.*, 2013). Rivoglitazone shows fit value of 7.08281 (Kong *et al.*, 2011; Schimke and Davis, 2007) with estimated activity value of 1053.62 nM

Although Malabaricone B shows highest fit value, it exists in a small quantity in nutmeg seeds which complicates the process being a marker and for extraction. Perhaps in future study, a new lead compound can be found or a better extraction technique for the nutmeg contents.

#### CONCLUSION

Active pharmacophoric features for PPAR gamma agonist are Hydrogen Bond Acceptor (HBA) and Hydrophobic (HY). The compounds of nutmeg seeds (*Myristica fragrans* Hout) have the highest fit value (8.10119) with a total estimated of activity value 100,997 nM.

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