

# Bioadhesive HPMC Gel Containing Gelatin Nanoparticles for Intravaginal Delivery of Tenofovir

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## ABSTRACT

The aim of the present study is to formulate tenofovir loaded gelatin nanoparticles by two step desolvation method for targeted release of drug by varying the concentration of polymer and cross-linking agent. Entrapment efficiency for all the formulations was found to be within  $67.32 \pm 1.24$  % to  $92.11 \pm 1.13$  %. Average particle size of different tenofovir loaded gelatin nanoparticle formulations was found within the range of 294.9 - 445.3 nm. *In-vitro* drug release study for glutaraldehyde cross linked gelatin nanoparticles were found between  $67.09 \pm 1.423$  -  $82.41 \pm 1.874$  after 8 h of dissolution. F5 (850 mg gelatin, 0.2 ml glutaraldehyde) was considered as the best formulation based on the entrapment efficiency and drug release from nanoparticle core. Kinetics study was performed for all the formulations and best fit model for drug release was determined depending on R squared values. HPMC K15M was used as a bioadhesive polymer as well as a gelling agent. Three different gel formulations were prepared by varying concentration of HPMC K15M and incorporated with the best formulation, F5. Membrane permeation and bio-adhesion study revealed F5B gel (5% HPMC K15M) as an optimum formulation with suitable bioadhesive strength and membrane permeability.

## INTRODUCTION

According to WHO and the Joint United Nation Program on HIV/AIDS (UNAIDS), HIV claimed more than 25 million lives worldwide since first recognised in December, 1981(WHO/EURO report.,1996; WHO/EM report., 1998). Since then, AIDS continues to be an opportunistic infection and a major threat to health globally (WHO., 1999; Espinoza *et al.*, 2007). In spite of prevention strategies such as safe sex practice and spreading awareness on the same, the incidence of STDs have not succumbed (Gewirtzman *et al.*, 2011). This scenario potentiates the need to combat the risk of STDs in near future. WHO recommended the class of new emerging anti-retrovirals can be widely used for pre-exposure prophylaxis (PrEP) of AIDS (Padian *et al.*, 2008; Anthony *et al.*, 2015). PrEP is the use of antiretroviral medications by HIV-negative persons to prevent

HIV infection upon high risked HIV exposure. Tenofovir is a nucleotide analogue reverse transcriptase inhibitor which has emerged as potential PrEP drug with a wide safety index. In spite of being an effective drug for children and adolescents (WHO, 2010), tenofovir suffers a range of disadvantages such as GI side effects, low permeability and low oral bioavailability (Peter *et al.*, 2011). Topical delivery of microbicides is a useful alternative for prevention strategies of sexually transmitted diseases like HIV. Avoidance of first pass effect, increased area for drug penetration, less enzymatic activity and rich vascularisation has made vaginal tract an important site for drug administration (Bernkop and Hornof, 2003). As per a current research by U. Parikh *et al.*, pig tail macaque monkeys treated with vaginally applied tenofovir before exposure to simian HIV (SHIV) remained free of infection after 20 challenges ( $p < 0.005$  versus control groups)(Parikh *et al.*, 2009). Furthermore, CAPRISA 004 trial confirmed effectiveness of a 1% vaginal gel of tenofovir for the prevention of HIV infection in women. This double-blind, randomized controlled clinical trial claimed that tenofovir gel could prove to be a promising brick in the HIV prevention gap (Abdool *et al.*, 2010).

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Gelatin nanoparticles (Jahanshahi *et al.*, 2008) have earlier been used for effective delivery of DNA and proteins with a major advantage of its ease of cross-linking, nonpyrogenicity, less cellular toxicity and low antigenicity (Schwick and Heide., 1969). In the present research, tenofovir was incorporated into cross linked, biodegradable gelatin nanoparticles by double desolvation method (Kreuter *et al.*, 2000).

The vaginal gel formulation of tenofovir will be particularly helpful in interfering with the sexual transmission of HIV in women. The mucoadhesive nature of the gel helps to retain the formulation for a longer period of time facilitating sustained release of the drug at site of action (Flynn, 1996), overcoming drug resistance and reduced toxicity (Brigger *et al.*, 2002; Deverdiere *et al.*, 1994).

## MATERIALS AND METHOD

### Materials

Tenofovir was received as a gift sample from Aurobindo Pharma, Hyderabad, India. Gelatin and HPMC K15M was received as a gift sample from Alembic Ltd, Vadodara, India. Glutaraldehyde was procured from Sigma Aldrich Chemicals Pvt. Ltd., Mumbai, India. The chemicals and reagents used were of analytical grades.

### Preparation of tenofovir nanoparticles by two step desolvation method

Gelatin nanoparticles of tenofovir were prepared by two step desolvation method (Weber *et al.*, 2000). Different concentrations of gelatin solution were prepared by dissolving required quantity of gelatin in 20 ml of distilled water in a constant heating temperature. 20 ml of acetone was added to the prepared gelatin solution for precipitating high molecular weight gelatin. The supernatant layer was discarded and high molecular weight gelatin was re-dissolved again by using 20 ml of distilled water. The pH of the gelatin solution was adjusted to 5.6 with the help of 0.1N sodium hydroxide solution.

Tenofovir was dispersed separately with tween 20 and then added drop-wise to the gelatin solution with continuous stirring. Glutaraldehyde was added in various concentrations to study the effect of cross-linking on particle size, drug entrapment and release of the drug from polymeric nanoparticles. Stirring was continued up to 8 h and then the prepared nanoparticles were separated by centrifugation at 2000 rpm. The nanoparticles were collected and washed with distilled water and then dried at 40°C.

### Characterization of tenofovir nanoparticles

#### Estimation of drug entrapment efficiency

Accurately weighed 50 mg of each nanoparticle formulations were taken and placed in 100 ml of acetate buffer solution having pH 4.5 was kept 24 h under constant stirring condition. After 24 h, the solution was filtered through Whatman filter paper and the drug content in the filtrate was determined

spectrophotometrically at 261nm by using UV-VIS spectrophotometer (Elico Lab, SL 210).

### Fourier transformed infrared spectroscopy (FTIR)

Drug- excipient compatibility study was carried out by the FTIR analysis of pure drug (tenofovir), pure polymer (gelatin) and nanoparticle formulation containing tenofovir using FTIR spectrometer (IRAffinity-1S, Shimadzu, Japan). Each sample grounded and mixed with KBr (Uvasol, Merck, KgaA, Germany). Pellets of the resulting mixture were prepared by using hydraulic press with an applied pressure of 100 Kg/cm<sup>2</sup> for 15 min. The pellets were then scanned from 4000 to 400 cm<sup>-1</sup> with a mirror speed of 2 mm/sec.

### Determination of particle size and zeta potential

The prepared tenofovir nanoparticles were dispersed in distilled water and sonicated for 5 minutes. The sonicated suspension was tested for particle size and zeta potential by using a light scattering particle size analyzer (Malvern Instruments Ltd., Sl. No.–MAL 1040985).

### Scanning electron microscopy

Tenofovir loaded gelatin nanoparticles were analyzed for surface morphology by field emission- scanning electron microscope (FE-SEM) (JEOL, JSM-6360, Japan). At first few samples were deposited on stub using one side of a double-sided adhesive dried carbon tape (NEM Tape, Nisshin Em. Co. Ltd. Tokyo, Japan) and then kept under saturated vapour of palladium to make them conductive. It was mounted on SEM instrument and the morphology of the nanoparticles was examined by secondary electro image (SEI) detector.

### In-vitro release study from tenofovir loaded gelatin nanoparticles

*In-vitro* release of tenofovir from prepared gelatin nanoparticles was performed across the membrane of dialysis bag. The drug release study was performed in USP type II dissolution apparatus (Lab India D5 8000) containing 900 ml of acetate buffer solution pH 4.5. The temperature was maintained at 37.4 ± 0.5°C under 50 rpm speed. Accurately weighed quantity of 50 mg nanoparticle formulation of tenofovir placed in dialysis bag (Cellophane membrane, Hi-media, India) containing 3 ml of acetate buffer solution. Then the dialysis bag was tied on both ends and placed in dissolution apparatus. The dialysis bag acted as a donor compartment, while the dissolution vessel acted as receptor compartment. 5 ml of aliquots was collected at regular intervals and replaced with fresh buffer solution to maintain sink condition throughout the experiment. The aliquots were filtered and diluted appropriately before determining the absorbance by using a UV-VIS spectrophotometer (Elico Lab, SL 210) at  $\lambda_{max}$  261 nm.

### Release kinetics

The drug release pattern from prepared gelatin nanoparticles of tenofovir were studied for various kinetic models like zero order, first order, Higuchi and Korsmeyer- Peppas model.

Zero order kinetics:  $Q_t - Q_0 = K_0t$ ; Where,  $Q_t$  is the amount of drug released after time  $t$ ,  $Q_0$  is the initial concentration of the drug in solution;  $K_0$  is the zero order rate constant (Hadjioannou *et al.*, 1993).

First order kinetics:  $\log Q_t = \log Q_0 - \frac{K_1t}{2.303}$ ; Where,  $Q_t$  is the amount of drug released after time  $t$ ,  $Q_0$  is the initial concentration of the drug in solution;  $K_1$  is the first order rate constant (Bourne., 2002).

Higuchi model:  $Q = Kt^{0.5}$ ; Where,  $Q$  is the amount of drug released after time  $t$ ; and  $K$  is the rate constant (Higuchi., 1963).

Korsmeyer-Peppas model:  $Q = Kt^n$ ; Where,  $Q$  is the amount of drug released after time  $t$ ,  $K$  is the rate constant and  $n$  is diffusional exponent for drug release.

Korsmeyer-Peppas model can also distinguish the exact release mechanism of drug. Depending on the  $n$  value obtained from the experiment, the data were analyzed for various release mechanisms: Fickian release (diffusion controlled release), non-Fickian release (anomalous transport) and case II transport (relaxation controlled release). In this context,  $n \leq 0.43$  indicates Fickian release,  $0.43 < n < 0.85$  indicates non-Fickian release, and  $n \geq 0.85$  symbolize a case II transport (Ritger and Peppas., 2002).

#### Preparation of HPMC K15M gel containing 1% tenofovir

Intravaginal nanogel of tenofovir was prepared by using HPMC K15M as a gel forming agent. Three different gel formulations F5A, F5B, F5C were prepared by dissolving different concentrations of HPMC K15M in 30 ml of acetate buffer (pH 4.0) with continuous stirring by magnetic stirrer (Remi motors, India) at 300 rpm to get a uniform clear dispersion. The resultant gels were kept undisturbed overnight for complete swelling (Ahmad *et al.*, 2008). Then accurately weighed quantity of tenofovir nanoparticles equivalent to 100 mg of tenofovir was mixed homogeneously with the HPMC K15M solution. Finally 1% tenofovir nanogel was prepared by using glycerine as humectant, citric acid as penetration enhancer and triethanolamine as pH modifier.

#### Characterization of HPMC K15M gel containing tenofovir loaded gelatin nanoparticles

##### Determination of pH

The pH of the prepared nanogel formulations were determined by using a digital pH meter (Elico Lab, India) by dipping the glass electrode in to the gel system completely.

##### Measurement of viscosity

The viscosity of the prepared 1% tenofovir gel was determined by using Brookfield viscometer DV-III Ultra (Middleboro, USA) at  $25 \pm 0.5^\circ\text{C}$ . Spindle number 6 was used to measure the viscosity at 10 rpm.

##### Determination of spreadability

Spreadability of the prepared gel was determined by placing 1 gm of gel in the centre of a glass plate which was

covered with another glass plate of same size ( $20'' \times 20''$ ). 1000 g of weight was carefully placed on the upper glass plate for expelling air and uniform spreading of gel. After 1 min, weight was removed and the spreading area of the gel between the glass plates was measured (Lardy *et al.*, 2000; Garg *et al.*, 2002).

#### Determination of bioadhesive strength

Bioadhesive property was determined using modified physical balance method (Harish *et al.*, 2009). Chick ileum mucosa was used as biological membrane, which was fixed under one pan of the balance with the help of glue and was hydrated with acetate buffer pH 4.5 maintained at  $37 \pm 0.5^\circ\text{C}$ . Accurately weighed amount of 1 g of gel was stuck to the inverted beaker (250 ml) using glue and the height of the balance was adjusted to accommodate a glass container below the pan where membrane was glued. A preload of 20 g was applied in order to allow the formation of mucoadhesive joints. After a 3 min rest period, the preload was removed and gradually the weight was added to the other pan until the gel was detached from the mucosal surface (Ellaithy and El-Shaboury, 2002). The total weight required for the complete detachment of the gel was recorded. The mucoadhesive force expressed as the detachment stress in  $\text{dynes/cm}^2$  was determined from the minimal weight that detached the mucosal tissue from surface of each formulation.

$$\text{Mucoadhesive Strength} = \frac{mg}{A}$$

Where,  $m$  = Weight required for detachment in gm,  
 $g$  = Acceleration due to gravity ( $981 \text{ cm/s}^2$ ),  
 $A$  = Area of mucosa exposed.

#### Membrane permeation study

The membrane permeation study for the prepared tenofovir gels were carried out to measure the cumulative amount of drug permeating through a biological membrane. Egg membrane was used as a biological membrane in this study.

##### Preparation of the membrane

The egg was kept in concentrated hydrochloric acid to dissolve the shell. Then by puncturing the membrane from one end, the contents were taken out. The membrane was then cleaned by using double distilled water. After thoroughly cleaning of membrane, it was kept in a petri dish containing acetate buffer pH 4.5 for overnight soaking.

##### Permeation study by Franz diffusion cell

The membrane permeation study was performed by using Franz diffusion cell which consist of two chambers- the donor compartment and receptor compartment. The egg membrane was mounted between the compartments by facing the outer side of the membrane towards the donor compartment. Acetate buffer pH 4.5 was used as a receptor medium. The total set up was done over a magnetic stirrer with a maintained temperature of  $37 \pm 0.5^\circ\text{C}$ . The

donor compartment was filled with 1 gm of nanogel formulation. At appropriate intervals, 2 ml of receptor medium was withdrawn and replaced with equal volume of fresh medium. The amount of permeated drug through the membrane was analyzed by using a UV-VIS spectrophotometer (Elico Lab, SL 210) at 261 nm wavelength in reference with an appropriate blank buffer solution (Narendra *et al.*, 2011).

#### Accelerated stability study of prepared gels

Accelerated stability test (Sanjay *et al.*, 2012) was carried out for all the prepared gels at accelerated conditions of temperature and humidity ( $40 \pm 2^\circ\text{C}$  and  $75 \pm 5\%$  RH) for a period of 90 days. Samples were withdrawn at regular time interval (1, 2 and 3 month) and analyzed for appearance, pH, drug content and compared with zero time samples as controls.

## RESULTS AND DISCUSSION

#### Drug entrapment efficiency

The entrapment efficiency of tenofovir loaded gelatin nanoparticle formulations were found to be within the range of  $67.32 \pm 1.24$  to  $92.11 \pm 1.13$  as shown in Table 3. F-5 was found to have the highest entrapment efficiency where as F-1 contains lowest amount of drug. From the drug entrapment result, we can also conclude that the increase in glutaraldehyde concentration is

directly proportional to drug entrapment efficiency. This may attribute due to high degree of cross-linking brought about by glutaraldehyde.

#### FTIR spectroscopy

FTIR spectrum of tenofovir, gelatin and glutaraldehyde crosslinked nanoparticle formulation (F5) was shown in Fig. 1. Hydrogen bond formation is observed in formulation, which was detected by broadening of peak at 3200 to 3500  $\text{cm}^{-1}$ . FTIR spectrum of tenofovir shows peak at 1762.94  $\text{cm}^{-1}$  for presence of carbonyl group (C=O) which can be observed in formulation (1734.01  $\text{cm}^{-1}$ ). Peaks for C-C stretching for aromatic ring, C-N stretching for aromatic amines, C-N stretching for aliphatic amines, C-H bending for alkenes, C-Cl stretching for alkyl halides were observed at 1409.96  $\text{cm}^{-1}$ , 1300.02  $\text{cm}^{-1}$ , 1246.02  $\text{cm}^{-1}$ , 952  $\text{cm}^{-1}$  and 746.45  $\text{cm}^{-1}$  respectively. The FTIR spectrum of tenofovir loaded nanoparticles showed peaks at 1421.54  $\text{cm}^{-1}$ , 1269.16  $\text{cm}^{-1}$ , 1186.22  $\text{cm}^{-1}$ , 952.84  $\text{cm}^{-1}$  and 788.89  $\text{cm}^{-1}$  for C-C stretching for aromatic ring, C-N stretching for aromatic amines, C-N stretching for aliphatic amines, C-H bending for alkenes, C-Cl stretching for alkyl halides respectively. The characteristic peaks appeared in the FTIR spectrum of tenofovir was also appeared in the spectrum of tenofovir loaded gelatin nanoparticles without any significant shifting of peaks, indicating absence of any chemical interaction during and after preparation.

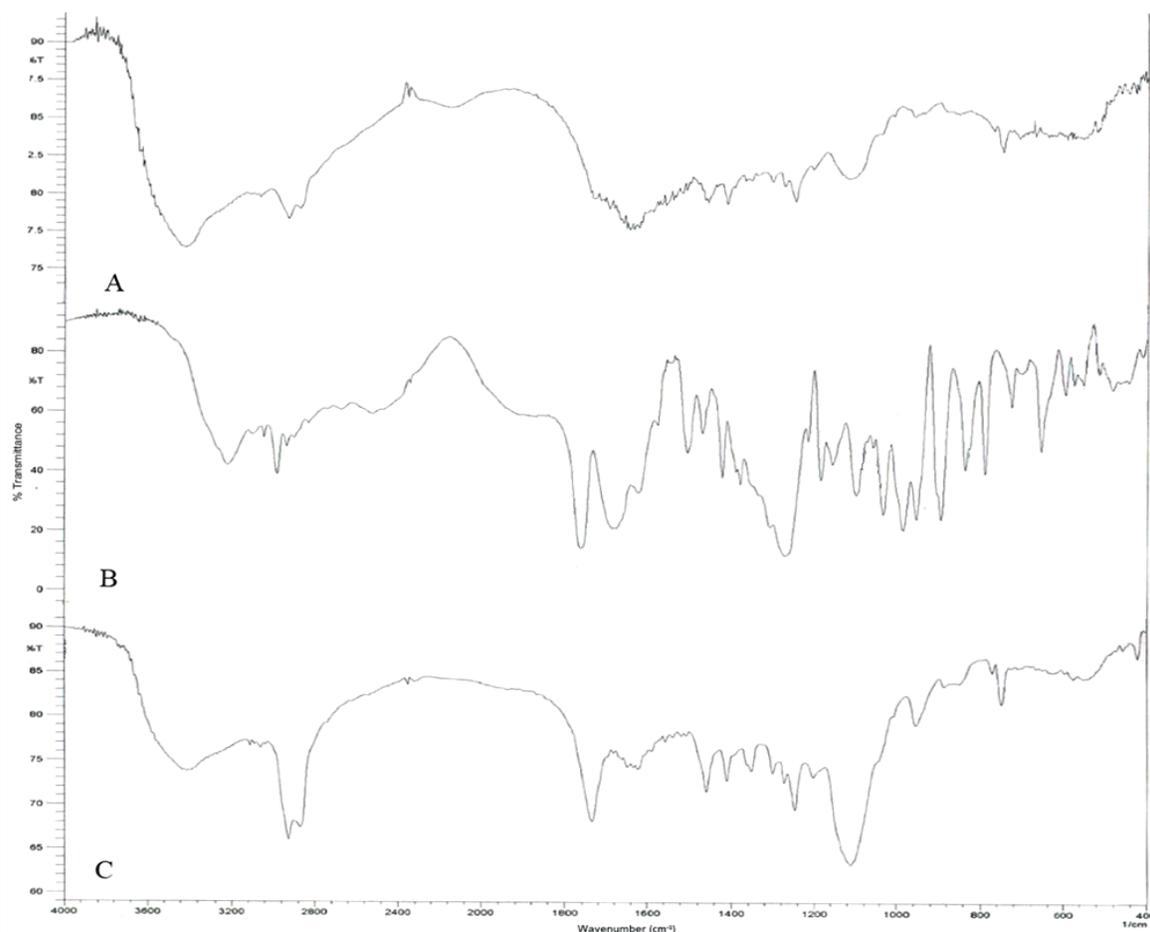
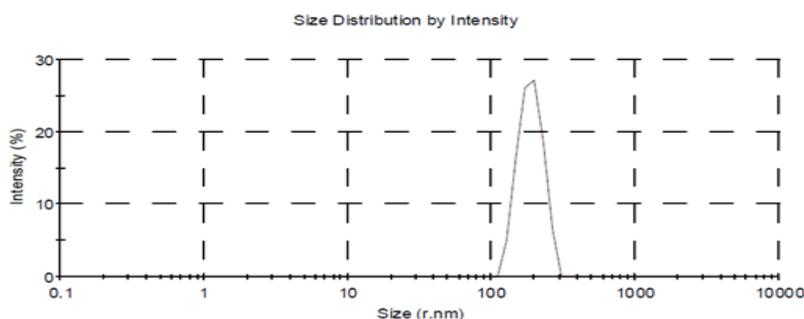


Fig. 1: FTIR spectrum of – A- pure tenofovir; B- Pure gelatin; C- tenofovir loaded nanoparticle formulation (F5)

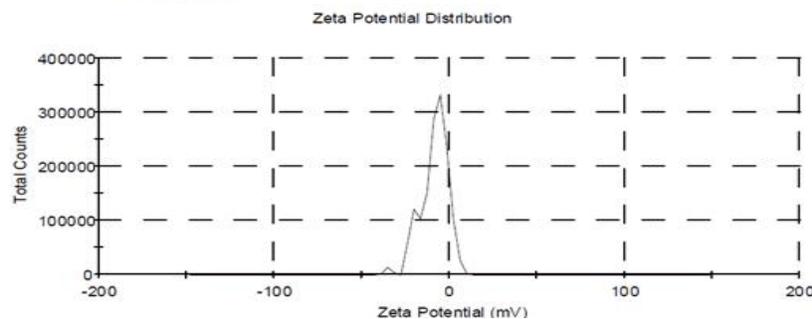
**Table 1:** Composition of various gelatin nanoparticle formulations.

Formulation code	Composition						Average particle size (nm)
	Tenofovir (mg)	Gelatin (mg)	Glutaraldehyde (ml)	Tween 20 (ml)	Acetone (ml)	Distilled water (ml)	
F1	100	500	–	2.0	20	20	445.3
F2	100	600	0.1	2.0	20	20	326.2
F3	100	700	0.1	3.0	20	20	337.8
F4	100	800	0.1	3.0	20	20	380.6
F5	100	850	0.2	2.5	20	20	294.9
F6	100	850	0.3	2.5	20	20	308.2

	Size (r.nm):	% Intensity	Width (r.nm):
<b>Z-Average (r.nm): 294.9</b>	<b>Peak 1: 189.5</b>	<b>100.0</b>	<b>35.71</b>
<b>Pdi: 0.584</b>	<b>Peak 2: 0.000</b>	<b>0.0</b>	<b>0.000</b>
<b>Intercept: 0.999</b>	<b>Peak 3: 0.000</b>	<b>0.0</b>	<b>0.000</b>
<b>Result quality : Refer to quality report</b>			

**Fig. 2:** Particle size distribution of tenofovir loaded gelatin nanoparticle formulation, F5.**Results**

	Mean (mV)	Area (%)	Width (mV)
<b>Zeta Potential (mV): -8.56</b>	<b>Peak 1: -6.27</b>	<b>80.3</b>	<b>5.49</b>
<b>Zeta Deviation (mV): 7.82</b>	<b>Peak 2: -19.6</b>	<b>18.3</b>	<b>2.77</b>
<b>Conductivity (mS/cm): 1.62</b>	<b>Peak 3: -34.7</b>	<b>1.3</b>	<b>2.05</b>
<b>Result quality : Good</b>			

**Fig. 3:** Zeta potential of tenofovir loaded nanoparticle formulation, F5.**Particle size and zeta potential**

Average particle size and particle size distribution of different tenofovir loaded gelatin nanoparticle formulations were found to be 294.9 - 445.3 nm as shown in Table 1. Among all the formulations, F5 was found to have smallest particle size of 294.9 nm and the particle size distribution was shown in Fig. 2. The average particle diameter was found to be lesser with increase in glutaraldehyde concentration. It is also observed that there was an increase in average particle diameter at higher concentration of

gelatin which could be due to variation in degree of cross-linking. The higher concentration of cross-linker (glutaraldehyde) provides higher degree of cross-linking which results the decrease in average diameter of nanoparticles (Jana *et al.*, 2013). The zeta potential of F5 measured by Malvern zeta sizer was found to be -8.56 mV with a zeta deviation of 7.82 mV as shown in Fig. 3. The data indicates good stability of nanoparticle formulation in a colloidal dispersion due to electrical repulsion of the charged particles.

### Surface morphology analysis

The morphological analysis of prepared tenofovir nanoparticles were visualised by scanning electron microscope. Fig. 4 displays the SEM image of F5 formulation. The particles were found to be hemispherical in shape with smooth surfaces. No wrinkles or pores been observed on the surface.

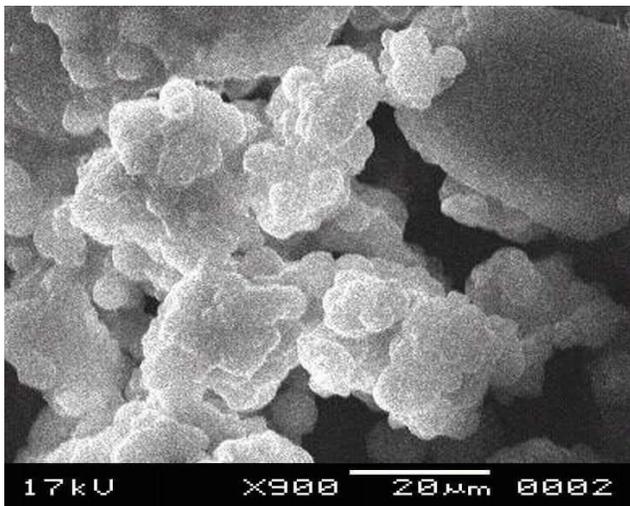


Fig. 4: Scanning electron micrograph of tenofovir loaded nanoparticles, F5.

Table 2: Composition of intra-vaginal nanogel of tenofovir.

Ingredients	F5A	F5B	F5C
Tenofovir loaded nanoparticles equivalent to tenofovir	1 %	1 %	1 %
HPMC K15M	4 %	5 %	6 %
Propyl paraben	0.004gm	0.004gm	0.004gm
Methyl paraben	0.072gm	0.072gm	0.072gm
Triethanolamine	0.02gm	0.02gm	0.02gm
Glycerine	2 ml	2 ml	2 ml
Citric acid USP	0.4gm	0.4gm	0.4gm
Purified water USP	23.2 ml	22.8 ml	22.2 ml

### In-vitro drug release from tenofovir loaded gelatin nanoparticles

*In-vitro* release of tenofovir from the tenofovir loaded gelatin nanoparticles which are ionically cross-linked by glutaraldehyde was examined. The cumulative percentage release was calculated for all the formulation and was found to be sustained up to 8 h. An initial burst release was observed for all the formulation which may be caused due to the drug present on the surface of the nanoparticles.

The cumulative percentage of drug released from glutaraldehyde cross-linked gelatin nanoparticles were found in between  $69.45 \pm 1.171 - 84.09 \pm 1.423$  as shown in Fig. 5.

After 8 h of time F-1 showed the highest amount of drug release which may happen because of loosely bound nanoparticle matrix (Sahana *et al.*, 2010) due to absence of cross-linker. It is observed that the release of the drug from nanoparticle matrix is slower with increase in glutaraldehyde concentration which indicates the formation of firmly bound matrix with gradual increase in cross-linker concentration.

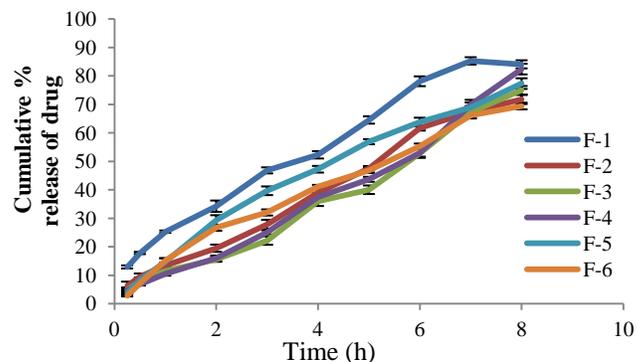


Fig. 5: *In-vitro* drug release from tenofovir loaded gelatin nanoparticles (mean  $\pm$  S. D.; n = 3).

### Release kinetics

*In-vitro* drug release study for prepared tenofovir nanoparticles were kinetically evaluated by using different mathematical models like zero order, first order, Higuchi and Korsmeyer- Peppas model and the results are shown in Table 3. Based on the accuracy of these mathematical models, the correlation coefficient ( $R^2$ ) values were determined for each formulation. When the respective  $R^2$  values of tenofovir loaded gelatin nanoparticles are compared, it is been observed that F1 is following Korsmeyer- Peppas model of Fickian release, F2, F3, F4 are following zero order kinetics and F5 and F6 are following Higuchi model.

### pH of the prepared nanogel formulations

In development of vaginal gel formulation, pH of the formulation plays an important role. Highly acidic or basic pH of vaginal gel formulation can alter the pH of surrounding environment, which may produce skin irritation on application of the gel. The pH of the prepared HPMC K15M gel containing tenofovir loaded gelatin nanoparticles were found to be 4.42, 4.45 and 4.47 for F5A, F5B and F5C respectively which are within the range of normal vaginal pH.

### Viscosity

The viscosity of prepared HPMC K15M gels containing tenofovir loaded gelatin nanoparticles were determined by using Brookfield viscometer. Formulation F5C ( $53454 \pm 22$  cps) was found more viscous in compare to formulation F5A ( $32398 \pm 19$  cps) and F5B ( $37398 \pm 15$  cps) due to increase in HPMC K15M concentration.

### Spreadability

The result for spreadability study of the prepared gel formulations was found to be satisfactory as shown in Table no 4. The high viscosity and bioadhesive property of the HPMC K15M could be responsible for decreasing the spreadability. From the data obtained, it can be clearly said that an increase in HPMC concentration is resulting in less spreadability of the prepared gel formulation.

**Table 3:** Results of curve fitting of *in-vitro* release profile of tenofovir nanoparticles.

Sl. no	Formulation	Zero order R <sup>2</sup>	First order R <sup>2</sup>	Higuchi R <sup>2</sup>	Koresmeyer–Peppas		Best fit model	Drug entrapment efficiency in % (mean ± S.D.; n = 3)
					R <sup>2</sup>	n		
1	F1	0.874	0.958	0.977	0.988	0.418	Koresmeyer-Peppas	67.32 ± 1.24
2	F2	0.982	0.971	0.952	0.978	0.113	Zero order	72.73 ± 0.87
3	F3	0.981	0.920	0.910	0.963	0.282	Zero order	80.24 ± 1.34
4	F4	0.982	0.882	0.912	0.972	0.389	Zero order	91.47 ± 1.08
5	F5	0.980	0.993	0.996	0.992	0.363	Higuchi	92.11 ± 1.13
6	F6	0.985	0.984	0.987	0.984	0.468	Higuchi	84.84 ± 0.86

**Table 4:** Results of various evaluation parameters of tenofovir nanogel.

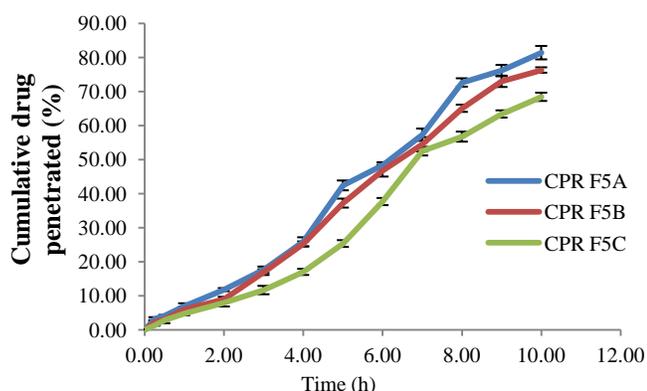
Formulation code	pH	Viscosity (mean ± S.D.; n = 3)	Spreadability (mean ± S.D.; n = 3)	Bioadhesive strength (mean ± S.D.; n = 3)	Appearance (after 3 months)	Drug content after three months (mean ± S.D.; n = 3)
F5A	4.34	32398 ± 19 cps	7.72 ± 0.16	5534 ± 0.11	No change	98.72% ± 0.73
F5B	4.38	37398 ± 15 cps	7.37 ± 0.13	5643 ± 0.13	No change	98.64% ± 0.35
F5C	4.40	53454 ± 22 cps	6.86 ± 0.08	5672 ± 0.14	No change	98.83% ± 0.28

### Bioadhesive strength

Bioadhesive strength for all the prepared tenofovir nanogel formulation was carried out by modified physical balance method by using chick ileum mucosa as biological membrane. The bioadhesive strength of nanogel formulation was found within 5534 ± 0.11, 5643 ± 0.13 and 5672 ± 0.14 dynes/cm<sup>2</sup> for F5A, F5B and F5C respectively as shown in Table 4. According to the requirement of the study, good bioadhesive strength was desired for better adherence and longer duration of action (Chatterjee *et al.*, 2011). The above result indicated that the bioadhesive strength increased with proportional increase in HPMC K15M concentration in nanogel formulation.

### Membrane permeation study

The HPMC K15M gels of tenofovir loaded gelatin nanoparticle formulation were evaluated for membrane permeation study by using egg membrane as a biological barrier. Fig. 6 displays controlled drug permeation from all the formulation was sustained over 10 h.

**Fig. 6:** Comparative drug permeation from HPMC K15M gel containing tenofovir loaded gelatin nanoparticles through egg membrane (mean ± S. D.; n = 3)

The penetration of tenofovir was significantly higher for F5A (81.36 ± 1.97) gel in compare with F5B (76.27 ± 0.79) and F5C (68.40 ± 1.20) gel formulation after 10 h of study.

### Accelerated stability study

All the gel formulation were visually inspected in every week of first month and then at 1, 2 and 3 month respectively. The clarity, transparency, and physical stability were compared with time zero gel formulation of each type. No significant changes were found in appearance of the gel formulations after 3 months. The pH of the prepared formulations F5A, F5B and F5C after 3 months was found to be 4.27, 4.28 and 4.36 respectively. After 3 months of exposure the drug content for F5A, F5B and F5C were resulted as 98.72% ± 0.73, 98.64% ± 0.35 and 98.83% ± 0.28 respectively as shown in Table 4. The above data confirms that there are no significant changes in formulation after storing at ambient condition for 3 months and indicates a stable formulation.

### CONCLUSION

Glutaraldehyde crosslinked gelatin nanoparticles were prepared successfully by two step desolvation method for sustained release of tenofovir. F5 was selected as best formulation based on the particle size, particle size distribution, drug entrapment efficiency and release pattern from polymeric matrix. The drug entrapment efficiency was found to be within the range of 67.32 ± 1.24 to 92.11 ± 1.13. The nanoparticles were found to be hemispherical in nature with an average particle size of 294.9 - 445.3 nm. Zeta potential for F5 (850 mg gelatin, 0.2 ml glutaraldehyde) was found -8.56 mv which assures a stable colloidal dispersion of nanoparticles. The *in-vitro* dissolution study for nanoparticle formulation showed a sustained release of tenofovir over 8 h. 4% (F5A), 5% (F5B) and 6% (F5C) HPMC K15M gel was prepared by taking 1% equivalent weight of tenofovir from nanoparticle formulation F5. Bioadhesive strength analysis was carried out by modified physical balance method using chick ileum.

The results showed higher bioadhesion for F5C (6% HPMC K15M gel). Membrane permeation study was carried out by using egg membrane which reveals a higher permeation rate for F5A (4% HPMC K15M) gel. By consolidating controlled drug permeation, viscosity, spreadability and bioadhesive strength, F5B

(5% HPMC K15M gel) was selected as optimum formulation with a standard bioadhesion and drug permeation profile.

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